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1	Envirotyping to control genotype x environment interactions for efficient
2	soybean breeding
3	
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9	
10	Highlights
11	• Northward soybean deployment requires environmental characterisation using eco-climatic
12	factors
13	• Envirotyping explained 88% of the genotype by environment interactions
14	• Five environment types contrasted by the intensity and timing of stresses
15	• Combine specific and broad adaptation is best suited in the target population of environments
16	
17	Abstract
18	In the context of the European protein deficit and the need for climate change mitigation by agriculture,
19	soybean (Glycine max L. Merr) is of major interest to farmers and breeders. Expanding the crop to new
20	cultivation areas requires understanding and control of the Genotype by Environment Interactions (GEI) that
21	impede the genetic gain. New envirotyping methods, including the Target Population of Environments (TPE)
22	characterisation, are key to the development of efficient breeding programs for specific or broad adaptations.
23	The objectives were to (i) determine the environment types describing the European early soybean TPE, (ii)
24	characterise the distribution and repeatability of the environment types across the TPE to identify the best
25	suited adaptation strategy and (iii) demonstrate the importance of assessing the alignment between multi-
26	environmental trials and the TPE for breeding decisions. In this study, 602 environments from France to
27	Russia were clustered into five environment types using twelve eco-climatic factors, <i>i.e.</i> environmental

28 variables that were calculated over specific phenological periods, such as the number of days below 15°C 29 between flower induction and first flower stages or the solar radiation quantity between first pod and first 30 seed stages. These factors were previously identified as main drivers of GEI for yield in early maturity soybean (maturity groups '000' and '00'). The environmental clustering explained 88% of the GEI effect on 31 32 soybean yield. The five environment types that composed the TPE, mainly contrasted in the intensity and 33 timing of stresses related to temperature (cold stress during the vegetative growth and heat stress during the 34 reproductive growth) and water availability (precipitation amount, evapotranspiration and drought throughout the crop cycle). Interestingly, we observed geographical and temporal variations in the 35 environment types distributions across the TPE as well as in their repeatability. These variations attested to 36 37 the TPE heterogeneity and thus suggested that selection strategies based on either specific or broad 38 adaptations should be combined. For example, specific adaptations to the third and fourth environment types 39 were best suited in Eastern Europe while the broad adaptation to all environment types could be 40 recommended in Western Europe. When broad adaptation was required, we demonstrated the need to assess the alignments between the environment types frequencies in the TPE and those observed when multi-41 42 environmental trials were conducted. This work will contribute to improving the existing soybean germplasm 43 by considering the risks linked with weather variations and unpredictability so as to design elite soybean 44 according to environment type.

45

46 Key words

47 Envirotyping, Multi-environment trial, Target population of environments, Soybean, Weighted selection

48 **1.** Introduction

49 Breeders aim to create cultivars that outperform the existing germplasm under a large set of growing 50 conditions across the cultivation area. Spatial considerations and year-to-year variations in weather scenarios can considerably affect the relative performances of cultivars and make breeding decisions more complex 51 52 (Cooper & DeLacy, 1994). This phenomenon, called Genotype by Environment Interactions (GEI), limits 53 the genetic gain by decreasing the effective heritability of traits, *i.e.* the ability to identify superior genotypes 54 statistically (Brennan et al., 1981; Basford & Cooper, 1998). When the GEI are low, cultivars can be identified that perform well across a wide range of conditions; this is referred to as broad adaptation (Cooper 55 56 & Byth, 1996). Alternatively, when there are greater GEI, certain cultivars can perform better than others 57 within a restricted set of conditions. This is referred to as specific adaptation (Cooper & Byth, 1996). Crop 58 growth models have become increasingly popular to unravel the complexity of the ecophysiological 59 mechanisms and processes underlying GEI (Millet et al., 2019; Rincent et al., 2019; Casadebaig et al., 2021; 60 Bustos-Korts et al., 2022). These models offer the possibility of connecting plant physiological processes 61 with environmental variables (Chenu et al., 2017). For instance, they can be used to calculate climatic 62 variables over specific simulated phenological periods. Known as environmental covariates or Eco-climatic Factors (EFs), they can be treated as explanatory variables of complex traits, including yield (Schoving et 63 al., 2022, Elmerich et al., 2023). 64

65 To capture the GEI effect, breeders traditionally evaluate genotypes across Multi-Environment Trials 66 (METs) that include a set of locations and years, referred to as environments. Such trials aim to characterise 67 the environmental conditions in which the crops are likely to be grown. These conditions correspond to the 68 Target Population of Environments (TPE) (Podlich & Cooper, 1998). Due to seasonal variability and the 69 practical constraints that restrict the number of genotypes and environments tested in METs the environments 70 sampled in METs often offer a biased representations of the TPE (Podlich et al., 1999). An extension of the 71 breeder's equation used to quantify genetic gain was recently developed to explicitly account for the 72 influence of the MET-TPE alignment on trait predictions (Cooper et al., 2023). Thus, weighted analyses 73 based on representativity assessments of trials can enable breeders to correct for this bias and select 74 germplasm that is better adapted to a TPE. This requires an upstream characterisation and understanding of 75 the TPE through identification of the key environmental classes affecting the performance of the genotypes.

76 These environmental classes, referred to as environment types, regroup environments (location x year) in 77 which genotypes have similar performances (Chenu et al., 2013). The classification can be performed 78 according to geographical regions (Döttinger et al., 2023), stress factors patterns (Chapman et al., 2000) or 79 effects on performance traits such as yield levels or maturity dates (Chauhan et al., 2013; Kurasch et al., 80 2017). While useful to describe environmental variables affecting crop productivity, these classification 81 criteria do not attest to the performance stability of the genotypes. Consequently, the best approach for 82 breeding purposes is grouping or clustering environments based on environment variables that are key drivers 83 of GEI (Annicchiarico, 2021). The challenge is to identify these drivers and use them to cluster environments 84 into environment types that define the TPE. This approach minimises GEI within environment types but 85 maximises them between environment types (Chenu et al., 2011; Bustos-Korts et al., 2022).

86 Leguminous crop deployments have been accelerated to address European protein deficits (Marraccini et 87 al., 2020; Ayerdi Gotor & Marraccini, 2022). Among legumes, soybean (Glycine max L. (Merrill)) seems to 88 be a good candidate to become a major oleo-protein crop for both food and feed (Sudarić, 2020). European 89 soybean production is currently low, and importations greatly exceed exportations (FAOSTAT, 2021). 90 Although recent studies demonstrate the potential of soybean crop in Europe, few breeding efforts have been 91 made to adapt soybean in Northern Europe (Boulch et al. 2021; Karges et al., 2022). Under highly divergent 92 weather conditions in France, Elmerich et al. (2023) identified the main EFs driving GEI in semi-93 indeterminate soybeans from two early maturity groups ('000' and '00'). Various GEI-drivers were identified 94 depending on maturity groups and the nature of the GEI (genotypes by location or locations by year). The 95 GEI-drivers occurred mainly during the overlapping period of vegetative and reproductive growths. Cold stress during the vegetative growth, solar radiation intensity during the pod emission and maximum 96 97 temperature before maturity were the main climatic variables that impacted GEI.

