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ABSTRACT:

Microduplications of the 17q21.31 chromosomal region encompassing the *MAPT* gene, which encodes the Tau protein, were identified in patients with a progressive disorder initially characterized by severe memory impairment with or without behavioral changes that can clinically mimic Alzheimer disease. The unique neuropathological report showed a primary tauopathy, which could not be unanimously classified in a given known subtype, showing both 4R- and 3R-tau inclusions, mainly within temporal cortical subregions and basal ganglia, without amyloid deposits. Recently, two subjects harboring the same duplication were reported with an atypical extrapyramidal syndrome and gait disorder. To decipher the phenotypic spectrum associated with *MAPT* duplications, we studied 10 carriers from 9 families, including 2 novel unrelated probands, gathering clinical (n=10), cerebrospinal fluid (N=6), MRI (n=8), dopamine transporter scan (n=4), functional (n=5), amyloid (n=3) and Tau-tracer (n=2) PET imaging data as well as neuropathological examination (n=4). Ages at onset ranged from 37 to 57 years, with prominent episodic memory impairment in 8/10 patients, associated with behavioral changes in four, while two patients showed atypical extrapyramidal syndrome with gait disorder at presentation, including one with associated cognitive deficits. Amyloid imaging was negative but Tau imaging showed significant deposits mainly in both mesiotemporal cortex. Dopaminergic denervation was found in 4/4 patients, including three without extrapyramidal symptoms. Neuropathological examination exclusively showed Tau-immunoreactive lesions. Distribution, aspect and 4R/3R tau aggregates composition suggested a spectrum from predominantly-3R, mainly cortical deposits well correlating with cognitive and behavioral changes, to predominantly-4R deposits, mainly in the basal ganglia and midbrain, in patients with prominent extrapyramidal syndrome. Finally, we performed *in vitro* seeding experiments in HEK-biosensor cells. Morphological features of aggregates induced by homogenates of three *MAPT* duplication carriers showed dense/granular ratios graduating between those induced by homogenates of a Pick disease and a progressive supranuclear palsy cases. These results suggest that *MAPT* duplication causes a primary tauopathy associated with diverse clinical and neuropathological features.

Keywords: tau, MAPT duplication, progressive supranuclear palsy, Pick disease, Tau seeding.

INTRODUCTION

The *MAPT* gene encodes tau, a microtubule binding protein, which forms pathological cellular aggregates in several neurodegenerative disorders termed tauopathies [14, 20]. In primary tauopathies, tau is the driving pathological factor whereas in secondary tauopathies such as Alzheimer Disease (AD), the pathological process is associated with the interaction of tau with other proteins or peptides, namely the Aβ peptide in AD. Additionally, tauopathies can be divided into 3R, 4R or mixed subtypes according to the isoforms that accumulate. The *MAPT* messenger RNA can be alternatively spliced resulting in 6 isoforms of the tau protein. 4R-tau refers to tau isoforms including 4 microtubule-binding repeats whereas 3R-tau includes only 3 repeats due to the splicing out of *MAPT* exon 10 [14].

The 17q21.31 microduplication is a rare recurrent rearrangement involving four genes *(CRHR1, MAPT, STH* and *KANSL1*) which causes a recently recognized neurodegenerative disorder with poorly defined phenotypic features [15]. Among the first few cases initially reported, a constant clinical feature was severe memory impairment, associated in some cases with behavioral changes leading either to a clinical diagnosis of typical Alzheimer disease (AD) phenotype or behavioral variant of Alzheimer disease [1, 12, 15, 21]. In a previous report, we found that the cerebrospinal (CSF) biomarkers profile of the 17q21.31 duplication was consistent with an AD diagnosis in 3/4 patients, including tau-related neurodegenerative processes and abnormal amyloid β (Aβ) levels. However, amyloid positron emission tomography (PET) imaging did not support this conclusion [15]. So far, detailed neuropathological examination has been published only for one patient [1]. This case showed a marked atrophy of the frontal and the temporal lobes, hippocampus, amygdala, and a depigmentation of the substantia nigra. Immunostaining for Aβ and α-synuclein remained negative in all regions examined. The most prominent neuropathological alterations were 4R-tau inclusions, although 3R-tau positive neurofibrillary tangles were also observed in the hippocampus, entorhinal cortex, striatum, nucleus basalis of Meynert and substantia nigra. Despite its clinical presentation and CSF biomarkers profile in some patients, the 17q21.31 duplication thus appears to cause a pure tauopathy. The main pathophysiological mechanism associated with this duplication is a 1.6-1.9 fold increase of the *MAPT* messenger RNA as measured in blood samples of duplication carriers [15], presumably increasing tau protein levels. To further complicate the matter, it has recently been reported that this duplication was present in two subjects clinically diagnosed with a different phenotype, *i.e.* atypical extra-pyramidal syndrome with gait disorder [4]. However, the in-depth description of neuropathological findings in those last two cases was not reported. In the present article, we describe novel cases, reanalyze the previously published

cases with additional imaging and CSF data and report the detailed results of four neuropathological examinations.

METHODS

Cases

Eight demented cases with an *MAPT* microduplication collected from year 2009 to year 2019 by our laboratory were studied. Six cases were previously published [15, 21]. For two of those six cases, we add here neuropathological data that were not available at the time and updated clinical and paraclinical information. Two additional unpublished cases were identified in the meantime and added to this series. Finally, two cases, enrolled in a series of extrapyramidal syndromes, were previously published without neuropathological details [4], which we are providing here.

All patients (or legal representatives) gave informed, written consent for genetic analyses. All patients underwent a comprehensive clinical examination including personal medical and family history, neurological examination, neuropsychological assessment and, for some, lumbar puncture with CSF biomarkers analysis. Structural Magnetic Resonance Imaging (MRI), ^{123}I-Ioflupane Dopamine Transporter Single Photon Emission Computerized Tomography (Datscans) and ^{18}Fluro-deoxy-glucose (FDG) Positon Emission Tomography (PET) or single-photon emission computed tomography (SPECT) with hexa-methyl-propylene-amine-oxime (HMPAO) imaging were collected when performed as well as Amyloid and Tau-PET cerebral imaging (see details below). We collected clinical and paraclinical information from medical charts of all participants. Autopsies for cases were performed with the authorization of the patient himself or the next of keen. Cases ALZ_441_005 and ALZ_596_006 were obtained through the French national brain bank network, Neuro-CEB. We certify that the study was performed in accordance with the ethical standards as laid down in the 1964 Declaration of Helsinki and its later amendments.

Genetic analyses

Copy Number Variants (CNVs) were identified from whole exome sequencing data and confirmed by quantitative multiplex PCR of short fluorescent fragments (QMPSF) as previously described [15], except for the two additional cases identified by DNA chips (SNP array genotyping) and validated by qPCR [4]. H1/H2 *MAPT* haplotypes genotyping was determined by SNaPshot experiments using the rs1052553 single nucleotide polymorphism as a tag. Sub-haplotypes could not be determined because, in most individuals, it was not possible

to unambiguously phase the polymorphisms used to tag these sub-haplotypes.
Exon 10 and at least 20 bp of the surrounding splicing regions were re-sequenced with the Sanger technique using specific primers (available upon request).

CSF biomarker analysis

CSF biomarker analyses were carried out in 6 different laboratories as part of the diagnostic workup using a standardized procedure. CSF samples were obtained using a Sprotte needle and collected in polypropylene tubes. They were aliquoted after centrifugation into polypropylene Eppendorf tubes, then frozen at −80°C within 1 h. $A\beta_{42}$, Tau (total-tau protein), and P-Tau (phosphorylated-tau protein in position 181) measurements were performed using an enzyme-linked immunosorbent assay (ELISA) (Fujirebio) according to the manufacturer's instructions. The analysis of all biomarkers was performed in two runs and averaged for statistical analyses. The quality of the results was ensured by the use of validated standard operating procedures and internal quality controls (QC) as all centers were already involved in nationwide studies. The following cut-offs were used: $A\beta_{42}$ <550 pg/mL, Tau >350 pg/mL, P-Tau <60 pg/mL. When $A\beta_{40}$ was available, the normal $A\beta_{42/}A\beta_{40}$ ratio cut-off was >0.05.

Cerebral Imaging Data

When available, MRI scans were assessed using native DICOM format images. The presence of a cortical atrophy was evaluated; hippocampal atrophy was bilaterally scored following Scheltens visual rating scale [24]. Cortical functional imaging was assessed to identify focal hypometabolism using ^{18}FDG-PET or hypoperfusion using HMPAO-SPECT. Available DaTscan interpretations of trained nuclear physicians were used to confirm or identify dopaminergic denervation.

Amyloid and Tau PET imaging

Amyloid-β PET imaging of three patients was performed using [^{18}F]-florbetapir (n=1), [^{18}F]-florbetaben (n=1) or [^{18}F]-flutemetamol (n=1) as previously described [15]. Tau-PET examination was performed in two patients (and compared to 13 already available control subjects, mean age = 68.8 years [64-75]) on a High-Resolution Research Tomograph (HRRT; CTI/Siemens Molecular Imaging). The acquisition was performed from 80 to 100 minutes after intravenous administration of 382 ± 11.3 MBq of [^{18}F]-flortaucipir. Dynamic list mode acquisitions were binned into successive 5-minute time frames. Parametric images were created using BrainVisa software (http://brainvisa.info) by averaging the original 5-min images over

80-100 min after administration of flortaucipir. Standard Uptake Value ratio (SUVr) parametric images were obtained by dividing each voxel by the average value of the cerebellar grey matter region-of-interest with an erosion of 4 mm. Seventy-six cortical volumes of interest (VOI) were defined based on individual MRI scans and then applied onto the subject's PET space following coregistration. The VOIs, defined separately for the left and right hemispheres, were pooled into broader anatomical volumes of interest: we considered three cortical VOIs on the left and right sides respectively encompassing the temporal (with its parahippocampal, fusiform and polar sub-regions), parietal and frontal lobes.

Neuropathological examination

For each autopsy case, one hemisphere was fixed in formalin (fixation times between 8 and 27 months) and the contralateral hemisphere was frozen at -80°C, except for case EXT_2000_001 in which the whole brain was fixed in formalin. Immunostaining was performed with the Benchmark station automated system (Ventana Medical system Inc., Roche) using the following antibodies: anti-phospho-tau (pS202,pT205): clone AT8 (mouse monoclonal, ThermoFischer, 1/500); anti 3R-Tau (RD3): clone 8E6/C11 (mouse monoclonal, Merck, 1/100); anti 4R-Tau (RD4): clone1E1/A6 (mouse monoclonal, Merck, 1/50), anti phospho-tau (pT212,pS214): clone AT100 (mouse monoclonal, ThermoFischer, 1/500), anti p-62: clone 3/P62 lck ligand (mouse monoclonal, BD Biosciences, 1/500), anti-Aβ antibody: clone 6F/3D (mouse monoclonal, Agilent, 1/200); anti-TDP43 (rabbit polyclonal, Proteintech, 1/1,000); anti α-synuclein: clone 5G4 (mouse monoclonal, Millipore, 1/4,000). Except for immunohistochemistry with the Aβ and p62 antibodies, tissue was pretreated using a buffer at pH8 at 95°C between 8 minutes and 1 hour before adding the antibodies. The antibodies were incubated at 37°C between 32 minutes and 2 hours. The antigen retrieval used for Aβ was 5 minutes in formic acid and incubation of the antibody overnight at room temperature. For p62, the antigen retrieval was done in a buffer at pH 6 for 76 minutes and incubation of the primary antibody for 96 minutes at 37°C. Gallyas and Bielschowsky silver techniques were used to assess argyrophilic properties of tau aggregates [29, 30].

