

Two RFC1 splicing variants in CANVAS

Sacha Weber, Giulia Coarelli, Anna Heinzmann, Marie-Lorraine Monin, Nicolas Richard, Marion Gerard, Alexandra Durr, Vincent Huin

▶ To cite this version:

Sacha Weber, Giulia Coarelli, Anna Heinzmann, Marie-Lorraine Monin, Nicolas Richard, et al.. Two RFC1 splicing variants in CANVAS. Brain - A Journal of Neurology , 2022, 10.1093/brain/awac466 . hal-03920011

HAL Id: hal-03920011 https://normandie-univ.hal.science/hal-03920011

Submitted on 12 Jan2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

LETTER TO THE EDITOR

2

Two RFC1 splicing variants in CANVAS

- Sacha Weber,^{1,2} Giulia Coarelli,³ Anna Heinzmann,³ Marie-Lorraine Monin,⁴ Nicolas
 Richard,⁵ Marion Gerard,¹ Alexandra Durr³ and Vincent Huin⁶
- 5

6 Author affiliations:

- 7 1 Medical Genetics Department, Caen University Hospital, Caen, France
- 8 2 Neurology Department, Caen University Hospital, Caen, France
- 9 3 Sorbonne Université, Paris Brain Institute, APHP, INSERM, CNRS, Paris, France
- 10 4 Reference Center for Rare "Neurogenetic" Diseases, Department of Medical Genetics,
- 11 Pellegrin Hospital, Bordeaux University Hospital, Bordeaux, France
- 12 5 Department of Genetics, Normandy University, UNICAEN, Caen University Hospital, EA
- 13 7450 BioTARGen, F-14000 Caen, France
- 14 6 Univ. Lille, Inserm, CHU Lille, U1172 LilNCog (JPARC) Lille Neuroscience &
- 15 Cognition, F-59000 Lille, France
- 16
- 17 Correspondence to: Dr Vincent Huin
- 18 Lille Neuroscience & Cognition, F-59000 Lille, France
- 19 E-mail: vincent.huin@inserm.fr
- 20

21 **ORCID**:

- 22 Sacha Weber = 0000-0001-9363-5141
- 23 Giulia Coarelli = 0000-0002-7824-8343
- 24 Anna Heinzmann = 0000-0001-8007-2597
- 25 Marie-Lorraine Monin = 0000-0002-0116-036X
- 26 Nicolas Richard = 0000-0003-0702-7399

© The Author(s) 2022. Published by Oxford University Press on behalf of the Guarantors of Brain. All rights reserved. For permissions, please e-mail: journals.permissions@oup.com This article is published and distributed under the terms of the Oxford University Press, Standard Journals Publication Model (https://academic.oup.com/journals/pages/open_access/funder_policies/chorus/standard_publication_model) 1

- 1 Marion Gerard = 0000-0003-2319-0305
- 2 Alexandra Durr = 0000-0002-8921-7104
- 3 Vincent Huin = 0000-0001-8201-5406
- 4

5 We read with great interest the original article *"RFC1* nonsense and frameshift 6 variants cause CANVAS: clues for an unsolved pathophysiology" by Benkirane *et al.*¹

Biallelic *RFC1* (AAGGG)_n expansions are responsible for cerebellar ataxia,
neuropathy, and vestibular areflexia syndrome (CANVAS), a frequent cause of late-onset
ataxia, with sporadic or autosomal recessive inheritance.² Since their discovery, other
pathogenic pentanucleotide motifs have been reported in the literature.³⁻⁵

The authors reported two cases of heterozygous compound truncating variants of 11 RFC1 in two patients with CANVAS. They were null variants (one frameshift and one 12 nonsense variant) and were associated with a classical RFC1 (AAGGG)_n intronic expansion 13 in *trans*. For the first time, Benkirane *et al.*¹ succeeded in advancing our understanding of the 14 pathophysiology of CANVAS, which, until now, has remained elusive. The authors showed a 15 decrease in RFC1 mRNA levels in the patients' blood, supporting the hypothesis of a loss-of-16 function of the gene as the cause of the disease. Here, we report genetic and clinical data from 17 two additional unrelated families with heterozygous compound expansions in RFC1 and loss-18 of-function pathogenic variants. 19

20

21 Case reports

22 Case 1

A 62-year-old French man who was referred to the Caen University Hospital presented 23 with progressive proprioceptive ataxia associated with cerebellar signs and self-reported hand 24 25 myotonia. His familial history revealed possible late-onset gait problems in his mother and maternal grandfather. He started at age 37 with gait problems which were worse in the dark. 26 27 He had a chronic and unexplained cough since age 20. Clinical examination showed severe proprioceptive ataxia, absent deep tendon reflexes, a flexor plantar response, and sensory 28 29 problems, with a decreased tactile sense and abolition of the vibration sense of the extremities. He had a cerebellar syndrome, shown by a SARA (Scale for the Assessment and 30