The potential early soybean cultivation area is wide in Northern Europe as it extends from France to Russia (approximately between the 45° and the 50° parallels). However, areas dedicated to soybean production remain low (3.6% of the total crops harvested in 2021) (FAOSTAT, 2021). Compared with other regions of the globe, Europe presents a substantial diversity of climatic scenarios (Woollings, 2010). Thus, GEI for yield are expected to be large, which will reduce the heritability of yield-related traits (Holland et al., 2003). 104 To date, the spatial and temporal characterisation of the European soybean TPE is unknown, especially 105 when GEI-drivers are used. Attempts to show the potential of dividing the TPE for specific adaptation have 106 not considered GEI per se nor the environmental characterisation (Kurasch et al., 2017; Döttinger et al., 107 2023). Those advances would enable breeders to position their cultivars on the market and contribute to 108 designing better breeding strategies. The objectives will be to (i) determine the environment types that 109 describe the European early soybean TPE using GEI-drivers, (ii) characterise the distribution and 110 repeatability of environment types across the TPE to propose the best-suited adaptation strategy, and (iii) 111 demonstrate the importance of MET-TPE alignment in breeding decisions and cultivars recommendations.

112

113 **2.** Material and Methods

114 2.1. Datasets

115 Two datasets were used in this study; they differed in data sources and use purposes. Tested and virtual 116 environments (a combination of location x year) were used in order to establish an adequate number of 117 environments. Dataset A consisted of 112 environments in an unbalanced combination of 60 locations and 118 five years (2017-2021). Terres Inovia, the French technical institute for oil and protein crops, used Multi-119 environment trials (METs) to test post-inscription cultivars in those environments (https://www.myvar.fr/resultats/campagne-177.html). The trials were conducted with rigorous chemical and 120 121 mechanical weed management. Even though biotic stresses are limited for soybean in Northern Europe, 122 annual trials are evaluated by a committee and then validated or rejected (and thus were not used in this 123 study). A total of 57 cultivars from two early maturity groups ('000' and '00') were tested (supplementary 124 table S1). The METs for the two maturity groups were conducted independently in France, between the forty-125 fifth and the fiftieth north parallels.

Dataset B consisted of a panel of untested environments across Europe. The studied early maturity groups '000' and '00' soybean cultivars are grown from France to Russia in locations situated between the fortyfifth° and the fifty-second parallels. This area includes regions in France, Germany, Belgium, Switzerland, the Czech Republic, Poland, Ukraine and Russia. Forty-nine locations were selected across this axis over a span of ten years (2012 to 2021) to produce 490 virtual environments. 131 Meteorological data for the two datasets were extracted from the Agri4Cast Resources Portal 132 (https://agri4cast.jrc.ec.europa.eu/dataportal/). The closest grid point (in kilometres) was attributed to each environmental location. Daily minimum and maximum temperatures (°C), daily solar radiation (MJ m⁻² 133 134 day⁻¹) and daily precipitation (mm) were extracted from 1 January to 31 December. Soil parameters extracted 135 from raster files produced by the European Soil Data Centre (https://esdac.jrc.ec.europa.eu/) were used to 136 characterise the soil of each location (1 km/1 km grid) using QGIS3 (v. 3.14.1). Clay content (%), silt content 137 (%), sand content (%), gravel content (%), organic carbon content (%), total nitrogen content (%) and bulk 138 density (g cm-3) from topsoil and subsoil were extracted as well as the depth available to roots (cm).

139

140 2.2. Eco-climatic factors calculation

141 A previous study using Dataset A (Elmerich et al., 2023), identified 20 out of 126 Eco-climatic Factors 142 (EFs) for their primary impact on Genotype by Environment Interactions (GEI). The 126 EFs used in this 143 study consisted of climatic variables calculated between two developmental stages (*i.e.* phenophases). Five 144 major categories of environmental variables were used: period duration, temperature, water, solar radiation 145 and stresses. Seven soybean phenophases were used: Sowing to EMergence (SEM), EMergence to Flower 146 Initiation (EMFI), Flower Induction to First Flower (FIFF), First Flower to First Pod (FFFP), First Pod to 147 First Seed (FPFS), First Seed to End of Pod (FSEP), End of Pod to Physiological Maturity (EPPM). These 148 phenophases were calculated based on the simulation of stages using the DSSAT-CROPGRO-soybean model 149 (Boote et al., 1998; Jones et al., 2003; Hoogenboom et al., 2019). The 126 EFs were sorted using partial least 150 square regression and the variable importance in projection scores. This enabled identification of the main 151 GEI-drivers.

152

153 2.3. Environmental classification based on k-means clustering

154 Clustering consists of grouping individuals into an optimal number of clusters based on one or a set of 155 variables. In our study, the aim was to group the 602 environments (Datasets A and B) into clusters that 156 represented the environment types; the intra-cluster GEI were minimised and the inter-cluster GEI were 157 maximised. The environmental classification should be defined according to crucial environmental factors 158 that affect GEI in the Target Population of Environments (TPE) (Annicchiarico, 2021). The set of 20 EFs identified by Elmerich et al. (2023) appeared to be too large for the clustering. Usually, the number of variables used for clustering ranges from 6 to 12 (Corlouer et al., 2019; Schoving et al., 2022). Thus, 888 896 clustering models were tested (*i.e.* the combination of 6 to 12 EFs among the 20). K-means clustering was performed with R software (v4.2.1) using the Euclidean distance measure and the complete linkage method. For each clustering model, the optimal number of clusters was determined using the *nbclust* package in the R software (v4.2.1).

165

166 2.4. Linear models

167 Each model was tested using R software (v4.2.1).

For each k-means model, a linear mixed-effects model – Equation 1 (E1) – was applied on the data of Dataset A (112 environments). Only cultivars tested at least three times in each cluster were kept. The model tested the fixed effects of Genotype (G), Cluster (C) and Genotype by Cluster Interactions (G x C), and the random effect of the Environments nested in the Clusters C(E) across the network:

172
$$Y_{ijk} = \mu + G_i + C_j + (G \times C)_{ii} + C_j(E_k) + \varepsilon_{ijk},$$
 (E1)

where Y_{ijk} is the seed yield of genotype *i* in cluster *j* in the environment *k*, μ is the population mean, G_i stands for the effect of genotype *i*, C_j stands for the effect of cluster *j*, $(G \ge C)_{ij}$ stands for the effect of the interaction between genotype *i* and cluster *j* and $C_j(E_k)$ stands for the effect of the environment *k* nested in the cluster *j*. The residual is ε_{ijk} .

A linear model – Equation 2 (E2) – was applied to the data of Dataset A (112 environments) to test the
effects of the EFs used in the clustering on the GxC:

179
$$GC_{ijk} = \sum_{n=1}^{p} \alpha_n EF_n + \varepsilon_{ijk}, (p \in [6; 12]),$$
 (E2)

180 where, GC_{ijk} is the genotype by cluster interaction effect of genotype *i* in cluster *j* in the environment *k*, and 181 α_n stands for the linear regression coefficient of the eco-climatic factor *n*, EF_n. The residual is ε_{ijk} .