Aβ oligomers were extracted from frontal cortex following Lesné et al. procedures [16] and analyzed by Western blot using WO2 antibody (Covance, Princeton, NJ).

Molecular profiles of tau aggregates were analyzed from sarkosyl-insoluble fraction following the protocol described by Sahara and Kimura [22] for patients ALZ_441_005 and ALZ _596_006 using brain samples extracted from frontal cortex and striatum. A patient with Pick's disease (PiD), an AD patient and a control individual were processed in parallel. Sarkosyl-

insoluble fractions from patient EXT_1998_001 and a subject with progressive supranuclear palsy (PSP) were obtained following the protocol described by Dan et al [6]. Indeed, the protocol from Sahara and Kimura did not enable us to obtain sufficient amounts of sarkosyl-extractable aggregates for EXT_1998_001. Hence, we observed a slightly different staining for patient EXT_1998_001; notably, the 3R staining revealed several bands from lower molecular weight, which were not observed for the other patients. For the sake of comparison, the AD patient and the control individual were also extracted with this method. Sarkosyl-insoluble fractions were analyzed by Western blot using the following antibodies: anti Phospho-tau (pT181) antibody: AT270 (mouse monoclonal, ThermoFischer, 1/200), anti 3R-Tau (RD3): clone 8E6/C11 (mouse monoclonal, Merck, 1/1,000), anti 4R-Tau (RD4): clone1E1/A6 (mouse monoclonal, Merck, 1/200).

Quantification of the 4R/3R tau ratio from brain mRNA

The mRNA from frontal cortex and striatum of three patients carrying the *MAPT* duplication and 5 normal controls was extracted using RNA extraction kit (Macherey-Nagel). Reverse transcription was performed using verso cDNA kit (ThermoFisher Scientific). Then, cDNAs were PCR-amplified in a limited number of cycles, using a 6-FAM labelled F primer located on exon 9 (5'-CCCAAGTCGCCGTCTTCC-3') and a R primer located on exon 11 (5'-TGGTTTATGATGGATGTTGCCT-3'). This resulted in the amplification of 2 fragments of 207 and 300 bp, depending on the exclusion (3R-Tau) or inclusion (4R-Tau) of exon 10, respectively. After migration on an ABIprism 3500 automated sequencer (Applied Biosystems), the peak heights were quantified using the GeneMapper5 software in order to determine the 4R/3R Tau ratios.

In vitro seeding experiments

Preparation of human brain homogenates. In addition to brain tissues from ALZ_441_005, ALZ_596_006 and EXT_1998_001, three additional *post-mortem* brains, collected in a brain donation project and stored in the French national brain biobank (Bioressource Research Impact Factor number = BRIF BB-0033-00011), were used for this study: one 72-year-old male corresponding to a control case devoid of any Aβ, phospho-tau or α-synuclein accumulation, one 64-year-old female with progressive supranuclear palsy (PSP case) and one 61-year-old male with Pick's disease (PiD case). The following samples were chosen: striatum of the three *MAPT* duplication cases, pons of the PSP case (as example of a 4R tauopathy) and frontal cortex of the PiD case (as example of a 3R tauopathy). The striatum of the control case was also

studied. Approximately 150 mg of frozen tissue were thawed on ice and mixed with ice-cold phosphate-buffered saline (PBS) containing protease and phosphatase inhibitor cocktail (Thermo Fisher Scientific), to reach a final solution of 20% weight/volume. Tissue was homogenized by 15-20 strokes in a 1 mL glass homogenizer (Fisher Scientific), briefly sonicated on ice with a Misonix Q125 sonicator (Q-sonica) and clarified by centrifugation (3000 g, 5 min, 4°C), as previously described [27]. Supernatants were collected, aliquoted and stored at -80°C. Concentrations of tau proteins were assessed by total tau (Total) Human ELISA Kit (Invitrogen, Thermo Fisher Scientific).

Treatment of biosensor cells sensitive to tau seeding. The seeding activity of each homogenate was evaluated on a misfolded tau biosensor (HEK cells expressing tau RD P301S coupled with YFP or CFP fluorochromes, #CRL-3275, ATCC, LGC Standards). Those cells were plated in 24-well plates and treated in duplicate, following the original protocol adapted at the laboratory [9, 11], with a few adjustments. A transduction mix was prepared with 43.75 µL Opti-MEM (Gibco, Thermo Fisher Scientific) and 6.25 µL Lipofectamine 2000 (Invitrogen, Thermo Fisher Scientific), combined with 50 µl of the homogenate diluted in Opti-MEM and normalized by total tau. The transduction mix was briefly spun, incubated for 30 min at room temperature and added to the cells. Cells were left in the incubator during 24h. The experiment was replicated three times.

Analysis. Treated cells were re-plated on 13 mm diameter coverslips coated with poly-D-lysine (Sigma-Aldrich) and left 6-7h in the incubator. Cells were fixed in 4% PFA and counterstained with DAPI. The coverslips were mounted with Fluoromount (Sigma-Aldrich). Slides were stored in the dark at 4°C until observation with an inverted confocal Leica TCS SP8 DLS (Leica Microsystems). The YFP/CFP positive cells were manually counted from max intensity projection images (4 microscopic fields per coverslip, 8 per condition). The morphology profiles of YFP/CFP positive aggregates were assessed from stack images (50 ± 10 aggregates per condition).

RESULTS

In addition to the six 17q21.31 duplication carriers already reported by our group [15, 21], and the two reported in the Chen et al. paper [4], we have identified two unrelated novel cases carrying the same rearrangement, leading to a total of 10 cases from 9 families. In all cases, the size of the duplicated segment was identical and encompassed the entire *MAPT* coding sequence. As displayed in Table 1, these microduplications occurred in different H1/H2 haplotypic backgrounds.

Clinical features

Clinical phenotypes of all 10 carriers are summarized in Table 1. Mean age of onset was 51.3 years (range: [37-57]) with 6/10 being male patients. In 8 cases, an amnestic syndrome was the main clinical phenotype. Among them, 4 patients initially showed exclusive memory impairment, 3 presented with memory loss associated with either apathy or other behavioral disturbances and one first exhibited behavioral disorder and then memory impairment. The cognitive follow-up of seven patients including a verbal episodic memory assessment by the Free and Cued Selective Reminding Test, revealed a hippocampal dysfunction defined as an abnormal free recall without normalization of the cued retrieval [23]. On the remaining cognitive domains, behavioral and cognitive dysexecutive syndromes (following criteria available at ref. [10]) were identified for 6 patients. Accordingly, the verbal fluency was in abnormal ranges in all of them. None of them presented with visuoperceptual impairment. Neurological examination of these 8 patients was otherwise unremarkable at first visit. In particular, there was not any motor, sensitive, pyramidal tract or extrapyramidal signs. No vertical gaze palsy was noticed. All of them gradually declined towards a severe memory deficit with misorientation and gesture apraxia inducing a progressive impact on instrumental activities of their daily living.

Both cases with an initial akinetic-rigid syndrome developed gait disorders with falls associated with dysarthria, resting tremor and genito-urinary dysfunction. Symptoms were considered levodopa moderately sensitive in EXT_2000_001 and levodopa-resistant in EXT_1998_001. No vertical gaze palsy was reported for EXT_1998_001; it was noted only during the last year before death of EXT_2000_001. In this last patient, executive and attention deficits with judgment impairment and perseverative ideas were reported after a time course of five years (one year before death). Unfortunately, no detailed cognitive assessment was available for this patient. In patient EXT_1998_001, a memory impairment was present at first examination and progressively worsened, 4 years after onset of motor signs and 3 years before death (MMSE scored 23/30 at year 4 and MMSE scored 17/30 with complete temporal disorientation at year 5). Due to these unusual clinical presentations, main clinical hypotheses during the follow-up of both patients were first multiple system atrophy with predominant parkinsonism and then atypical progressive supranuclear palsy (PSP).

Among this group of *MAPT* duplication carriers, at the present time, five patients have died from their disease. Autopsies were available in 4 of them. Mean disease duration was 10.1 years [5-20] when considering the whole group of carriers and 12.0 years when restricting to deceased

patients.

CSF biomarkers

As can be seen in Table 1, CSF tau and p-tau values were in the abnormal ranges while CSF Aβ was normal in the two novel patients (EXT_1593_001 and EXT_1687_001). In one previously reported individual (ALZ_596_001) with an $A\beta_{42}$ value below the threshold, a second CSF biomarker assessment following a novel lumbar puncture (4 years after the first one) showed an $A\beta_{42}$ value in the normal range (646 pg/mL). Overall, CSF $A\beta_{42}$ values were considered in the normal ranges in 4/6 patients tested.

Brain imaging patterns (MRI, FDG-PET, DAT-SCAN)

MRI scans, performed between one and five years after disease onset, were available for eight patients. White matter hyperintensities on FLAIR-weighted images were present in 3/8 patients but were mild, punctiform and periventricular for ALZ_596_001 and EXT_1114_001 (with a medical history of vascular risk factors) or posterior, moderate and periventricular for EXT_1593_001 (who presented smoking addiction) (Fig. 1a). Cortical atrophy was present in all patients although differentially located. Internal temporal atrophy was found in all cognitive patients, associated with anterior temporal atrophy in three patients (ALZ_596_001, EXT_1687_001, ALZ_441_005) or parietal and occipital atrophy in another three (EXT_1593_001, EXT_1114_001 and ROU_1373_001) (Fig. 1a to 1d). Conversely, no cortical atrophy was noticed for patients EXT_1998_001 and EXT_2000_001.

Functional imaging using FDG-PET or HMPAO-SPECT, showed a pattern consistent with the atrophy profiles on MRI. Tracer uptake was decreased in the temporal cortex of all five analyzed patients. The uptake was additionally decreased in the parietal regions of three patients: ROU_1373_001 (Fig. 1e), EXT_1593_001 and ALZ_ 441_005. Interestingly, that last patient also showed a severe frontal right hypometabolism.