Rating of Ataxia) score of 26/40, with action tremor and dysarthria. He exhibited vertical 1 nystagmus on videonystagmography, diplopia in the lateral view, a positive head impulse test, 2 and abolished visually-enhanced vestibulo-ocular reflex. He suffered from severe 3 dysautonomia with orthostatic hypotension, abnormal secretion and vomiting of mucus, 4 constipation, dysuria, erectile dysfunction, tachycardia, excessive sweating, and a disorder of 5 thermoregulation, with an intolerance to hot humid weather. He had normal hearing but had 6 7 had tinnitus since age 55. Myotonia was not found during the examination nor by EMG and he was diagnosed instead with myokymia. He developed akineto-rigid syndrome at age 56 8 9 and his gait problems and orthostatic hypotension progressively worsened and confined the patient to bed or a wheelchair at age 60. 10

EMG at age 50 showed sensory neuronopathy associated with moderate reductions in motor action potentials of the lower limbs. EMG at age 57 confirmed the previous results and showed a small decrease in motor potentials in the lower limbs associated with prolonged distal motor latency in the upper limbs. Brain MRI showed mild cerebellar atrophy involving the vermis. Dopamine transporter (DaT) SPECT showed bilateral and symmetrical reduction of striatal fixation. Computed tomography of the chest, abdomen, and pelvis was normal. The patient fully met the criteria for the diagnosis of clinically definitive CANVAS.⁶

We excluded acquired or metabolic causes of cerebellar atrophy, including infectious, 18 inflammatory, immune, carential, and neo- and paraneoplasic causes. Previous genetic 19 screening of the POLG and DMPK genes, dystrophic myotonia and episodic ataxia panel 20 genes (CACNA1A, CLCN1, KNJ2, KCN1A) by next-generation sequencing (NGS), and 21 polyglutamine expansions in the ATXN1, ATXN2, ATXN3, CACNA1A, ATXN7, and TBP 22 genes were negative. The FMR1 locus was normal. Analysis of mitochondrial DNA by NGS 23 was normal. Screening for Friedreich ataxia showed an expanded allele of ≈1000 triplets and 24 a normal allele without other pathogenic variants shown by Sanger sequencing and MLPA of 25 the FXN gene. We screened the RFC1 gene by fluorescent duplex PCR, repeat primer-PCR, 26 and LR-PCR with Southern-blotting of the PCR product, as previously described.⁷ The patient 27 28 carried an expanded $(AAGGG)_n$ allele and an allele of normal length with 10 pentaplets. Finally, we performed exome sequencing and found a heterozygous splicing variant in the 29 canonical acceptor site of exon 19 in the RFC1 gene: NM_002913.5: c.2535+2T>C, p?. 30 Segregation analyses performed on the proband's mother and son confirmed the compound 31 32 heterozygous status (Figure 1).

33

1 Case 2

A 38-year-old French man with a personal history of rheumatoid arthritis and no family history of neurodegenerative diseases presented with cerebellar and proprioceptive ataxia at the Neurogenetics Reference Center for Rare Diseases at the Salpêtrière Hospital in Paris. He started at age 32 with gait problems. Clinical examination showed a decreased sense of vibration at the ankles (2/8), cerebellar ataxia, and cerebellar dysarthria, with a SARA score of 8.5/40. He had brisk deep tendon reflexes and fasciculations of the lower limbs. He had suffered from a chronic and unexplained cough since age 18.

EMG showed generalized sensory neuronopathy. Somatosensory-evoked potentials 9 showed a somatosensory deficit. Eye movement recording showed dysmetric horizontal 10 11 saccades, decreased velocity of vertical saccades, numerous square waves, and downbeat nystagmus. Brain MRI showed cerebellar atrophy of both the vermis and cerebellar 12 hemispheres. Previous genetic screening, including for Friedreich ataxia and the search for 13 polyglutamine expansions in the ATXN1, ATXN2, ATXN3, CACNA1A, ATXN7, and TBP 14 genes showed no abnormal expansions. RFC1 gene screening showed an expanded 15 (AAGGG)_n allele and an allele of normal length with nine pentaplets. Exome sequencing 16 showed a heterozygous splicing variant in the canonical acceptor site of exon 20 in the RFC1 17 gene: NM_002913.5: c.2690+1G>A, p.? (Figure 1). 18

19

20 **Discussion**

Here, we confirm the report of Benkirane *et al.*¹ by adding two additional patients 21 22 whose phenotypes are characteristic of RFC1-related disorders and who carry heterozygous compound mutations in the RFC1 gene. Such compound heterozygous mutations are 23 reminiscent of the loss-of-function mutations associated with (GAA)_n intronic expansion in 24 the FXN gene, responsible for Friedreich ataxia, for which the phenotype of late onset forms 25 is very similar to that of CANVAS.^{8,9} Consistent with this hypothesis, we noted that the *RFC1* 26 gene is intolerant to loss-of-function variants based on the constraint score in the gnomAD 27 database (loss-of-function observed/expected upper bound fraction = 0.305). There are only 28 121 allele counts with a loss-of-function variant recorded in gnomAD v2.1.1, including three 29 counts with disputable annotation or quality and 98 counts of a splice variant with an 30 uncertain loss-of-function effect due to weak/unrecognized splice rescue. Moreover, Zhao et 31 *al.*¹⁰ showed that *RFC1^{-/-}* knock-out mice are embryonic lethal. 32

The two patients have first motor neuron signs (brisk deep tendon reflexes in case 2) 1 or second motor neuron signs (myokymia in case 1 and fasciculations in case 2). Mixed motor 2 and sensory neuronopathy was also observed in case 1. This is consistent with our previous 3 report of frequent motor neuron involvement in *RFC1* patients.⁷ Interestingly, the two cases 4 that we report had earlier onset than that previously reported for classical CANVAS. The 5 second difference is the presence of an MSA-like phenotype in the first case, not present in 6 7 the second, indicating that other factors are involved in this phenotypic expression of RFCI-8 related disorders.