The evaluation of each model was based first on the significance of the GxC effect. A significant GxC effect indicated that there was some variation among genotypes in their rank orders between clusters. To ensure that the significance of the GxC effect was not driven by a single or a few genotypes and clusters, the percentage of significant interactions between specific genotypes and clusters was calculated. Finally, the capacity to explain the GxC effect using the EFs drawn on in the clustering model was evaluated.

187

188 2.5. Genotype plus genotype by block of environments biplot

Complementary principal component analyses were conducted on environment-centred genotype by environment, grouped by environment type, matrix (Cooper & DeLacy, 1994; Yan & Kang, 2003; Laffont et al., 2013). The Genotype plus Genotype by Block of environments biplot (GGB biplot) displays the genotype and genotype by block effects of a multi-environment trial dataset. The blocks of environments in our study were the environment types.

The GGB biplots can be interpreted such that genotypes located near the origin might either have all their values close to the environment means (low performance variation), given that the data were environment centred, or their variability is located in another dimension. Similarly, environments close to the origin may have little variability across genotypes or may not fit well in two dimensions. Genotypes that are close together have similar performance across environment types (Chapman et al., 1997). The R package *gge* was used for the analyses (Laffont et al., 2013).

The GGB biplots allowed us to assess genotypic performance across environment types. Their construction required a balanced multi-environmental trial dataset. Thus, two multi-environmental trials including varied years and locations were used in our study. The first multi-environmental trial (MET1) included seven cultivars from maturity group '00' and 28 environments grouped in three environment types. The second multi-environment trial (MET2) included five cultivars from maturity group '000' and 44 environments grouped in three environment types.

206

207 2.6. Weighted and unweighted multi-environment trial data

A MET dataset from 2018 was used that included ten '00' cultivars that were tested in twelve locations. For the unweighted selection strategy, the mean performance by genotypes was calculated as the arithmetic mean of the performance in the twelve locations. The means of the ten genotypes were ranked from 1 to 10. For the weighted selection strategy, the mean performance was calculated – Equation 3 (E3) – (Podlich et al., 1999).

213
$$w_k = \sum_{i=1}^{E} e_i \sum_{j=1}^{E_i} \frac{y_{kj}}{E_i}$$
(E3)

where w_k is the weighted mean performance of the genotype k, e_i is the frequency of occurrence of the environment type i in the TPE, y_{jk} is the phenotypic performance of the genotype k in environment j, and E_i is the number of environments of type i in the MET.

217

218 **3. Results**

219 3.1. Eco-climatic factors set used for environmental clustering

A total of 888 896 clustering models were tested. Each model used a unique combination of Eco-climatic Factors (EFs) that were identified as major Genotype by Environment Interactions (GEI) drivers in a previous study conducted by Elmerich et al. (2023). The best clustering model was identified based on the selection criteria detailed in *section 2.5*. In this model, 1.3% of the genotype yield variance was explained by the Genotype by Cluster Interactions (GxC) effect. The complete analyse of variance table can be found in supplementary table 2. Finally, the combination of EFs used in this clustering model allowed 88% of the GxC effect to be explained.

This model used a set of 12 EFs to define five clusters. The 12 EFs were: the number of days below 15°C from Flower Induction to First Flower (FIFF) and from the First Flower to First Pod (FFFP), the duration of the FFFP period, the photothermal quotient during FFFP, the minimal temperature from First Pod to First Seed (FPFS), the solar radiation quantity during FPFS, the evapotranspiration potential during FPFS, the photoperiod from First Seed to End of Pod (FSEP), the number of days above 30 and 34°C during FSEP, the photoperiod from End of Pod to Physiological Maturity (EPPM) and the number of days above 30°C during EPPM.

234

235 3.2. Characterisation of the five environment types

The selected clustering model allowed the identification of five clusters that were referred to as environment types. The twelve EFs used for the clustering strongly discriminated the environment types (pvalue < 0.001 for each EF) (Fig. 1).



239

Fig. 1. Characterisation of the five environment types by the 12 eco-climatic factors used for the clustering. The phenophases are indicated in parenthesis with the following stages: Flower Induction (FI), First Flower (FF), First Seed (FS), First Pod (FP), End Pod (EP) and Physiological Maturity (PM). The smaller inner circle represents the minimum value and the outer cycle represents the maximal value for each eco-climatic factor. The colours of the lines indicate the environment type: blue for the first environment type (ET-1), green for the second environment type (ET-2), orange for the third environment type (ET-3), yellow for the fourth environment type (ET-4) and red for the fifth environment type (ET-5).

247

248 As each environment was initially characterised by a set of 126 EFs, the latter were used to precisely 249 describe the environment types if a significant difference was observed (p-value ≤ 0.05) (Table 1). The first 250 environment type (ET-1) differed greatly in the high quantity of solar radiation (ca. 1088 MJ m⁻²), 251 precipitation (ca. 153 mm) and potential evapotranspiration (ca. 231 mm) as well as important cold stress (ca. 26 days below 10°C) during the vegetative growth. The ET-1 reproductive growth conditions showed 252 253 low precipitation (ca. 82 mm) and ETP (ca. 187 mm) as well as high thermal amplitude (ca. 13°C) and heat 254 stress (ca. 12.7 days above 30°C). The second environment type (ET-2) was characterised by non-stressful 255 conditions during the vegetative growth followed, during the reproductive growth, by high water stress (ca. 0.47), solar radiations (ca. 1131 MJ m⁻²) and thermal amplitude (ca. 12.8 °C), with both chilling (ca. 17 days 256 257 below 15°C) and heat stress (ca. 7.3 days above 30°C) at the beginning and the end of the period respectively. 258 The third environment type (ET-3) was distinguishable by the absence of stressful or extreme conditions 259 throughout the crop cycle. The fourth environment type (ET-4) was denoted by the low precipitation (ca. 70 260 mm), warm temperature (ca. 18 °C) and short duration (ca. 39 days) of the vegetative growth. The ET-4

reproductive growth conditions were differentiated by high solar radiation intensity (ca. 23 MJ m⁻² d⁻¹), high 261 262 water (ca. 0.37) and severe heat stress (ca. 6 days above 34°C). The fifth environment type (ET-5) showed high precipitation (ca. 154 mm), solar radiation quantity (ca. 1084 MJ m⁻²), cold stress (ca. 27 days below 263 10°C) and period duration (ca. 56 days) during the vegetative growth. The ET-5 reproductive growth was 264 differentiated by the high precipitation (ca. 164 mm) and the low water stress index (ca. 0.11), the solar 265 radiation intensity (ca. 18.1 MJ m⁻² d⁻¹), the thermal amplitude (ca. 10.3 °C) and the heat stress (ca. 4 days 266 267 above 30°C). Among the 602 environments used for the clustering, 83 (13.8%) belonged to ET-1, 106 (17.6%) belonged to ET-2, 161 (26.7%) belonged to ET-3, 93 (15.4%) belonged to ET-4 and 159 (26.4%) 268 269 belonged to ET-5.