DaTscans were performed in four patients. Only one of them (EXT_2000_001) presented with an extrapyramidal syndrome. The three other patients had no extrapyramidal syndrome, neither at first visit, nor at follow up. Strikingly, all four patients showed significantly decreased uptake of the tracer, suggesting bilateral nigro-striatal dopaminergic neuronal loss, either symmetrical for ROU_1373_001 and EXT_2000_001, or asymmetrical for EXT 1593_001 and EXT_1687_001 (left predominance for both).

Molecular imaging

Amyloid PET was available in three patients (already reported in [15]) including in patient ALZ_596_001 with initially low and then normal CSF $A\beta_{42}$ values, and in patient EXT_1114_001 with abnormal $A\beta_{42}$ values. Amyloid PET scans were all considered as normal (Table 1).

Tau-PET imaging using [^{18}F]-flortaucipir was performed in two patients. On visual inspection, we found that the tracer binding, eight years after disease onset, was strongly increased in the temporal lobes and in the posterior and superior region of the frontal lobes of ALZ_596_001 (Fig. 1f). The other patient (ROU_1373_001) had less pronounced [^{18}F]-flortaucipir binding on the imaging performed nine years after onset. The uptake was restricted to the medial and anterior temporal regions. A control group of 13 individuals was used to calculate the mean and the standard deviation (SD) of the SUVRs in the regions of interest. The SUVRs of ALZ 596_001 were above mean plus 1.96 SD of controls (95% confidence interval), in the temporal lobes (parahippocampal, fusiform, inferior temporal and temporo-polar regions), as well as in the supplementary motor areas. In patient ROU 1373_001, only the parahippocampal gyri and right temporal pole were above mean plus 1.96 SD (Fig. 1g).

Neuropathological findings

Neuropathological findings are summarized in Fig. 2, Fig. 3, Table 2, supplementary Fig. 1, supplementary Fig. 2, supplementary Fig. 3, Table 2 and supplementary Table 1.

ALZ_441_005

At autopsy, the brain was extremely atrophic, the left hemisphere weighted 347 g after formalin fixation. Macroscopic examination of the left hemisphere revealed a severe atrophy of the frontal, temporal and parietal lobes, mostly affecting the association cortices. The striatum was also atrophic, in particular the caudate nucleus. The hippocampus and amygdala were barely identifiable (Supplementary Fig.1a, b). The brainstem also showed a generalized atrophy. There was a loss of pigment of the *substantia nigra* and the *locus cæruleus* was not identifiable. Finally, in the cerebellum, in addition to a chronic cavitary infarction of the left hemisphere, there was an atrophy of the dentate nucleus.

Histology revealed massive neuronal loss, severe astrogliosis and marked spongiosis in the regions where atrophy was seen at macroscopic examination. These neurodegenerative changes were associated with severe degeneration of the adjacent subcortical white matter.

Immunohistochemistry using the AT8 anti-tau antibody revealed a great number of hyperphosphorylated tau deposits in neurons, astrocytes and less frequently in oligodendrocytes

as well as in the neuropil. Tau positive aggregates were numerous and seen predominantly in the neocortex, striatum, pallidum, limbic regions, including the hippocampus and entorhinal cortex (Fig. 2a, b, Fig. 3) and the cerebellum. Neocortical pathology involved the frontal, temporal and parietal lobes with very little tau pathology in the occipital cortex. Tau pathology predominated in the grey matter and was scant to moderate in the white matter. In the brainstem, AT8-positive aggregates were numerous in the midbrain and the pons but not in the medulla oblongata. In neurons, the majority of the deposits were spherical inclusions reminiscent of Pick bodies (Fig. 2a). Although the aggregates were mainly 3R-tau, a significant number of inclusions were also 4R-tau immunoreactive. They did not show argyrophilic properties with the Bielschowsky silver staining, except for a few neurons in the dentate gyrus and CA1 regions of the hippocampus, but were positive for Gallyas silver staining (suppl table 1). Neurons with diffuse cytoplasmic neuronal immunoreactivity or globose neurofibrillary tangles (NFTs) were also observed but in a much lesser number. In the dentate nucleus of the cerebellum, coarse grain intracytoplasmic neuronal inclusions predominated (Supplementary Fig. 2). The astrocytic deposits reminded of ramified astrocytes as in Pick's disease (Fig. 2a). They showed tau positivity close to the nucleus and lacked fine prolongations in the astrocytic processes. Of note was the predominance of neuronal intracytoplasmic spherical inclusions and neuropil threads (NTs) with very few glial lesions seen in the striatum (Fig. 2b). Aggregates in oligodendrocytes were not numerous and they were predominant in the white matter of affected regions. Small round spherical inclusions, smaller than the nuclei, similar to those seen in Pick disease, were more abundant than coiled bodies (Supplementary Fig. 2 and 3).

Tau aggregates in neurons and astrocytes were labeled with 3R-tau and 4R-tau whereas oligodendrocytes were labelled with 3R-tau and less frequently with 4R-tau. As can be seen in Table 2 summarizing the presence of 3R and 4R-tau deposits among nine brain regions in each available autopsy case, there was a predominance of 3R-tau isoforms over 4R isoforms in neurons and astrocytes of all examined brain regions. It is noteworthy that the striatal neuronal deposits were strictly labeled by 3R but not 4R-tau (Fig. 2b). Neuronal, astroglial and oligodendrocytic aggregates were immunoreactive for p62 and AT100. Argentophilic properties were variable depending on the region. A few neuronal globular deposits showed argentophilia with Bielschowsky staining in the dentate gyrus of the hippocampus while they were negative for Gallyas, as seen in Pick disease (Supplementary Fig. 3c). However, in the frontal cortex and striatum they were positive for Gallyas and negative for Bielschowsky silver staining. Small globular oligodendroglial deposits in the white matter showed argyrophilic properties both with the Gallyas and Bielschowsky stainings (Supplementary Fig. 3b).

Sarkosyl-insoluble fractions from the frontal cortex and striatum blotted with phospho-tau antibody revealed two main bands at 55 and 64 kDa, similar to the pattern observed in a PiD case processed in parallel. Consistently, when blotted with a 3R-tau antibody, these western blots showed intense immunoreactivity whereas no immunoreactivity was detected with the 4R-tau specific antibody (Fig. 4a).

Overall, the morphology of tau aggregates was reminiscent of that observed in PiD. However, immunohistochemical and argyrophilic properties of the aggregates showed some differences and there was abundant pathology in the dentate nucleus of the cerebellum, which is not usually observed in PiD. Furthermore, white matter pathology was scant and the globular oligodendroglial aggregates were of small size compared to the ones seen in globular glial tauopathy.

ALZ_596_006

The fixed left hemisphere weighted 587 g. Macroscopic examination revealed a moderate atrophy of the medial region of the temporal lobe associated to a moderate ventricular dilatation (Supplementary Fig. 1c).

Histology showed marked neuronal loss associated with reactive gliosis in the amygdala, the hippocampus and the entorhinal cortex. A mild to moderate neuronal loss was seen in the nucleus basalis of Meynert, the *substantia nigra* and the *locus cæruleus*. The cerebellum appeared intact at hematoxylin and eosin (H&E) examination.

Immunohistochemistry revealed marked tau pathology in the neocortex where there was a predominance of tau positive astrocytes, with moderate number of NFTs and NTs (Fig. 2a). Tau deposits in the astrocytes had various morphologies: some accumulated tau near the nucleus, others had fine deposits in the distal part of the processes and in others the deposits were seen along the length of the processes. In the striatum, the number of tau aggregates was moderate and involved the neurons with the presence of many NFTs and NTs (Fig. 2b). Very few tau positive astrocytes were seen in the striatum (Fig. 3). Tau pathology in the pallidum was rare (Supplementary Fig. 2). In the brainstem, tau pathology was abundant in the *substantia nigra*, which showed many globose NFTs and NTs (Supplementary Fig. 2). In the pons, tau deposits were observed mainly in the tegmentum, including the *locus cæruleus*. Both neuronal and astrocytic deposits were seen. In the medulla oblongata, tau pathology predominated in the reticular formation. The cerebellum had very few aggregates in the dentate nucleus (Supplementary Fig. 2). Oligodendrocytes were only observed in the reticular formation of the medulla and the amygdala. Subependymal, perivascular and subpial thorny astrocytes were

seen in the medial region of the temporal lobe and striatum suggesting the presence of associated aging-related tau astrogliopathy (ARTAG) (Supplementary figure 3). A few clusters of astrocytes in the white matter of the frontal, parietal and occipital lobe were also seen. Tau pathology in the medial temporal lobe with numerous NFTs and NTs in the hippocampus, the entorhinal cortex and the amygdala were also observed (Supplementary figure 3). Occasional diffuse Aβ plaques were seen in the cortex. All of the deposits were positive for p62 and AT100. Neuronal aggregates were immunoreactive for 3R-tau and 4R-tau. In contrast, astrocytes where only 4R-tau positive. Silver staining was variable. Neurons and astrocytes where positive for Gallyas silver staining and only occasional neuronal aggregates and rarely astrocytes where argyrophilic with Bielschowsky (suppl figure 3). As can be seen in Table 2 and Fig. 2, when analyzed for tau isoforms, neuronal aggregates were 3R- and 4R-tau positive with a predominant 3R labelling in almost all structures examined. As in case ALZ_441_005 the striatum was only immunoreactive for 3R-tau. Western blot of insoluble fractions from frontal cortex and striatum, labeled with AT270 antibody showed three bands similar to those seen in AD. The two lower bands were intensively labelled by 3R-tau while the upper and middle bands were labelled by 4R-tau (Fig. 4b).

Overall, this case shows a tauopathy (3R>4R) that mainly involves neurons and astrocytes, in cortical and subcortical regions, in which tau aggregates adopt diverse morphologies. Distribution and biochemical properties of the lesions did not fulfill the criteria for either of the known tauopathies.

EXT_1998_001

The right hemisphere fixed in formalin weighted 391g (brainstem and cerebellum not included). Gross examination of the coronal sections did not show atrophy of the brain parenchyma (Supplementary Fig. 1d). The study of the mesencephalon was limited, as the brainstem had been previously separated from the hemisphere through this region. However, the rest of the brainstem appeared to have a normal size at macroscopic examination. The *locus cæruleus* was not identified.