9

10 Data availability

11 The data that support the findings of this study are available from the corresponding authors

12 upon reasonable request.

13

14 Acknowledgements

15 We are grateful to the patients for their participation in this study.

16

17 Funding

This work was funded by the University of Lille, the "Institut National de la Santé et de la Recherche Médicale" (INSERM), the Lille University Hospital, the Association des Aidants et Malades à Corps de Lewy (A2MCL) charity, and the France Alzheimer charity. This work was also supported by grants from the Programs d'Investissements d'Avenir LabEx (excellence laboratory) DISTALZ (Development of Innovative Strategies for a Transdisciplinary approach to ALZheimer's disease).

24

25 **Competing interests**

26 The authors report no competing interests.

27

28

1 References

- 2 1 Benkirane M, Da Cunha D, Marelli C, et al. RFC1 nonsense and frameshift variants cause
- 3 CANVAS: clues for an unsolved pathophysiology. Brain. Published online July 27,
- 4 2022:awac280. doi:10.1093/brain/awac280
- 5 2 Cortese A, Simone R, Sullivan R, et al. Biallelic expansion of an intronic repeat in RFC1 is
- a common cause of late-onset ataxia. Nat Genet. 2019;51(4):649-658. doi:10.1038/s41588019-0372-4
- 3 Tsuchiya M, Nan H, Koh K, et al. RFC1 repeat expansion in Japanese patients with lateonset cerebellar ataxia. J Hum Genet. 2020;65(12):1143-1147. doi:10.1038/s10038-020-0807-
- 10 X

4 Beecroft SJ, Cortese A, Sullivan R, et al. A Māori specific RFC1 pathogenic repeat
configuration in CANVAS, likely due to a founder allele. Brain. 2020;143(9):2673-2680.

- 13 doi:10.1093/brain/awaa203
- 5 Miyatake S, Yoshida K, Koshimizu E, et al. Repeat conformation heterogeneity in
 cerebellar ataxia, neuropathy, vestibular areflexia syndrome. Brain. 2022;145(3):1139-1150.
 doi:10.1093/brain/awab363
- 6 Szmulewicz DJ, Roberts L, McLean CA, MacDougall HG, Halmagyi GM, Storey E.
 Proposed diagnostic criteria for cerebellar ataxia with neuropathy and vestibular areflexia
 syndrome (CANVAS). Neurol Clin Pract. 2016;6(1):61-68.
 doi:10.1212/CPJ.0000000000215
- 7 Huin V, Coarelli G, Guemy C, et al. Motor neuron pathology in CANVAS due to RFC1
 expansions. Brain. 2022;145(6):2121-2132. doi:10.1093/brain/awab449
- 8 Campuzano V, Montermini L, Moltò MD, et al. Friedreich's Ataxia: Autosomal Recessive
 Disease Caused by an Intronic GAA Triplet Repeat Expansion. Science.
 1996;271(5254):1423-1427. doi:10.1126/science.271.5254.1423
- 9 Galea CA, Huq A, Lockhart PJ, et al. Compound heterozygous FXN mutations and clinical
 outcome in friedreich ataxia: Insights From Frataxin Structure and Function. Ann Neurol.
 2016;79(3):485-495. doi:10.1002/ana.24595
- 10 Zhao R, Russell RG, Wang Y, et al. Rescue of Embryonic Lethality in Reduced Folate
 Carrier-deficient Mice by Maternal Folic Acid Supplementation Reveals Early Neonatal

- 1 Failure of Hematopoietic Organs. Journal of Biological Chemistry. 2001;276(13):10224-
- 2 10228. doi:10.1074/jbc.C000905200
- 3
- 4

1 Figure legend

Figure 1 Pedigrees and RFC1 variants of the two families. A. Pedigrees of family1 (left) 2 and family 2 (right). The probands are indicated by an arrow. Filled black symbols denote 3 clinically affected members. Grey filled symbols denote members clinically affected by late-4 onset gait problems but who had not been examined by one of us. Numbers indicate the age at 5 onset in years for neurological signs (cough excluded). Open symbols indicate unaffected 6 7 individuals. A forward slash indicates deceased individuals. Intronic (AAGGG)_n RFC1 expansions are shown in bold. The two RFC1 splicing variants are shown in bold and in red. 8 **B.** Electropherograms of the heterozygous splicing variants in the *RFC1* gene in the probands 9

10 from family 1 (left) and family 2 (right).

- 11
- 12