Table 1. Description of the five environment types identified by clustering. For each Eco-climatic Factor (EF), the environment type effect was tested by Kruskal-Wallis non-parametric tests. When this effect was significant, a Dunn's multiple comparison test was performed in order to determine environment types (using Bonferroni adjusted *p*-values). In the table, yellow and blue shadings correspond to the highest and lowest EF means, respectively, that significantly differed from the others. The EFs written in bold correspond to those used for the k-means clustering. Abbreviations used in the table: evapotranspiration (ET), potential evapotranspiration (ETP), photothermal quotient (PTQ). The duration ranged from 5 to 52 days. The photoperiod ranged from 12.83 to 16.31 hours. The solar radiation intensity ranged from 5.3 to 29.7 MJ m² d⁻¹. The solar radiation quantity ranged from 86 to 758 MJ m². The PTQ ranged from 0.59 to 3.72. The precipitation amount ranged from 0 to 246 mm. The ET ranged from 2 to 141 mm. The ETP ranged from 8 to 154 mm. The water stress index ranged from 0 to 0.99. The minimum, average and maximum temperatures ranged from 0 to 22.9°C, 1.8 to 29.5°C and 3 to 37.1°C, respectively. The number of days with a minimal temperature below 10 or 15°C ranged from 0 to 50 days in both cases. The number of days with a minimal temperature above 30 or 34°C ranged from 0 to 18 days and 0 to 10 days, respectively. The thermal amplitude ranged from 2.5 to 18.7°C.

Enviro nment types	Variables	Sowing - Emergence	Emergence - Flower Induction	Flower Induction - First Flower	First Flower - First Pod	First Pod - First Seed	First Seed - End Pod	End Pod - Physiological Maturity	Overview		
ET-1	Duration/Photoperiod		Long period			Short days	Short days		Wet, cold and		
	Solar radiation	Quantity and PTQ	Quantity		PTQ	Quantity			shaded conditions		
	Water availability	Precipitation and ETP	Precipitation, ET and ETP	ET	Precipitation		ETP	ETP	during vegetative growth. Dry, warm		
	min	Cold stress	Cold stress			\mathbf{T}° min			and sunny		
	Temperature moy				Amplitude	Amplitude	Amplitude		reproductive		
	max	T° max		Heat stress	Heat stress		Heat stress		growth		
	Duration/Photoperiod				Long period	Long period	Short period				
ET-2	Solar radiation				PTQ	Quantity	PTQ		No stressful		
	Water availability				Precipitation	ETP and Water	Precipitation and ET	Precipitation and ET	conditions during vegetative growth. Very dry sunny		
					ETP and Water stress	stress	Water stress	Water stress	and variable temperatures		
	min				Chilling stress	T° min			during the		
	Temperature moy					Amplitude		Amplitude	reproductive		
	max						Heat stress (34°C)	Heat stress (30°C)	growin		
ET 2	Duration/Photoperiod					Long days			Low stressful		
E1-3	Solar radiation			PTQ	PTQ				conditions		

	Water availabilit	у					ET		throughout the		
	mi	n			Cold stress	T° min			crop cycle.		
	Temperature mo	у		Amplitude							
	ma	X		Heat stress			Heat stress				
	Duration/Photoper	iod Short peric	od Short period	Short period	Short period		Long days	Long days			
	Solar radiation	Quantity and	PTQ Quantity and PTQ	Quantity and PTQ	PTQ	Intensity, PTQ	Intensity	Intensity			
ET-4	Water availability	Precipitation	, ET Precipitation, ET	Precipitation, ET	Dessinitation		Precipitation	ET	Short, dry and		
		and ETP	and ETP	and ETP	Precipitation		ETP and Water stress	ETP and Water stress	warm conditions during the		
		T° min	T° min						Dry, hot and sunny		
	mi Temperature	n Cold stres	s Cold stress	Chilling stress	Chilling stress	T ^o min	T ^o min	T ^o min	reproductive arowth		
	mc	у					T° average	T° average	growin		
	ma	x T° max	T° max and Heat stress	T° max	T° max	T° max and Heat stress	Heat stress	Heat stress			
	Duration/Photoper	iod Long perio	d	Long period			Short days	Short days			
ET-5	Solar radiation	Intensity Quantity and	Quantity		PTQ	Quantity	Intensity and PTQ	Intensity	Long, wet and cold		
	Water availability	Precipitation, ET		Draginitation	Draginitation	Precipitation	Precipitation and ET	Precipitation and ET	conditions during vegetative growth.		
		and ETP		Precipitation	Precipitation	ETP and Water stress	ETP and Water stress	ETP and Water stress	shaded conditions		
	mi	n Cold stres	S	Chilling stress	Chilling stress	\mathbf{T}° min		T° min	reproductive		
	Temperature mc	re moy			Amplitude	Amplitude	Amplitude and T° average	Amplitude and T° average	growth		
	ma	x T° max			T° max and Heat stress	T° max and Heat stress	Heat stress	Heat stress			

224 3.3. Environment types occurrences in the European early soybean target population of environments 225 Fig. 2 displays the frequencies of environment types for the Dataset B (49 locations over 10 years). The 226 first environment type (ET-1) (10.6% in Dataset B) occurred principally in France and Germany. The second 227 environment type (ET-2) (19.1% in Dataset B) was present at least one year in all locations from France to 228 the middle of Ukraine. The third environment type (ET-3) (28% in Dataset B) occurred at least one year in 229 42 locations but showed high frequencies in locations situated in central Europe. The fourth environment 230 type (ET-4) (16.7% in Dataset B) characterised mostly Eastern Europe locations and a few Southern France 231 environments. The fifth environment type (ET-5) (25.5% in Dataset B) was present from France to Poland 232 with high frequencies in Northern France, Belgium, Switzerland and Germany.

Moreover, around 12% of the TPE locations presented low environment types repeatability (*i.e.* most frequent environment type frequency < 33%), 53% presented intermediate environment types repeatability (*i.e.* 33% < most frequent environment type frequency \leq 66%) and 35% presented high environment types repeatability (*i.e.* most frequent environment type frequency > 66%) (Couëdel et al., 2021).



Ź . .

Fig. 2. Environment types frequency of occurrence from France to Russia (European early soybean production area) over ten years. For the 49 locations of Dataset B, the environment types' frequency of occurrence was calculated and represented on the map using pie charts. The colours of each pie section indicate the environment type: blue for the first environment type (ET-1), green for the second environment type (ET-2), orange for the third environment type (ET-3), yellow for the fourth environment type (ET-4) and red for the fifth environment type (ET-5).

244

The temporal distribution (2012-2021) of the five environment types was highly dependent on the environment types considered (**Fig. 3**). In contrast to the other years, the years 2013, 2014, 2014, 2017, 2018 and 2020 were largely dominated by only one environment type. The ET-3 and ET-4 were present each year (at least in 6 or 4 locations, respectively), while ET-1, ET-2 and ET-5 were not represented in 2, 2 and 1 year, respectively.



250

Fig. 3. Temporal distribution of the five environment types from 2012 to 2021 using the 49 locations in Dataset B. The colours indicate the environment type: blue for the first environment type (ET-1), green for the second environment type (ET-2), orange for the third environment type (ET-3), yellow for the fourth environment type (ET-4) and red for the fifth environment type (ET-5).