Histology showed, in addition to widespread anoxic lesions, a tauopathy involving the brainstem, pallidum, striatum, thalamus and dentate nucleus of the cerebellum. In the brainstem, the pallidum and the dentate nucleus, the most prominent AT8 positive tau inclusions were globular bodies, diffuse neuronal intracytoplasmic granular immunoreactivity and NTs (Supplementary Fig. 2). Coiled bodies were seen in the pallidum. Tufted astrocytes were the only tau pathology seen in the striatum (Fig. 2b). In the frontal cortex, the pathology was limited

but the extensive laminar necrosis and massive neuronal loss could have explained the scarcity of tau deposits which were predominantly found in tufted astrocytes (Fig. 2a). A few NTs and coiled bodies in the subjacent white matter were seen. In the medial and lateral regions of the temporal lobe, tau pathology which included NFTs, granular intracytoplasmic aggregates, NTs and ghost tangles was severe. Immunohistochemistry for tau antibodies in the frontal cortex and striatum revealed that tau pathology was only of the 4R isoforms, while 3R-tau immunohistochemistry was negative (Fig. 2). Neuronal and astrocytic aggregates were immunoreactive for p62 and AT100. Oligodendrocytes showed frequent immunoreactivity for AT100 but it was rare for p62. Silver staining showed positivity for Gallyas while Bielschowsky was negative (Supplementary Table 1). In medial temporal brain regions, the deposits, consistent with Braak stage II-III tau pathology, were labeled with both 3R and 4R-tau (Table 2).

Sarkosyl-insoluble fraction from the frontal cortex and striatum blotted with AT270 antibody showed a two-band profile reminiscent of PSP. When blotted with a 3R-tau antibody, samples from frontal cortex did not show any immunoreactivity, while samples extracted from the striatum showed a faint, 2-band pattern, corresponding to 55 and 64kDa Tau bands. Both samples were labeled by the 4R-tau antibody, with a similar pattern to that seen with the AT270 antibody, confirming that tau aggregated, in these regions, was mainly 4R (Fig. 4c).

EXT_2000_001

The formalin fixed whole brain weighted 1200 g. At gross examination, the *substantia nigra* was pale and the subthalamic nucleus, atrophic. There was a moderate dilation of the 3rd ventricle (Supplementary Fig. 1e). Histologic examination with hematoxylin-eosin (H-E) evidenced, in accordance with the macroscopic observations, a massive neuronal loss in the *substantia nigra*. The neuronal loss was severe in the subthalamic nucleus. The pallidum was gliotic. The neocortex and the hippocampus appeared normal.

Tau positive aggregates were mainly detected in subcortical regions (especially in the basal ganglia and in the brainstem) and were minimal in the frontal cortex and hippocampus (Fig. 2a, b; Supplementary Fig. 1). The predominant lesions were NFTs, sometimes globose, NTs and astrocytic deposits. In the astrocytes, AT8 tau positive aggregates were atypical, close to the nuclei with little ramifications. In the caudate nucleus, however, the morphology of the astrocytic aggregates was similar to that of tufted astrocytes (Fig. 2a, b). Coiled bodies in the oligodendrocytes were infrequent. In the striatum, aggregates were seen in the caudate nucleus while they were scarce in the putamen. The internal capsule was the only region that showed

many coiled bodies and NTs were also frequent. Tau pathology was marked in the pallidum (Supplementary Fig. 2). While the thalamus was moderately involved the subthalamic nucleus had massive pathology with a predominance of neuronal deposits (NFTs and NTs). Mainly globose NFTs were seen in the nucleus basalis of Meynert. In the brainstem there was a large number of deposits in the *substantia nigra* and the periaqueductal grey of the mesencephalon. In the pons, tau aggregates were numerous in the tegmentum while they were rare in the ventral pons (Supplementary Fig. 2). In the medulla oblongata, the aggregates predominated in the reticular formation, in the cranial nerve nuclei and in the inferior olive, where mainly NFTs were seen. In the cerebellum the presence of many coarsed granular neuronal intracytoplasmic aggregates and NTs in the dentate nucleus was surprising compared to its normal appearance after H-E staining. All of the tau deposits seen were positive for the 4R isoforms and negative for the 3R isoforms (Table 2, Fig. 2). In the entorhinal cortex tau aggregates expressed both 3R and 4R isoforms, consistent with Braak stage II tau pathology. As with the previous case, all of the tau aggregates were positive for AT100. P62 immunoreactivity was also seen in all types of aggregates, however it was scant compared to the amount of tau pathology seen. Similarly to the previous case, tau aggregates were positive for Gallyas and negative for Bielschowsky silver staining. No Western blot analysis was undertaken due to the lack of frozen tissue.
Overall, the distribution and characteristics of tau pathology in cases EXT_1998_001 and EXT_2000_001 was reminsicent of that observed in progressive supranuclear palsy (PSP) cases. However, compared to sporadic PSP, in our two cases, oligodendrocytes with tau pathology (coiled bodies) were scant and tau aggregates in the pontine basis were rare.

No evidence for associated proteinopathies

There was no associated α-synuclein or TDP-43 proteinopathies in any of the cases (Supplementary Fig. 7). No Aβ deposits were observed except in case ALZ_596_006 where rare diffuse Aβ plaques were seen in the cortex of the middle frontal gyrus, the subcallosal area and the inferior temporal gyrus. No Aβ oligomers were detected by western blot analysis of the frontal cortex of the three cases with available frozen tissue (Supplementary Fig. 4)

Analysis of exon 10 splicing and determination of 4R/3R tau mRNA ratios

The alternative splicing of exon 10 in mature *MAPT* mRNA naturally leads to either 3R-tau (in case of exon 10 skipping) or 4R-tau (in case of exon 10 inclusion). In an attempt to identify genetic variants influencing alternative splicing of exon 10, we sequenced exon 10 boundaries in the nine cases with available DNA. No single nucleotide variant, short insertion or deletion

putatively influencing splicing was found.

MAPT 4R/3R mRNA ratios were analyzed in two cerebral regions (frontal cortex and striatum) for the three patients for whom frozen tissue was available (EXT_1998_001, ALZ_596_006 and ALZ_441_005) and for five normal controls. In controls, the 4R/3R mean ratio was 0.69 ± 0.10 (SEM) in the frontal cortex and 0.97 ± 0.16 (SEM) in the striatum. For each control, the 4R/3R ratio was systematically increased in the frontal cortex compared to striatum, but this increase was collectively not significant (p=0.125, Wilcoxon signed ranked test). In the frontal cortex of the patient with 3R-tau aggregates resembling Pick bodies (ALZ_441_005), we noted a 243% increase of the 4R/3R mRNA ratio as compared to the mean value in controls, which is in the same range as published data in patients with Pick disease [13]. In the striatum, 4R/3R mRNA ratios were higher in both patients with mainly 3R-tau aggregates (ALZ_441_005 and ALZ_596_006) (136% and 139 % increase, respectively) and lower (74%) for the patient with mainly 4R-tau aggregates (EXT_1998_001) as compared with the controls mean value (Supplementary Fig. 5).

In vitro seeding experiments

All homogenates derived from samples containing misfolded tau induced the aggregation of the endogenous tau RD P301S proteins expressed by the biosensor (Fig. 5; Supplementary Fig. 6). The intracellular aggregates of tau proteins observed in biosensor cells was of different types: some aggregates were granular (types 1 and 2), while others appeared larger and denser (types 3 and 4; Fig. 5a). Morphological features of induced aggregates of three *MAPT* duplication carriers (ALZ_441_005, ALZ_596_006 and EXT_1998_001) showed dense/granular ratios between those induced by homogenates of a PiD and a PSP control (Fig 5b), at both extremes. Indeed, the seeding-competent homogenates derived from ALZ_441_005 and PiD cases led to a majority of granular aggregates of tau proteins, that represented respectively 66.44% [95%CI: 58.86;74.02] and 67.35% [95%CI: 59.77;74.93] of total aggregates (Z-test comparison to the null hypothesis of equal proportions: $p < 0.0001$ for ALZ 441_005 and PiD cases, as shown in Fig. 5b). Interestingly, dense/granular ratios did not differ significantly between ALZ_441_005 and the PiD case (Fig 5c). Conversely, samples obtained from EXT_1998_001 and PSP cases both induced a majority of dense aggregates, reaching a proportion of 63.83% [95%CI: 55.90;71.76] and 76.97% [95%CI: 70.28;83.67], Z-test comparison to the null hypothesis of equal proportions: p=0.0014 for EXT_1998_001 case and $p < 0.0001$ for PSP case, as shown in Fig. 5b). Finally, the samples obtained from ALZ_596_006

showed an intermediate proportion of dense/granular aggregates (proportion of dense aggregates: 56.21% [95%CI: 48.73;63.69]; Z-test comparison to the null hypothesis of equal proportions: p =0.1239).

DISCUSSION

The biological, imaging and neuropathological data presented here definitely settle one issue: the 17q21.31 duplication causes a primary tauopathy showing various combinations of well-characterized primary tauopathies, and not an atypical form of AD. Previous CSF biomarker data suggesting an Aβ pathology in 3/4 *MAPT* duplication carriers were not supported by a reassessment in one patient and more recent data obtained in two additional patients in whom Aβ levels were in the normal range. Furthermore, all three amyloid PET scans were negative; there were no senile plaques at neuropathological examination and minimal Aβ staining was present in only one case. We further ensured that no Aβ oligomers were detected, ruling out any involvement of this peptide in the pathology. Of note, the case reported by Alexander et al. [1] showed asymmetric TDP-43 deposits. We hypothesize that it could be co-incidental in this case, as none of our patients showed such immuno-reactivity, even though only one hemisphere was examined in all four cases.

Clinically, we identified eight patients presenting with episodic memory impairment following a profile observed in typical AD [8] with or without behavioral symptoms upon presentation, on one side, and one patient with an isolated atypical extrapyramidal presentation resembling the one observed in progressive supranuclear palsy (PSP), on the other side. These clinical presentations appear as two extremes, belonging to a spectrum. That hypothesis is supported by (i) the association, in the remaining case with an atypical extrapyramidal syndrome upon presentation, of an early memory impairment that worsened during follow up and (ii) the fact that three cases with memory impairment upon presentation also exhibited significant dopaminergic nigrostriatal neuronal depletion as observed on DaTscan imaging despite normal physical examination, thus suggesting partial alteration of dopaminergic pathways however insufficient to induce the emergence of extrapyramidal signs as described in premotor Parkinson's disease stages [25]. Interestingly, in the family described by Alexander et al.[1], both the proband and his sister presented memory and behavioral impairment, with neuropsychological assessments and MRI consistent with a diagnosis of probable AD, but also exhibited mild to moderate extrapyramidal symptoms upon examination.

Neuropathological examination combining immuno-histochemical and biochemical analyses supports such a continuum by showing that different tau isoforms formed aggregates among

the four examined brains. In one case with a cognitive and behavioral disorder presentation (ALZ_441_005), we observed widespread tau deposits in the hippocampus and neocortex, which were mainly composed of 3R-tau as in Pick disease although globular inclusions were Gallyas positive and Bielschowsky negative in the striatum and frontal cortex. Thus, this case harbors a novel variety of Pick body-like inclusions in addition to those already associated with some *MAPT* pathogenic variants [2, 19, 26]. In contrast, in the case presenting with isolated atypical extrapyramidal syndrome (EXT_2000_001), the picture was reversed: we observed tau aggregates mainly located in subcortical regions, which were exclusively composed of 4R-tau protein isoforms. Those two cases may be seen as the extremes of the spectrum: between those two extremes, patient EXT_1998_001, who exhibited an akinetic-rigid syndrome associated with a severe memory deficit, showed a mixed 3R/4R tauopathy, albeit with 4R-tau predominant deposits in the striatum and frontal cortex. The clinical picture, the 4R-tau predominance, the presence of 3R-tau accumulation in the subcortical nuclei is reminiscent of the case published by Alexander et al. [1]. In the last case (ALZ_596_006), closer to the dementia side, a mixed 3R/4R tauopathy similar to that seen in AD was found with severe involvement of the medial region of the temporal lobe.