256 3.4. Genotypic performance linked with envirotyping

257 Stratifying environments by environment types using Genotype plus Genotype by Block of environments 258 biplots (GGB biplot) enabled the interpretation of cultivar responses in the TPE. With the GGB biplot 259 representation cultivars can be compared by projecting a perpendicular from the genotype to the environment 260 type vector for any particular environment type. Cultivars that are further along in the positive direction of 261 the environment type vector have higher yields than other cultivars in said environment type, and vice versa. 262 In the multi-environmental trial 1 (MET1), which included seven cultivars and 28 environments grouped in 263 three environment types, the cultivar GEN2 was strongly positively correlated with ET-4 and positively 264 correlated with ET-1 and ET-5, and GEN4 was also positively correlated with ET-4 but negatively correlated 265 with ET-1 and ET-5 (Fig. 4a). In the multi-environmental trial 2 (MET2), which included five cultivars and 266 44 environments grouped in three environment types, the cultivars GEN8 and GEN11 were both negatively

267 correlated with all environment types whereas GEN9, GEN10 and GEN11 were positively correlated with



268 ET-1 but negatively correlated with ET-5 (Fig. 4b).

- 270 Fig. 4. Environment-standardised Genotype plus Genotype by Block of environments biplots illustrating 271 contrasted cultivars' performances depending on environment types in two multi-environmental trials from the Dataset A. (a) The first multi-environmental trial included seven cultivars from maturity group '00' (GEN 272 273 1 to GEN 7) and 28 environments (e1 to e28) grouped in three environment types (ET-1, ET-4 and ET-5). 274 (b) The second multi-environment trial included five cultivars from maturity group '000' (GEN 8 to GEN 275 12) and 44 environments (e29 to e72) grouped in three environment types (ET-1, ET-3 and ET-5). The percentages of the total genotype plus genotype by environment variation explained by the main two principal 276 277 components are shown in parentheses. The colours indicate the environment type: blue for the first (ET-1), 278 orange for the third (ET-3), yellow for the fourth (ET-4) and red for the fifth (ET-5).
- 279

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280 3.5. MET-TPE alignment for MET data evaluation

281 In the 2018 Multi-Environment Trials (MET) dataset presented in Table 2, one environment belonged to 282 ET-1 (8.3%), seven belonged to ET-2 (58.3%), three belonged to ET-3 (25%), one belonged to ET-4 (8.3%) 283 and zero belonged to ET-5 (0%). This MET did not satisfactorily represent the frequencies of environment 284 types within the TPE (14, 18, 27, 15 and 26%, respectively for ET-1 to ET-5). Changes in cultivar ranks 285 between unweighted and weighted data were observed and ranged from no change (Genotype 3, 6 and 8) to 286 a six-rank position change (Genotype 9). If a breeder was allowed to keep only 10% of the tested cultivars 287 for future testing and/or deployment, his choices would differed depending on the strategy. For instance, Genotype 4 would surely be advanced if data were weighted by the environment types frequencies observed 288 289 in the TPE but would most likely have been eliminated if data were unweighted.

291 **Table 2.** Example of the use of unweighted vs. weighted data analyses on the final breeding decisions. The Target Population of Environments (TPE) consists of five types of environments with a frequency of 292 293 occurrence of 0.14, 0.18, 0.27, 0.15 and 0.26, respectively. Ten genotypes were evaluated across 12 environments within a Multi-Environment Trial (MET) included in the Dataset A. In this MET, the first 294 environment type (ET-1) has been sampled in the MET environment 1, the second environment type (ET-2) 295 296 has been sampled in MET environments 2 to 8, the third environment type (ET-3) has been sampled in MET 297 environments 9 to 11, the fourth environment type (ET-4) has been sampled in MET environment 12 and the 298 fifth environment type (ET-5) has not been sampled in the MET. The grain yield performance of the 299 genotypes is expressed in guintals by hectare. The estimated mean yield performance and rank are given for 300 each genotype.

	Environment Types in MET											Data analyses				
	ET-1 $(e_1 = 0.14)^1$		ET-2 $(e_2 = 0.18)^1$					ET-3 $(e_3 = 0.27)^1$			ET-4 $(e_4 = 0.15)^{-1}$	1 Unweighted		Weighted		
Env. in MET	1	2	3	4	5	6	7	8	9	10	11	12	Mean	Rank	Mean	Rank
Genotype 1	51.1	28.4	46.4	25.4	30.2	34.1	43.7	33.6	41.7	34.9	38.6	37.7	37.2	9	29.2	6
Genotype 2	50.3	40.0	44.1	24.0	27.5	36.2	48.8	30.6	44.5	38.8	42.2	34.6	38.5	4	29.8	3
Genotype 3	41.1	28.7	43.2	20.9	30.4	36.6	43.1	31.5	45.3	37.4	32.1	37.1	35.6	10	27.5	10
Genotype 4	52.5	30.5	47.0	24.6	34.5	30.4	45.3	33.1	43.1	42.0	31.7	40.8	38.0	6	30.1	2
Genotype 5	49.1	30.5	48.0	23.0	32.2	39.7	46.8	32.4	42.2	35.7	40.4	36.1	38.0	5	29.2	5
Genotype 6	54.5	27.4	45.2	25.4	36.2	42.2	47.9	33.2	42.9	38.0	43.1	39.0	39.6	1	31.0	1
Genotype 7	52.4	33.7	52.8	24.5	34.5	35.8	46.8	32.9	35.7	36.3	42.2	37.1	38.7	3	29.7	4
Genotype 8	48.0	33.3	51.5	24.3	29.7	31.2	45.1	33.6	46.5	37.3	36.4	36.1	37.8	7	29.1	7
Genotype 9	42.0	35.2	54.8	25.6	31.2	41.4	48.4	33.9	43.9	39.6	38.9	35.0	39.2	2	28.9	8
Genotype 10	41.0	31.4	48.9	25.2	31.5	38.3	44.9	33.2	38.3	39.0	40.5	38.4	37.6	8	28.4	9

³⁰¹ ¹Frequency of each environment type in the TPE

302

303 4. Discussion

4.1. Five environment types characterised the European early soybean target population of environments
 The environmental clustering using twelve Eco-climatic Factors (EFs) that contrasted in terms of climatic

variables and phenophases explained 88% of the cultivar yield variation attributed to Genotype by Cluster interaction (GxC). Thanks to the clustering, breeders will be able to minimise the GxC variance in their prediction models and focus more on the genetic variation *per se* (Messina et al., 2018). The highly significant and large GxC variance component compared to the genotype and cluster effects demonstrated the interest in defining environment types within the European early maturity soybean production area. It would be interesting to confront this model performance with a richer database.

The results highlighted five environment types that represent the European early soybean TPE. The first environment type (ET-1) was characterised by wet, cold and shaded conditions during the vegetative growth 314 and warmth and high radiation but no drought conditions during the reproductive growth. The second 315 environment type (ET-2) showed no stressful conditions during the vegetative growth but showed drought 316 stress, high radiation and variable temperatures during the reproductive growth. The third environment type 317 (ET-3) was distinguishable by the absence of stressful or extreme conditions throughout the crop cycle. The 318 fourth environment type (ET-4) was marked by the short, dry and warm conditions during the vegetative 319 growth as well as by the dry, hot and high radiation conditions during the reproductive growth. The fifth 320 environment type (ET-5) was characterised by long, wet and cold conditions during the vegetative growth as 321 well as wet, cool and shaded conditions during the reproductive growth. The environment types could be 322 referred to as follows: 'Temperate Humid', 'Temperate dry', 'Temperate with no stress', 'Warm dry' and 323 'Cool humid', respectively (IPCC, 1997). The different environment types were primarily contrasted by the 324 intensity and timing of stresses relative to temperatures (cold stress during the vegetative growth and heat 325 stress during the reproductive growth) and water availability (precipitation amount, evapotranspiration and 326 drought). Despite disconnected/juxtaposed geographical areas, our classification is consistent with the 327 classification established by Schoving et al. (2022) that identified environment classes on the basis of levels 328 of temperatures and drought stresses. Our non-a priori approach allowed us to identify the combinations of 329 stresses and conditions to which soybean cultivars are expected to similarly respond. This envirotyping 330 approach allowed to define the first European typology of environments for soybean and can be used to 331 design new trial networks or improve existing ones.

332

333 4.2. Varied occurrences of environment types in the target population of environments

334 The results demonstrated that the environment types distribution in the TPE over ten years (from 2012 to 335 2021) was heterogenous and lack generalities. Contrary to the six mega-environments defined by Kurasch et 336 al. (2017), in the early soybean TPE, environment types' occurrences neither followed annual (one year 337 corresponding to one environment type) nor geographical (longitudinal or latitudinal) distributions. This 338 difference could be attributed to our wider study area and to the environmental descriptors that were based 339 on maturity dates in their study as opposed to EFs. The ET-1 showed low repeatability, ET-2 and ET-5 340 showed intermediate repeatability, the ET-4 showed high repeatability and the ET-3 equally showed 341 intermediate and high repeatability. The identification of highly repeatable environment types (mainly

342 located in Eastern Europe) is key to designing breeding strategy for specific adaptation. In environments where specific adaptation appears best suited, the interactions between genotypes and other environmental 343 344 characteristics, such as soil properties, cultural practices or biotic stress incidences could be investigated. On 345 the other hand, the highlight of low environment types repeatability in Western Europe will help breeders 346 aim for broad adaptation, as future growth conditions are highly unpredictable (de la Vega & Chapman, 347 2006). The broad adaptation of cultivars could be reached by designing more plastic cultivars, *i.e.* cultivars 348 having traits with a large reaction norm, leading to a stable performance under a large set of growing 349 conditions (Sambatti & Caylor, 2007). Future work should evaluate soybean above and below ground traits 350 plasticity in response to the main EFs driving GEI to identify relevant traits to focus on for broad adaptation. 351 Among these, cold stress during the early vegetative growth has never been investigated and seems of great 352 importance (Elmerich et al., 2023).

Variations in the spatial and temporal distribution of the environment types across the TPE revealed the complexity to design efficient breeding programs that will control possible genotype by environment interactions and select for climate adaptation as much as. In the context of climate change, these five environment types could be conserved but their spatial and temporal distributions could be affected. Thus, monitoring the environment types distribution will be essential. The perspectives will be to simulate virtual environments using different forecasting climate scenarios, *e.g.* SSP5-8.5 and SSP2-4.5 (IPCC, 2022) and confront our results to a projected environment types distribution (Bustos-Korts et al., 2016).

360

361 4.3. Environmental characterisation, an informative tool for breeding decisions

The environmental characterisation aimed to increase the understanding of GEI effects. This is a crucial step when attempting to expand a new crop northward, and to assist breeders dealing with GEI (Chenu, 2015). In our study, the GEI effect, was larger for yield, compared with the genotype effect. The same ranking was previously reported in the literature (Dreccer et al., 2007; Chenu et al., 2011).

Using METs conducted in France, differences in genotypes performances were observed across environment types. Thanks to the environmental characterisation of the European early soybean TPE, predictions can be made on genotypes performances in non-tested locations that belong to the same environment type. Better precision in these predictions could be achieved especially in Eastern Europe where

370 specific adaptation appears to be best suited. For instance, GEN 2 showed its best performance in ET-4; 371 therefore, it could be advised for use in Eastern Europe or used as a parent to develop bi-parental populations. 372 Envirotyping could increase performances predictions for new targeted areas in locations where trials have 373 not been conducted. The environment types frequencies within the TPE constitute an interesting result to 374 position current elite soybean cultivars on the market, according to their performance in each environment 375 type. It could also be used to design new cultivars specifically adapted to an environment type. This approach 376 allows the crop yield potential to be maximised for a targeted region and to match farmers' needs for 377 improved and locally designed germplasm (Annicchiarico, 2021).

378 A huge challenge when analysing METs for breeding purposes is evaluating how METs data match the 379 TPE in order to improve breeding decisions and enhance the genetic gain over generations. Comparisons 380 between weighted and unweighted MET phenotypic data support the importance of MET-TPE alignment 381 evaluation (Podlich et al., 1999; Cooper et al., 2023). Our results confirmed the efficiency of this approach 382 when climatic scenarios at the MET scale derived from the TPE (Table 2). When dealing with large and 383 complex GEI effects, as observed in the Western European soybean production area, breeders should weight 384 their phenotypic data and consider the MET-TPE alignment, particularly if they are selecting for broad 385 adaptation.

386

387 **5.** Conclusion

388 This study is the first to European described early soybean Target Population of Environments (TPE) to 389 control Genotype by Environment Interactions (GEI) for breeding purposes. Environmental characterisation 390 based on GEI-drivers is a powerful tool to tackle GEI and accelerate soybean genetic gain. Our method 391 explained a large part (88%) of the yield variation that is due to the GEI effects. The five environment types 392 observed across Europe and their repeatability varied in terms of geographical and temporal distributions. 393 The results underlined the necessity to combine specific (maximising cultivars performances in a given 394 environment type) and broad adaptations (maximising cultivars yield stabilities over different environment 395 types) depending on locations. Envirotyping represents a necessary progression within the era of data-driven 396 plant improvement (Xu, 2016; Crossa et al., 2021). These approaches allowed us to better predict genotypic 397 performances within specific environment types and improve the positioning of cultivars in the marketplace. Moreover, as demonstrated in this study, a better MET data interpretation and utilisation can be reached through MET-TPE alignment evaluation. To enhance the deployment of soybean northward, perspectives should be to design ideotypes that outperform the existing germplasm for specific or multiple environment types across the TPE.

402

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409

410 Authors contribution

411 CE, GB and BL designed the study. CE, MG and BL conducted the data analysis. CE, GB, PJ, MG, MPF
412 and BL wrote the manuscript. All authors read, commented on and approved the final version.