Overall, *MAPT* duplication can thus lead to a spectrum of primary tauopathies with 3R- or 4R-tau predominant aggregates. Our observations suggest that 4R-tau deposits involve together basal nuclei, brainstem and basal ganglia, and are associated with motor symptoms. On the other hand, 3R-tau predominant cortical inclusions were associated with Pick-like inclusions. Those conclusions, based on the analysis of a small number of cases, obviously await confirmation by further studies.

Of note, *in vivo* tau PET imaging could reflect neuropathological heterogeneity of the tau pathology. Postmortem studies showed that the affinity of the tau tracer for non-AD tau lesions may be lower than that for AD tauopathy, with low or absent binding of [^{18}F]-Flortaucipir to tau straight filaments [17]. The distinct tau PET binding profiles between ALZ_596_006, and ROU_1373_001 could reflect a qualitative rather than a quantitative difference of the tau pathology, with a possible different mixed 3R/4R tauopathy in ALZ_596_001, whose tau tracer uptake was clearly increased in the temporal lobes. The tau tracer binding was far less pronounced in ROU_1373_001, who had nevertheless a similar clinical presentation, disease duration, and FDG-PET pattern. This could reflect a different type of tau lesions for which the tracer has less affinity.

We sought to identify a putative correlation between the preferential accumulation of 4R- or 3R-tau and the relative expression of tau transcripts. Focusing on the striatum where either 4R

or 3R-predominant tau aggregates were present in patients, we failed to find a positive correlation with 4R/3R mRNA ratios. Rather, the relative proportion of 4R-containing transcripts was moderately increased in patients ALZ_441_005 and ALZ_596_006 with 3R-tau aggregates whereas the converse was observed in patient EXT_1998_001 with 4R deposits. Consistent with previous findings in PSP and PiD patients [13], the preferential accumulation of 4R- or 3R- tau aggregates is neither driven by the corresponding preferential expression of the 4R or 3R-encoding transcripts, nor by the *MAPT* haplotype associated with the duplication. It has been suggested, although this claim remains controversial [28], that the *MAPT* H1 haplotype [3] or sub-haplotypes [18], is associated with increased expression of 4R tau-encoding transcripts. Accumulation of 3R-tau in ALZ_441_005 occurred in the 2H1-1H2 background whereas accumulation of 4R-tau in EXT_1998_001 occurred in the 1H1-2H2 background: only one H1 haplotype was thus present in a case with 4R-tau predominant accumulation while they were two in a case in which 3R-tau was the preferential isoform, in contradiction with the hypothetical role of the H1 haplotype. Finally, the search of rare or common variants in exon 10 and surrounding splicing regions, which could have influenced the 3R/4R tau mRNA ratios remained negative. We conclude that the shift toward 3R or 4R-tau predominant pathology and the topography of lesions do not appear to be directly related to a change in the balance between the corresponding transcripts.

Local factors influencing availability or aggregation properties of different tau isoforms might explain why, in the general context of tau overexpression, aggregates are preferentially formed of 3R- or 4R-tau in discrete brain regions of a same individual. The striking inter-individual differences in the composition of aggregates observed in identical brain regions suggest, however, that this process could also be driven by the formation of different pathogenic seeds: peculiar sets of 3R or 4R misfolded tau proteins could aggregate in specific parts of the brain and subsequently spread in other cerebral regions along anatomically connected networks [5]. Tau pathology located in the striatum of ALZ_441_005, ALZ_596_006 and EXT_1998_001 cases induced different types of tau aggregates in a biosensor cell line expressing tau RD P301S (4R-tau): the striatum of ALZ_441_005 containing 3R-tau inclusions mainly led to granular aggregates (similarly to a PiD homogenate), whereas the striatum of EXT_1998_001 presenting 4R-tau inclusions mostly resulted in dense aggregates (as observed with a PSP homogenate). The striatum of ALZ_596_006, containing a mixture of 3R- and 4R-tau lesions, seeded the formation of granular and dense aggregates in similar proportions. Those morphological features were independent of the sample seeding potential, as tau seeding activity appeared higher in ALZ_441_005 and PSP homogenates than in EXT_1998_001 and PiD samples, after

a total tau normalization (Supplementary Fig. 6). Our results thus support the hypothesis of different seeds among *MAPT* duplication cases. In the context of moderate *MAPT* overexpression related to the duplication, tau monomers could adopt different abnormal conformations leading to several seed competent species. Depending on the "strains", mainly 3R or 4R-tau could be preferentially incorporated, leading to various protein assemblies, resulting ultimately in different pathologies characterized by fibril morphology, cellular type specificity and anatomical spreading patterns. Also supporting this hypothesis, it has recently been shown that stochastic misfolding of tau conformers occurs in brain tissues of patients carrying the same P301L *MAPT* mutation, followed by templated conversion of native monomers [7].

In summary, we have identified different types of *MAPT* duplication carriers exhibiting, at least in early stages of their diseases, different cognitive or motor symptoms related either to cortical or subcortical dysfunctions and characterized by the accumulation of different tau isoforms. Given the diversity of clinical and imaging presentations and the simplicity of the genetic test, we suggest searching *MAPT* duplications in all cases with early-onset idiopathic neurodegenerative dementia or early-onset idiopathic atypical extrapyramidal syndromes. They should also be sought when neuropathologically-identified primary tauopathies are not related to *MAPT* mutations, particularly when the unusual constellation of tau pathology cannot be clearly put in one of the boxes of currently described primary tauopathies.

FIGURE LEGENDS

Figure 1. Structural, functional and Tau PET Cerebral imaging of MAPT duplication carriers

a. FLAIR-weighted axial MRI scan of EXT_1593_001 showing parietal cortical atrophy and mild white matter hyperintensities

b. T1-weighted coronal MRI scan of EXT_1114_001

c. T1-weighted coronal MRI scan of ROU_1373_001

d. FLAIR-weighted sequence coronal MRI scan of EXT_1687_001

e. FDG-PET scan of ROU_1373_001 showing parietal bilateral hypometabolism (arrows)

f. Tau-PET imaging of ALZ_596_001 with color scale corresponding to the SUVR scale

g. Tau-PET imaging of ROU_1373_001 with color scale corresponding to the SUVR scale

Figure 2. Tau pathology of the four cases carrying an *MAPT* duplication in the frontal cerebral cortex (A) and the caudate nucleus (B).

a. Diversity of tau pathology and 3R- and 4R-tau isoform expression in the frontal cortex. In ALZ_441_005, pathology was abundant with a predominance of neuronal deposits with many globular Pick-like bodies in the neurons. Astrocytic deposits were also numerous and reminded of tufted astrocytes with deposits close to the nucleus and without the ramifications. In ALZ_596_006, there was a predominance of astrocytic pathology with many tufted astrocytes and some neurofibrillary tangles (NFTs) in neurons. Tau pathology in EXT_1998_001 and EXT_2000_001 was scant and only a few deposits in the form of tufted astrocytes and a few NFTs in neurons were seen. Tau aggregates in ALZ_441_005 and ALZ_596_006 included the 3R- and 4R-tau isoforms. EXT_1998_001 and EXT_2000_001 tau aggregates expressed solely 4R-tau. *Frontal cortex. Immunohistochemistry: AT8, 3R-tau, 4R-tau. Scale bar in first row 100µm, Scale bar in four last rows 25µm*.

b. Diversity of tau pathology and 3R- and 4R-tau isoform expression in the caudate nucleus. In ALZ_441_005, pathology was abundant with a predominance of neuronal pathology with many globular Pick-like bodies in neuronal soma and abundant neuropil threads. In ALZ_596_006 neuropil threads were many, however neuronal neurofibrillary tangles (NFT) were moderate in number. In EXT_1998_001 and EXT_2000_001, pathology was predominantly astroglial with astrocytic deposits resembling tuffted astrocytes. ALZ_441_005 and ALZ_596_006 expressed only 3R-tau isoforms and EXT_1998_001 and EXT_2000_001 were positive only for 4R-tau immmunohistochemistry. *Caudate nucleus. Immunohistochemistry: AT8, 3R-tau, 4R-tau. Scale bar in first row 100µm, Scale bar in last*

three rows 25µm.

Figure 3. Distribution and intensity of tau pathology in the brain of *MAPT* duplication carriers.

Color-coded table following a quantitative evaluation of tau pathology by regions of the brain. Semiquantification of total tau pathology **(total)** in **neurons, astrocytes, oligodendrocytes** and neuropil **(neuropil threads)** in relevant brain regions. Blue=0; Green=rare or scant; Orange=moderate; Red=abundant. NA: not available. (a) Tau pathology is seen in the anterior part of the putamen in case ALZ_596_006 and EXT_1998_001 (b) Tau pathology only involves the external pallidum in case ALZ_441_005. ctx: cortex; wm: white matter; CA: cornu Ammonis; DG: dentate gyrus; Sub: subiculum; Subth: subthalamic nucleus; SN substantia nigra; PGM: periventricular gray matter; RN: red nucleus; PT: pontine tegmentum; BP: basis pontis; RT reticular formation; inf olive: inferior olive; dentate: dentate nucleus

Figure 4. Western blots of sarkosyl-insoluble 4R and 3R-Tau in patients and controls.

a. Analysis of patient ALZ_441_005 (ALZ 441) ; b. Analysis of patient ALZ_596_006. c. Analysis of patient EXT_1998_001. For all patients, western blotting was performed on brain samples extracted from frontal cortex (left panels) or striatum (right panels). Blots were revealed with antibodies against Tau phosphorylated on Thr181 (AT270 tracks), 3R-Tau (RD3 tracks) or 4R-Tau (RD4 tracks). Tau profiles obtained from the *MAPT* duplication carriers were compared to those of patients showing Alzheimer's disease (AD), Pick's disease (PiD) or PSP neuropathology, and to a control individual (Ctrl).

Note that western blots are presented here as a composite image from different times of exposure of a unique blot for each panel (all samples from each panel migrated on a same blot) to better compare Tau profiles between each antibody despite varying staining intensity.

Note that RD4 staining requires longer exposure than using RD3 antibody, so that minor 4R aggregates as observed in IHC may be hardly detectable in WB performed in similar regions.

Figure 5. *In vitro* characterization of tau seeding activity in the striatum of *MAPT* duplication cases in a cell biosensor.

HEK cells expressing tau RD P301S coupled with fluorochromes (YFP or CFP) were exposed to brain homogenates normalized for total tau proteins, obtained from the striatum of a control case and of three *MAPT* duplication cases, pons of a PSP case and frontal cortex of a PiD case. a. Illustration of the four types of tau aggregates. Type 1: unique and small grain (~ 10% of cell

volume or less); Type 2: multiple granules of homogeneous size; Type 3: multiple granules associated with a larger dense core; Type 4: large and dense aggregate (~ 30% of cell volume or more). Representative confocal images with max YFP (yellow), CFP (cyan) and DAPI (blue) intensity projections from a stack covering all the cells contained in the field (scale bar: 10 µm); no aggregates: example of cells devoid of any aggregates in HEK cells exposed to control brain homogenate, showing diffuse YFP/CFP positive immunofluorescent staining due to non-aggregated tau-RD b. Mean percentages of each type of tau aggregates observed for each case, obtained from three independent experiments in which a total of 50 ± 10 YFP/CFP positive cells were analyzed per condition. Proportions of "granular" (type 1 + type 2) and "dense" types of aggregates (type 3 + type 4) were compared to the null hypothesis of equal proportion using a Z-test for proportions. c. Proportion of cells with dense aggregates. Comparison between two individuals using a Z-test for proportions.

Supplementary Figures

Supplementary Fig. 1. Morphological variability at macroscopic examination between the four postmortem studied cases: ALZ_441_005 (A and B), ALZ_595_006 (C), EXT_1198_001 (D), EXT_2001_001 (E).

a. Marked atrophy of the frontal, parietal and temporal association cortices. b. Coronal sections at two levels, the head of the caudate and the mamillary bodies, showed marked dilatation of the lateral ventricle, severe atrophy of the striatum (white arrow) and of the temporal lobe (black arrow). The hippocampus and amygdala are barely identifiable. c.) Moderate atrophy of the medial temporal lobe (red arrow) associated to a moderate dilatation of the temporal horn of the lateral ventricle. d and e) Absence of brain atrophy. Moderate dilatation of the 3rd ventricle in EXT_2000_001 (e). (a and b) left hemisphere; (c) right hemisphere.

Supplementary Fig. 2. Diversity of tau pathology in different regions of the brain. In the pallidum tau pathology was abundant in all cases except in ALZ_596_006. In the *substantia nigra* tau aggregates where numerous in all cases seen where tissue was available. Globular tau aggregates in neuronal cell body and neuropil threads were the predominating deposits. In the pons, the *basis pontis* showed a great number of tau positive aggregates which were globular in ALZ_441_005. Tau deposits were scant in EXT_2000_001 and were absent in ALZ_596_006 and EXT_1998_001. The dentate nucleus showed abundant coarse granular neuronal intracytoplasmic aggregates in ALZ_441_005, deposits were moderate in EXT_2000_001 and scant in the other two cases. The white matter showed little pathology in ALZ_596_006 and EXT_2000_001 with few neuropil threads. In ALZ_441_005 a moderate number of oligodendrocytic aggregates were seen and were predominantly globular. In EXT_1998_001 the few oligodendroglial pathology seen adopted a more typical coiled body shape. Immunohistochemical staining with AT8. *Scale bar in rows 1-4: 50µm, Scale bar in last row: 25µm.*

Supplementary Fig. 3. a. Immunohistochemical and histochemical characteristics of tau aggregates. Tau aggregates seen in neurons and astrocytes in the four histological studied cases (ALZ_441_005, ALZ_596_006, EXT_1998_001, EXT_2000_001**)** were positive for phosphorylation dependent AT100 antibody, p62 and Gallyas silver staining. Occasional neurons and rarely astrocytes in the frontal cortex of case ALZ_596_006 showed argyrophilic properties with the Bielschowsky silver staining. *Frontal cortex (ALZ_441_005 and ALZ_596_006) and striatum (EXT_1998_001 and EXT_2000_001). Immunohistochemistry:*

AT100 and p62. Silver staining: Gallyas and Bielschowsky. Scale bar 50µm except for Bielschowsy staining of case ALZ_596_006 which is 25µm.

b. W**hite matter pathology is scant to moderate in ALZ_441_005**. Tau aggregates are seen in some of the oligodendrocytes. There is a predominance of small round aggregates (black arrow) but some coiled bodies are also seen (white arrow head). Tau aggregates express the 3R-tau and 4R-tau isoforms, are AT100 and p62 positive and are argyrophilic with both Gallyas and Bielschowsky silver stains. *Immunohistochemistry: AT8, 3R-tau, 4R-tau, AT100 and p62. Silver staining: Gallyas and Bielschowsky. Scale bar for top left image 100µm, for top right image 50µm and for inferior row 25µm.*

c. Only rare neurons in the dentate gyrus and CA2 region of the hippocampus are positive for Bielschowsky in ALZ_441_005. *Bielschowsky silver stain. Scale bar: 25µm.*

d. Round neuronal intracytoplasmic tau aggregates were observed in the dentate gyrus in ALZ_596_006. The aggregates were immunoreactive for both 3R-tau and 4R-tau antibodies and were positive for Gallyas and Bielschowsky silver staining. *Immunohistochemistry: 3R-tau and 4R-tau. Silver stains: Gallyas and Bielschowsky. Scale bar: 25µm.*

e. Perivascular thorny astrocytes reminiscent of aging-related tau astrogliopathy (ARTAG) were seen in the periamygdaloid region. Additionally, there were occasional astrocytes containing tau aggregates in the subcortical white matter (parietal lobe). *Immunohistochemistry: AT8. Scale bar: 50µm.*

f. In case ALZ_596_006, 3R tau and 4R tau neurofibrillary tangles and neuropil threads in the hippocampus-CA1. *Immunohistochemistry: 3R-tau, 4R-tau. Silver staining: Gallyas and Bielschowsky. Scale bar: 50µm.*

Supplementary Fig. 4. Western blot using WO2 antibody from the frontal cortex of three MAPT duplication carriers, a subject with Alzheimer disease due to an APP duplication a one normal control.

Supplementary Fig. 5. Analysis of the 4R/3R ratio of MAPT mRNA. Extracted from frontal cortex or striatum. The control group (ctrls) consists in the mean of 5 normal individuals. Error bars represent SEM.

ALZ 441: ALZ_441_005; ALZ 596: ALZ_596_006, EXT 1998: EXT_1998_001

Supplementary Fig. 6. Quantitative analysis of tau seeding activity in a cell biosensor. Proportion of HEK cells containing endogenous tau intracellular aggregates for each tested sample, obtained after 2 x 4 repeated observations in three independent experiments. Patients were compared into a linear mixed effect model for repeated data using Tukey correction for multiple comparisons (Mean of observed proportions ± SEM: CTRL: 0.22% ± 0.11%, PiD cortex: 2.90% ± 0.51%; ALZ_441_005 striatum: 12.22 ± 0.81; ALZ_596_006: 4.48 ± 0.61% ; EXT_1998_001 striatum: 5.95% ± 0.83%; PSP : 14.93% ± 0.90%). Proportion of cells with aggregates is significantly higher in all patients compared to controls (PiD vs CTRL: $p=0.01$; other comparison to controls: all $p<0.001$). Tau seeding activities detected in patients were all pairwise significantly different (ALZ_441_005 vs PSP: $p=0.03$, EXT_1998_001 vs PiD: $p = 0.01$, all other $p<0.001$) except for PiD vs ALZ_596_006 ($p=0.6$) and EXT_1998_001 vs ALZ_596_006 ($p=0.6$).

Supplementary Fig 7. Absence of associated α-synucleinopathy or TDP-43-pathy. Immunohistochemistry for α-synuclein and TDP-43 was negative in all histological studied cases. Aβ was negative in three cases (ALZ_441_005, EXT_1998_001, EXT_2000_001). A few diffuse plaques were seen in the neocortex of ALZ_596_006. First column: frontal cortex. Second column: substantia nigra (SN). Third column: dentate gyrus. Scale bar: 50 µm

Supplementary Table 1. Staining properties of tau immunoreactive aggregates in different cell types in MAPT duplication carriers.

3R-tau, 4R-tau, AT100 and p62 antibodies. Gallyas and Bielschowsky silver staining. R: tau aggregates are sparsely positive. ND: not determined.

	ALZ_596_001	ALZ_596_006	EXT_1593_001	EXT_1114_001	ROU_1373_001	ROU_747_001	EXT_1687_001	ALZ _ 441_005	EXT_1998_001	EXT_2000_001
Published in / novel case	[15]	[15]	novel	[15]	[15]	[21]	novel	[15]	[4]	[4]
Age at onset (years)	50	55	56	57	54	49	53	45	57	37
Sex	M	M	M	F	M	F	F	M	F	M
MAPT haplotype	H1/2H2	H1/2H2	H1/2H2	H1/2H2	2H1/H2	3H1	H1/2H2	2H1/H2	H1/2H2	NA
First reported symptoms	memory	memory	memory, behavior	memory, behavior	memory	behavior	memory	behavior, memory	akinetic-rigid synd.	akinetic-rigid synd.
Family history of dementia	+ (same family as ALZ_596_006)	+ (same family as ALZ_596_001)	+	+	-	+	+	- (de novo mutation)		
Extrapyramidal syndrome at onset	-	-	-	-	-	-	-	-	+	+
behavioral or cognitive dysexec. synd.	-		+	+	-	+	-	+	+	+
visuoperceptive impairment	-		-	-	-		-	-		
Disease duration (years)	11	16 (to death)	6	8	10	20 (to death)	6	13 (to death)	6 (to death)	5 (to death)
MRI (years ADO∔)	5		2	1	4		3	2	4	3
presence of WMH	+		+	+	-		-	-	-	-
anterior temporal atrophy	+		-	-	-		+	+	-	-
internal temporal atrophy	+		+	+	+		+	+	-	-

Scheltens score (R/L)	4-4		3-3	3-3	3-3		4-3	3-3		0-0
frontal atrophy	-		-	-	-		-	+	-	-
parietal/occipital atrophy	-		+	+	+		-	-	-	-
HMPAO-SPECT/FDG-PET (years ADO✢)	3		3	3	5			3		
temporal cortex	**P**		**P**	**P**	**P**			**P**		
parietal cortex	N		**P**	N	**P**			**P**		
frontal cortex	N		N	N	N			**P**		
DaTScan (years ADO✢)			**P (3)**		**P (10)**		**P (6)**			**P (2)**
Amyloid-PET (years ADO✢)	N (6)			N (3)	N (4)					
Tau-PET (years ADO✢)	**P (8)**				**P (9)**					
CSF Biomarkers (years ADO✢)	3		3	3	2		2	1		
Aβ42 (pg.mL^{-1})	**496** / 646*		1372	**372**	864		850	**260**		
Tau (pg.mL^{-1})	**558 / 558***		**789**	**866**	**696**		**1284**	**507**		
P-Tau (pg.mL^{-1})	**66** / 56*		**82**	**94**	**98**		12	**67**		
Aβ42/Aβ40	n.a / 0.07*		0,16				0,06			

Table 1. Demographic, genetic, clinical, imaging and biological data of all *MAPT* duplication carriers

Cells in grey mean data are not available; +: present; -: absent; dysexec. synd.: dysexecutive syndrome; WMH: white matter hyperintensities; HMPAO-SPECT: hexamethylpropanolamine oxime single photon emission computed tomography; FDG-PET: fluorodeoxyglucose positron emission tomography; ADO✢: after disease onset; R: right; L: left; DaTScan: dopamine transporter scan; amyloïd-PET: amyloid-tracer positron emission tomography; tau-PET: Flortaucipir tau-tracer positron emission tomography; CSF: cerebrospinal fluid; Aβ: β-amyloid 1-42 peptide; Aβ40: β-amyloid 1-40 peptide; P-Tau: phosphoryled-tau protein; *: novel analysis after second lumbar puncture. Abnormal CSF biomarker values appear in bold. Normative values for CSF biomarkers were: Aβ42 >550 (pg.mL^{-1}), Tau <350 (pg.mL^{-1}), P-Tau<60 (pg.mL^{-1}), Aβ42/Aβ40>0.055. P:pathological; N:Normal.