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414 **References**

- 415 Annicchiarico, P. (2021). Breeding gain from exploitation of regional adaptation: An alfalfa case study.
- 416 Crop Science, 61(4), 2254–2271. https://doi.org/10.1002/csc2.20423

417 Ayerdi Gotor, A., & Marraccini, E. (2022). Innovative Pulses for Western European Temperate Regions:

418 A Review. Agronomy, 12(1), Article 1. https://doi.org/10.3390/agronomy12010170

419 Basford, K. E., & Cooper, M. (1998). Genotype×environment interactions and some considerations of

420 their implications for wheat breeding in Australia This review is one of a series commissioned by the

421 Advisory Committee of the Journal. Australian Journal of Agricultural Research, 49(2), 153-174.

- 422 https://doi.org/10.1071/a97035
- 423 Boote, K. J., Jones, J. W., Hoogenboom, G., & Pickering, N. B. (1998). The CROPGRO model for grain
- 424 legumes. In G. Y. Tsuji, G. Hoogenboom, & P. K. Thornton (Eds.), Understanding Options for Agricultural
- 425 Production (Vol. 7, pp. 99–128). Springer Netherlands. https://doi.org/10.1007/978-94-017-3624-4_6

- Brennan, P. S., Byth, D. E., Drake, D. W., Lacy, Ihd., & Butler, D. G. (1981). Determination of the
 location and number of test environments for a wheat cultivar evaluation program. *Australian Journal of Agricultural Research*, *32*(2), 189–201. https://doi.org/10.1071/ar9810189
- 429 Bustos-Korts, D., Boer, M. P., Layton, J., Gehringer, A., Tang, T., Wehrens, R., Messina, C., de la Vega,

430 A. J., & van Eeuwijk, F. A. (2022). Identification of environment types and adaptation zones with self-

431 organizing maps; applications to sunflower multi-environment data in Europe. *Theoretical and Applied*

432 *Genetics*. https://doi.org/10.1007/s00122-022-04098-9

433 Bustos-Korts, D., Malosetti, M., Chapman, S., & van Eeuwijk, F. (2016). Chapitre 3-Modelling of

434 Genotype by Environment Interaction and Prediction of Complex Traits across Multiple Environments as a

435 Synthesis of Crop Growth Modelling, Genetics and Statistics. In X. Yin & P. C. Struik (Eds.), Crop Systems

- 436 Biology (pp. 55–82). Springer International Publishing. https://doi.org/10.1007/978-3-319-20562-5_3
- 437 Casadebaig, P., Gauffreteau, A., Landré, A., Langlade, N., Mestries, E., Sarron, J., Trépos, R., Vincourt,

438 P., & Debaeke, P. (2021). *Optimized cultivar deployment improves the efficiency and stability of sunflower*

439 crop production at national scale. https://doi.org/10.1101/2020.09.21.306076

- Chapman, S. C., Cooper, M., Hammer, G. L., & Butler, D. G. (2000). Genotype by environment
 interactions affecting grain sorghum. II. Frequencies of different seasonal patterns of drought stress are
 related to location effects on hybrid yields. *Australian Journal of Agricultural Research*, *51*(2), 209–222.
 https://doi.org/10.1071/ar99021
- Chapman, S. C., Edmeades, G. O., & Crossa, J. (1997). Genotype by environment effects and selection
 for drought tolerance in tropical maize. I. Two mode pattern analysis of yield | SpringerLink. *Euphytica*, *95*,
 1–9.
- Chauhan, Y. S., Solomon, K. F., & Rodriguez, D. (2013). Characterization of north-eastern Australian
 environments using APSIM for increasing rainfed maize production. *Field Crops Research*, *144*, 245–255.
- 449 https://doi.org/10.1016/j.fcr.2013.01.018
- 450 Chenu, K. (2015). Chapter 13-Characterizing the crop environment nature, significance and
- 451 applications. In V. O. Sadras & D. F. Calderini (Eds.), Crop Physiology (Second Edition) (pp. 321-348).
- 452 Academic Press. https://doi.org/10.1016/B978-0-12-417104-6.00013-3

- Chenu, K., Cooper, M., Hammer, G. L., Mathews, K. L., Dreccer, M. F., & Chapman, S. C. (2011).
 Environment characterization as an aid to wheat improvement: Interpreting genotype–environment
 interactions by modelling water-deficit patterns in North-Eastern Australia. *Journal of Experimental Botany*,
- 456 62(6), 1743–1755. https://doi.org/10.1093/jxb/erq459
- Chenu, K., Deihimfard, R., & Chapman, S. C. (2013). Large-scale characterization of drought pattern: A
 continent-wide modelling approach applied to the Australian wheatbelt spatial and temporal trends. *New*
- 459 Phytologist, 198(3), 801–820. https://doi.org/10.1111/nph.12192
- 460 Chenu, K., Porter, J. R., Martre, P., Basso, B., Chapman, S. C., Ewert, F., Bindi, M., & Asseng, S. (2017).
- 461 Contribution of Crop Models to Adaptation in Wheat. Trends in Plant Science, 22(6), 472-490.
- 462 https://doi.org/10.1016/j.tplants.2017.02.003
- 463 Cooper, M., & Byth, D. E. (1996). Understanding plant adaptation to achieve systematic applied crop
- 464 improvement—A fundamental challenge. In *Plant Adaptation and Crop Improvement* (M. Cooper and G. L.
- 465 Hammer, pp. 5–23). IRRI.
- Cooper, M., & DeLacy, I. H. (1994). Relationships among analytical methods used to study genotypic
 variation and genotype-by-environment interaction in plant breeding multi-environment experiments. *Theoretical and Applied Genetics*, 88(5), 561–572. https://doi.org/10.1007/BF01240919
- Cooper, M., Powell, O., Gho, C., Tang, T., & Messina, C. (2023). Extending the breeder's equation to
 take aim at the target population of environments. *Frontiers in Plant Science*, 14.
 https://www.frontiersin.org/articles/10.3389/fpls.2023.1129591
- 472 Corlouer, E., Gauffreteau, A., Bouchet, A.-S., Bissuel-Bélaygue, C., Nesi, N., & Laperche, A. (2019).
- 473 Envirotypes Based on Seed Yield Limiting Factors Allow to Tackle $G \times E$ Interactions. Agronomy, 9(12),
- 474 Article 12. https://doi.org/10.3390/agronomy9120798
- 475 Couëdel, A., Edreira, J. I. R., Pisa Lollato, R., Archontoulis, S., Sadras, V., & Grassini, P. (2021).
- 476 Assessing environment types for maize, soybean, and wheat in the United States as determined by spatio-
- 477 temporal variation in drought and heat stress. Agricultural and Forest Meteorology, 307, 108513.
- 478 https://doi.org/10.1016/j.agrformet.2021.108513
- 479 Crossa, J., Fritsche-Neto, R., Montesinos-Lopez, O. A., Costa-Neto, G., Dreisigacker, S., Montesinos-
- 480 Lopez, A., & Bentley, A. R. (2021). The Modern Plant Breeding Triangle: Optimizing the Use of Genomics,