	ALZ_441_005	ALZ_596_006	EXT_1998_001	Case III-2 of [1]	EXT_2000_001
frontal cortex	3R>4R	4R>3R#	4R	4R	negative*
hippocampus	3R>4R	3R>4R	3R/4R	3R/4R	4R*
enthorhinal cortex	3R>4R	3R>4R	3R>4R	3R/4R	3R/4R
amygdala	3R>4R	3R>4R	3R/4R	4R > 3R	4R
striatum	3R	3R	4R	4R >> 3R	4R
Pallidum	3R>>4R	3R/4R	4R	4R >> 3R	4R
SN	3R>4R	4R>3R	ND	4R >> 3R	4R
NBM	3R>>4R	3R>4R	3R/4R	3R/4R	4R
Cerebellum	NA	3R/4R	4R	4R	4R

Table 2. Distribution of 4R and 3R aggregates among brain regions in *MAPT* duplication carriers (patients ordered from right to left based on the main aggregated tau isoforms)

3R: positive immunostaining of 3R tau protein isoforms; 4R: positive immunostaining of 4R tau proteins isoforms. Case III-2 of [1]: immunohistochemistry data from the published case by Alexander et al., 2016 [1]; negative*: rare AT8-positive deposits which were negative for 3R and 4R staining; 4R*: rare AT8-positive deposits which were negative for 3R staining; 4R>3R#: In the frontal cortex 3R and 4R were similar in neurons but astrocytes were only positive for 4R. NA: not available.

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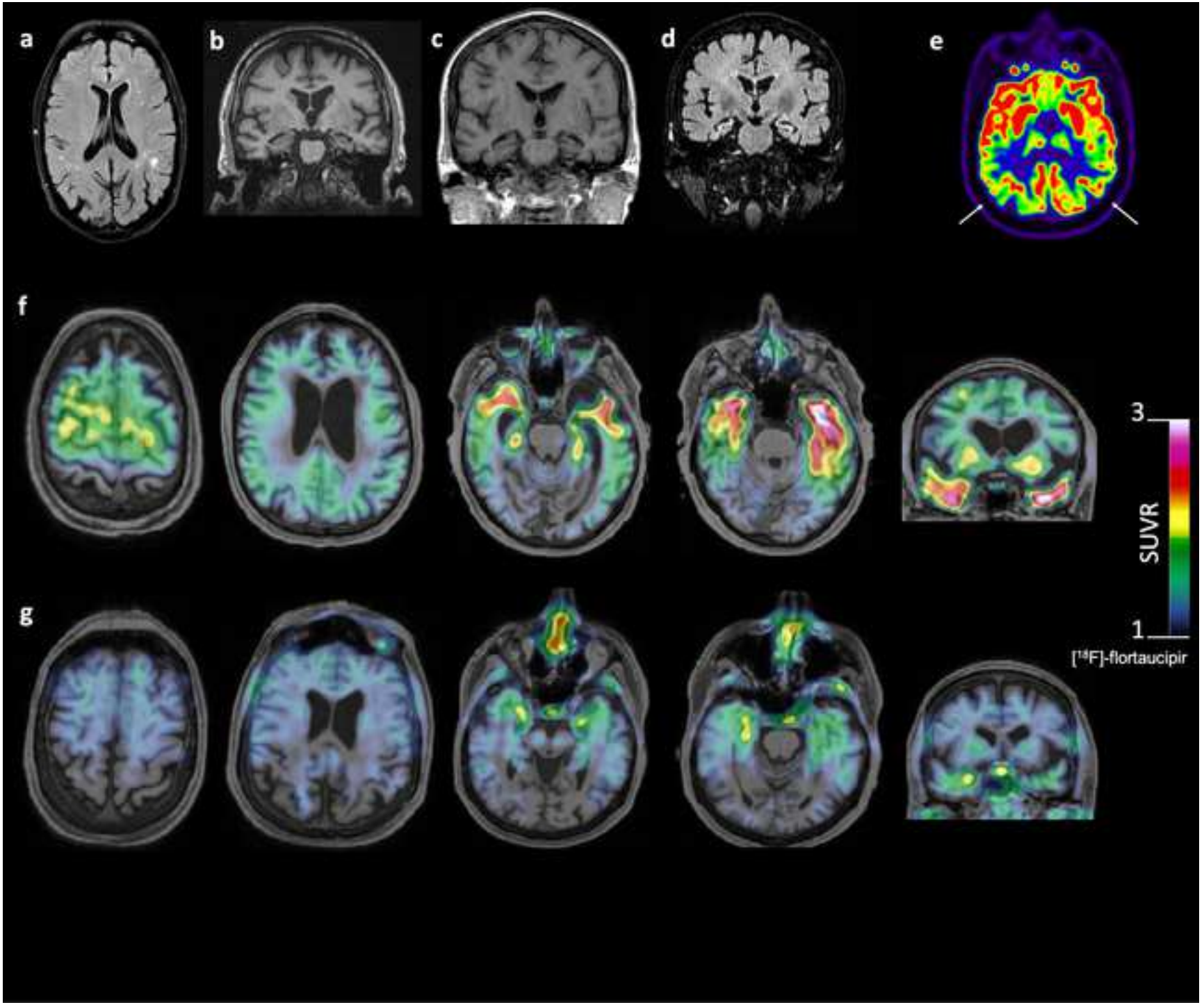

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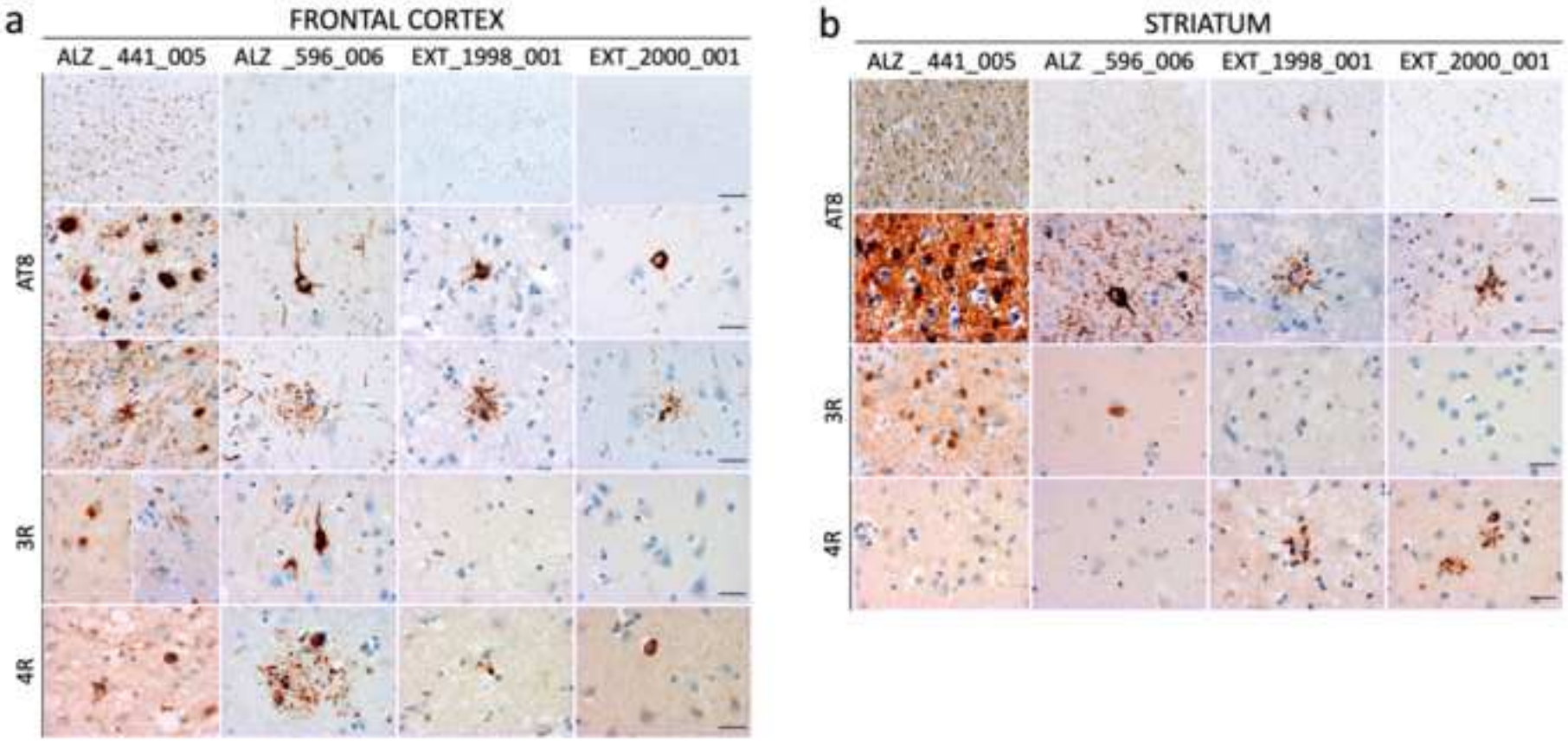

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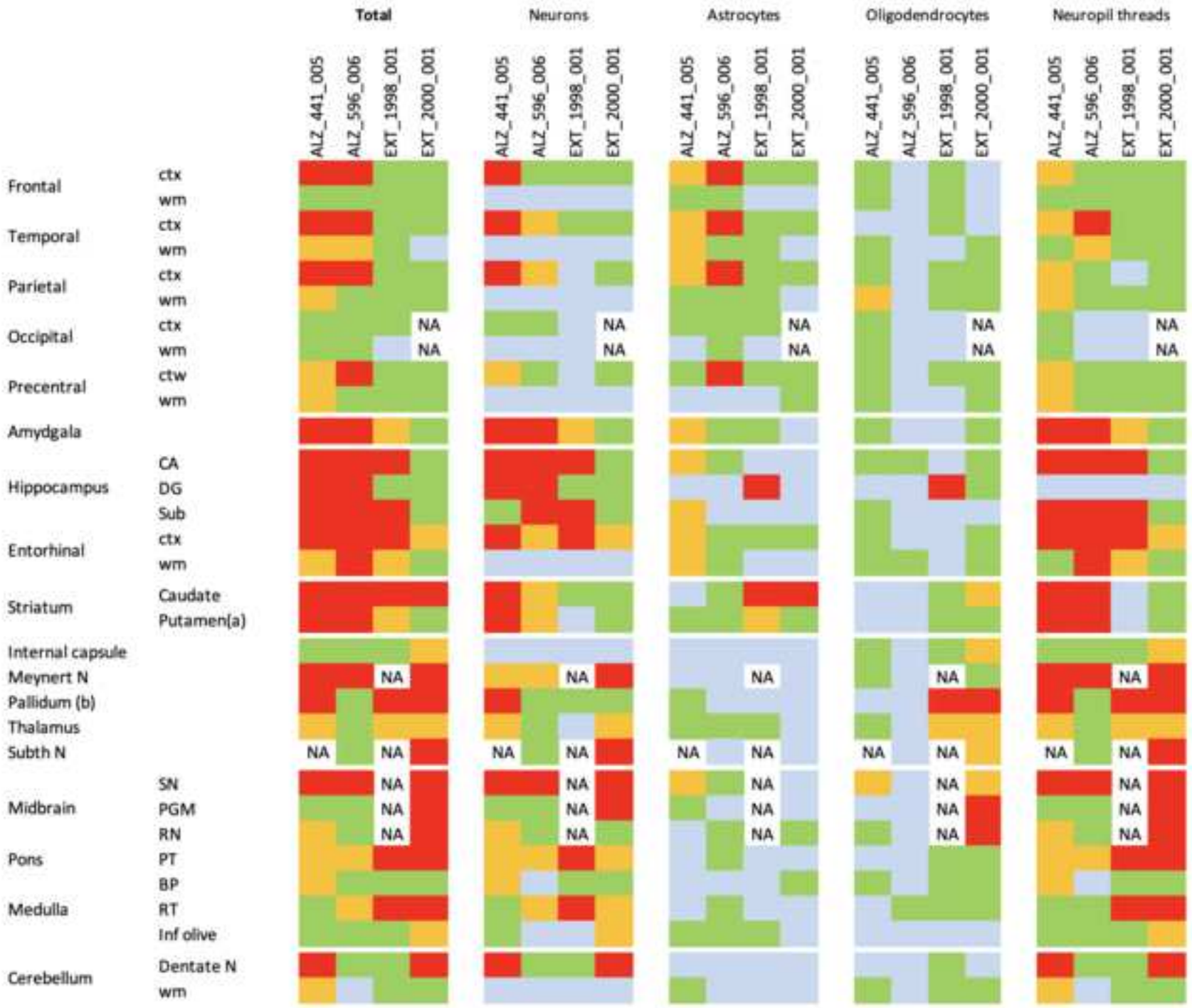

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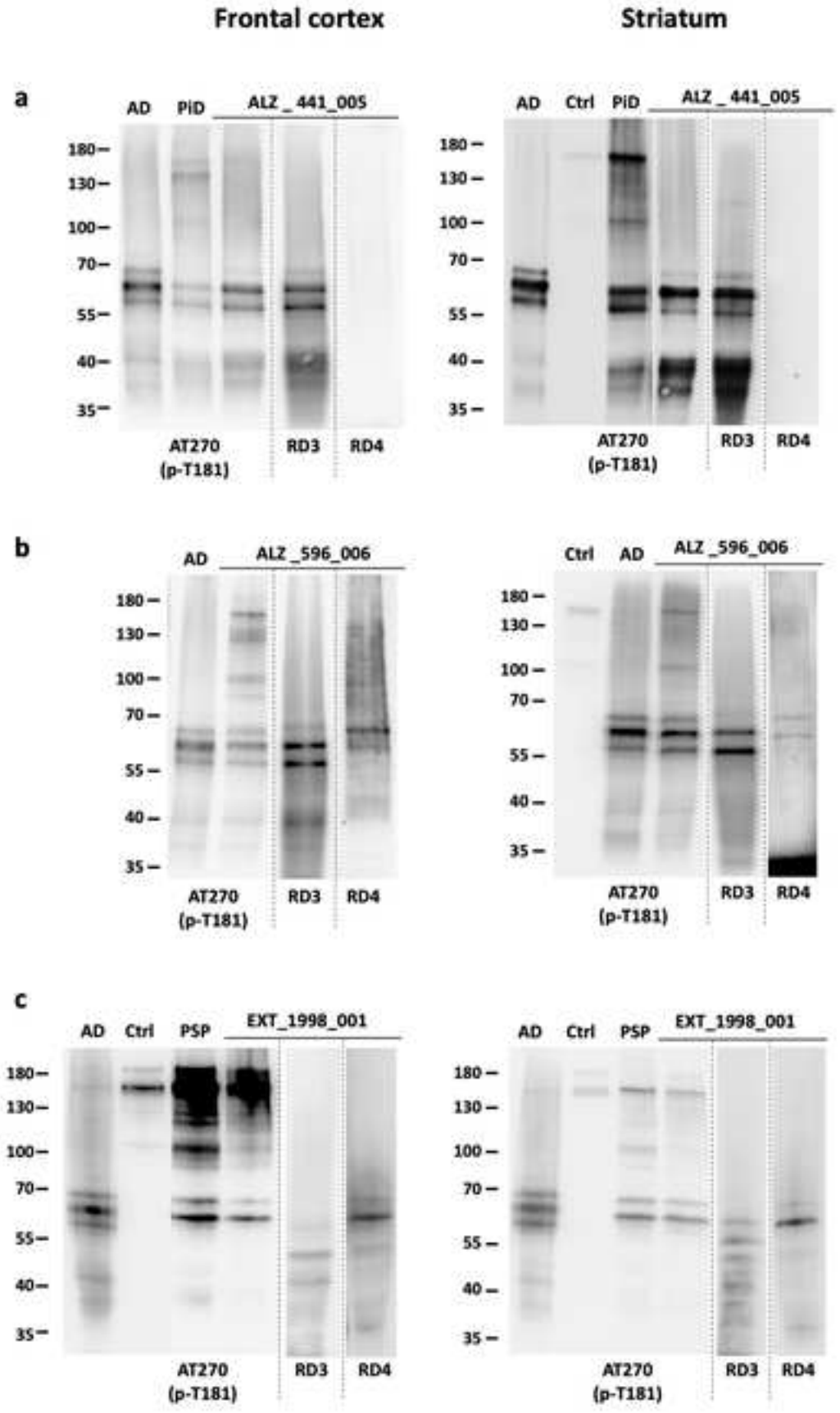

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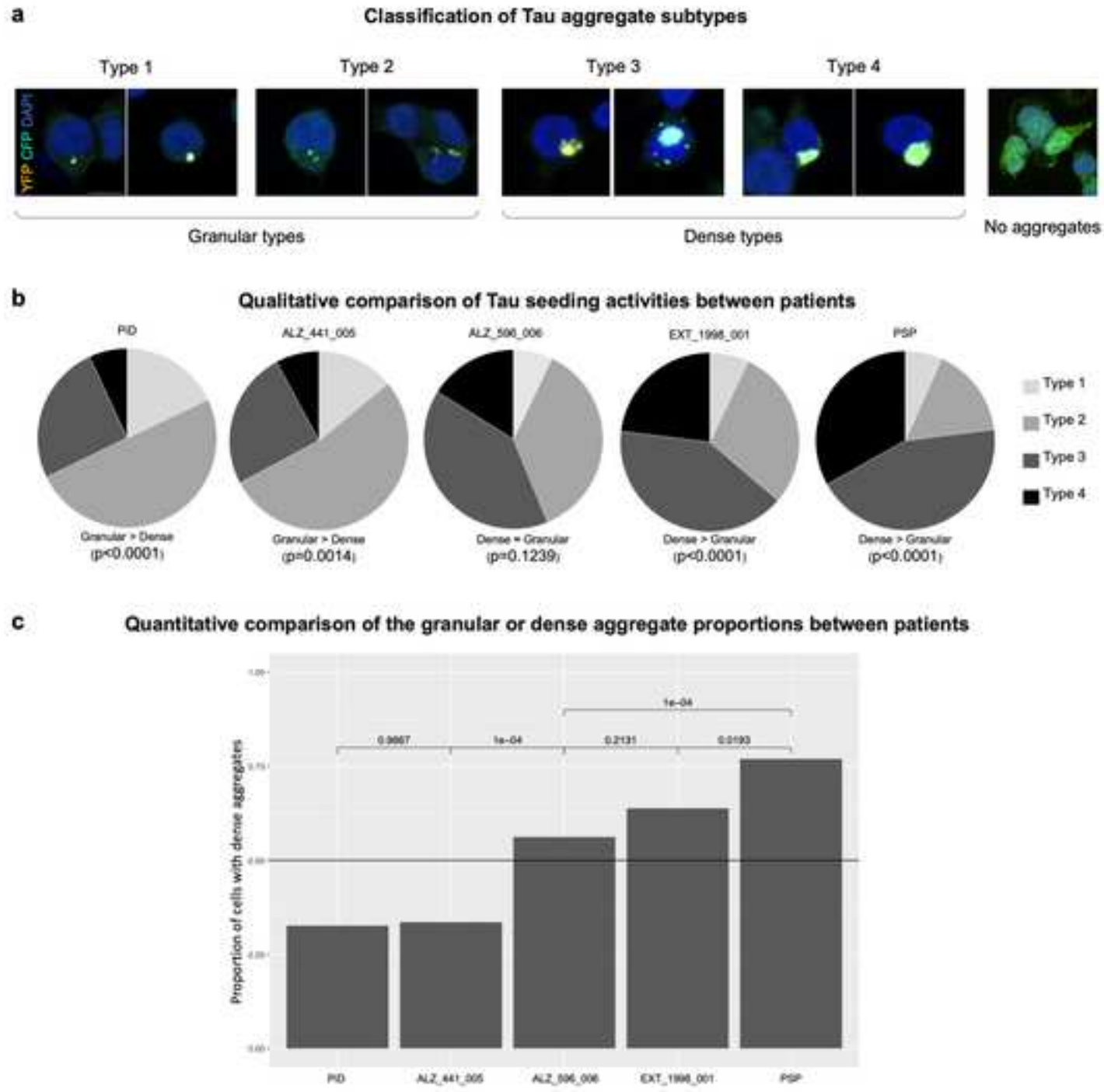

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