- 481 Phenomics, and Environics Data. Frontiers in Plant Science, 12.
 482 https://www.frontiersin.org/article/10.3389/fpls.2021.651480
- de la Vega, A. J., & Chapman, S. C. (2006). Defining Sunflower Selection Strategies for a Highly
 Heterogeneous Target Population of Environments. *Crop Science*, 46(1), 136.
 https://doi.org/10.2135/cropsci2005.0170
- 486 Döttinger, C. A., Hahn, V., Leiser, W. L., & Würschum, T. (2023). Do We Need to Breed for Regional
- 487 Adaptation in Soybean? Evaluation of Genotype-by-Location Interaction and Trait Stability of Soybean in
 488 Germany. *Plants*, *12*(4), Article 4. https://doi.org/10.3390/plants12040756
- 489 Dreccer, M. F., Borgognone, M. G., Ogbonnaya, F. C., Trethowan, R. M., & Winter, B. (2007).
- 490 CIMMYT-selected derived synthetic bread wheats for rainfed environments: Yield evaluation in Mexico and
- 491 Australia. Field Crops Research, 100(2), 218–228. https://doi.org/10.1016/j.fcr.2006.07.005
- 492 Elmerich, C., Boulch, G., Faucon, M.-P., Lakhal, L., & Lange, B. (2023). Identification of Eco-Climatic
- Factors Driving Yields and Genotype by Environment Interactions for Yield in Early Maturity Soybean
 Using Crop Simulation. *Agronomy*, *13*(2), 322. https://doi.org/10.3390/agronomy13020322
- 495 FAOSTAT. (2021). Food and Agriculture Organization Corporate Statistical Database.
 496 https://www.fao.org/faostat/en/#data/QCL
- Holland, J. B., Nyquist, W. E., & Cervantes-Martínez, C. T. (2003). Estimating and interpreting
 heritability for plant breeding: An update. In *Plant breeding reviews* (Jules Janick, Vol. 22). John Wiley &
 Sons.
- 500 Hoogenboom, G., Porter, C. H., Boote, K. J., Shelia, V., Wilkens, P. W., Singh, U., White, J. W., Asseng,
- 501 S., Lizaso, J. I., Moreno, L. P., Pavan, W., Ogoshi, R., Hunt, L. A., Tsuji, G. Y., & Jones, J. W. (2019). The
- 502 DSSAT crop modeling ecosystem. In Advances in Crop Modeling for a Sustainable Agriculture (pp. 173–
- 503 216). Burleigh Dodds Science Publishing. https://doi.org/10.19103/AS.2019.0061.10
- 504 IPCC. (1997). Revised 1996 IPCC guidelines for national greenhouse gas inventories: Reference Manual
- 505 (Volume 3) (Houghton J.T., Meira L.G., Filho L.G., Lim B., Treanton K., Mamaty I., Bonduki Y., Griggs
- 506 D.J. and Callender B.A.). Intergovernmental Panel on Climate Change.
- 507 IPCC. (2022). Climate Change 2022: Impacts, Adaptation, and Vulnerability. (Contribution of Working
- 508 Group II to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change).

- Jones, J. W., Hoogenboom, G., Porter, C. H., Boote, K. J., Batchelor, W. D., Hunt, L. A., Wilkens, P. W.,
- 510 Singh, U., Gijsman, A. J., & Ritchie, J. T. (2003). The DSSAT cropping system model. European Journal
- 511 of Agronomy, 18(3-4), 235-265. https://doi.org/10.1016/S1161-0301(02)00107-7
- 512 Karges, K., Bellingrath-Kimura, S. D., Watson, C. A., Stoddard, F. L., Halwani, M., & Reckling, M.
- 513 (2022). Agro-economic prospects for expanding soybean production beyond its current northerly limit in
- 514 Europe. European Journal of Agronomy, 133, 126415. https://doi.org/10.1016/j.eja.2021.126415
- 515 Kurasch, A. K., Hahn, V., Leiser, W. L., Vollmann, J., Schori, A., Bétrix, C.-A., Mayr, B., Winkler, J.,
- 516 Mechtler, K., Aper, J., Sudaric, A., Pejic, I., Sarcevic, H., Jeanson, P., Balko, C., Signor, M., Miceli, F.,
- 517 Strijk, P., Rietman, H., ... Würschum, T. (2017). Identification of mega-environments in Europe and effect
- of allelic variation at maturity *E* loci on adaptation of European soybean: Soybean Adaptation to Europe.
- 519 Plant, Cell & Environment, 40(5), 765–778. https://doi.org/10.1111/pce.12896
- Laffont, J.-L., Wright, K., & Hanafi, M. (2013). Genotype Plus Genotype × Block of Environments
 Biplots. *Crop Science*, 53(6), 2332–2341. https://doi.org/10.2135/cropsci2013.03.0178
- 522 Marraccini, E., Gotor, A. A., Scheurer, O., & Leclercq, C. (2020). An Innovative Land Suitability Method
- 523 to Assess the Potential for the Introduction of a New Crop at a Regional Level. Agronomy, 10(3), 1–24.
- 524 https://doi.org/10.3390/agronomy10030330
- Messina, C. D., Technow, F., Tang, T., Totir, R., Gho, C., & Cooper, M. (2018). Leveraging biological insight and environmental variation to improve phenotypic prediction: Integrating crop growth models (CGM) with whole genome prediction (WGP). *European Journal of Agronomy*, *100*, 151–162. https://doi.org/10.1016/j.eja.2018.01.007
- Millet, E. J., Kruijer, W., Coupel-Ledru, A., Alvarez Prado, S., Cabrera-Bosquet, L., Lacube, S.,
 Charcosset, A., Welcker, C., van Eeuwijk, F., & Tardieu, F. (2019). Genomic prediction of maize yield across
 European environmental conditions. *Nature Genetics*, *51*(6), 952–956. https://doi.org/10.1038/s41588-0190414-y
- Podlich, D. W., & Cooper, M. (1998). QU-GENE: A simulation platform for quantitative analysis of
 genetic models. *Bioinformatics*, 14(7), 632–653. https://doi.org/10.1093/bioinformatics/14.7.632

- Podlich, D. W., Cooper, M., Basford, K. E., & Geiger, H. H. (1999). Computer simulation of a selection
 strategy to accommodate genotype environment interactions in a wheat recurrent selection programme. *Plant Breeding*, *118*(1), 17–28, https://doi.org/10.1046/j.1439-0523.1999.118001017.x
- 538 Rincent, R., Malosetti, M., Ababaei, B., Touzy, G., Mini, A., Bogard, M., Martre, P., Le Gouis, J., & van
- 539 Eeuwijk, F. (2019). Using crop growth model stress covariates and AMMI decomposition to better predict
- 540 genotype-by-environment interactions. Theoretical and Applied Genetics, 132(12), 3399-3411.
- 541 https://doi.org/10.1007/s00122-019-03432-y
- Sambatti, J. B. M., & Caylor, K. K. (2007). When is breeding for drought tolerance optimal if drought is
 random? *New Phytologist*, *175*(1), 70–80. https://doi.org/10.1111/j.1469-8137.2007.02067.x
- 544 Schoving, C., Champolivier, L., Maury, P., & Debaeke, P. (2022). Combining multi-environmental trials
- and crop simulation to understand soybean response to early sowings under contrasting water conditions.
- 546 European Journal of Agronomy, 133, 126439. https://doi.org/10.1016/j.eja.2021.126439
- 547 Sudarić, A. (2020). Soybean for Human Consumption and Animal Feed. IntechOpen.
- 548 Woollings, T. (2010). Dynamical influences on European climate: An uncertain future. *Philosophical*
- 549 Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences, 368(1924), 3733–
- 550 3756. https://doi.org/10.1098/rsta.2010.0040
- Xu, Y. (2016). Envirotyping for deciphering environmental impacts on crop plants. *Theoretical and Applied Genetics*, *129*(4), 653–673. https://doi.org/10.1007/s00122-016-2691-5
- 553 Yan, W., & Kang, M. S. (2003). *GGE biplot analysis: A graphical tool for breeders, geneticists, and* 554 *agronomists*. CRC Press.
- 555