

D-serine in physiological and pathological brain aging

E. Ploux, T Freret, Jean-Marie Billard

▶ To cite this version:

E. Ploux, T Freret, Jean-Marie Billard. D-serine in physiological and pathological brain aging. Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics, 2020, pp.140542. 10.1016/j.bbapap.2020.140542. hal-02947335

HAL Id: hal-02947335 https://normandie-univ.hal.science/hal-02947335

Submitted on 23 Sep 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

d-serine in physiological and pathological brain aging

E. Ploux, T. Freret, J.-M. Billard



PII:S1570-9639(20)30189-8DOI:https://doi.org/10.1016/j.bbapap.2020.140542Reference:BBAPAP 140542To appear in:BBA - Proteins and ProteomicsReceived date:8 July 2020Revised date:10 September 2020Accepted date:13 September 2020

Please cite this article as: E. Ploux, T. Freret and J.-M. Billard, d-serine in physiological and pathological brain aging, *BBA - Proteins and Proteomics* (2020), https://doi.org/10.1016/j.bbapap.2020.140542

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2020 Published by Elsevier.

D-serine in physiological and pathological brain aging

E. Ploux^{a,*} eva.ploux@unicaen.fr, T. Freret^a and J-M Billard^{a,*} jean-marie.billard@inserm.fr

^aNormandie Univ, UNICAEN, INSERM, CYCERON, COMETE, 14000 Caen, France. *Corresponding authors at: UNICAEN COMETE, UMR S-1075, Bat GMPc, Campus Horowitz, Bd Henri Becquerel, CS14032 Caen Cedex 5, France.

Abstract

Among aging-induced impairments, thos a rifecting cognitive functions certainly represent one the most major challenge to face to hip, ove elderly quality of life. In last decades, our knowledge on changes in the morphology and function of neuronal networks associated with normal and pathological brain agins, bas rapidly progressed, initiating the development of different pharmacological and b havoural strategies to alleviate cognitive aging. In particular, experimental evidences have cocumulated indicating that the communication between neurons and its plasticity gradually veakens with aging. Because of its pivotal role for brain functional plasticity, the *N*-Methyl *D*-Aspartate receptor subtype of glutamate receptors (NMDAr) has gathered much of the experimental interest. NMDAr activation is regulated by many mechanisms. Among is the mandatory binding of a co-agonist, such as the amino acid D-serine, in order to activate NMDAr. This mini-review presents the most recent information indicating how D-serine could contribute to mechanisms of physiological cognitive aging and also considers the divergent views relative of the role of the NMDAr co-agonist in Alzheimer's disease. **Keywords:** synaptic plasticity, NMDA receptors, serine racemase, Alzheimer's disease, glutamate, L-serine

1. Introduction

Beyond increasing life expectancy, improving health status of elderly is now one of the most current important challenge to face in the field of research on "aging processes". Aging is a natural biological process that affects the entire body. Notably, while affecting the brain, cognitive decline and especially memory processes alterations, would play a pivotal role in the poor quality of life of elders.

Although initially associated with neuronal loss, underpining mechanisms of brain aging still remain debated despite an extensive literature [1-5]. This may be related in part to limits of the initial stereological methods used to estimate the neuronal density [6-8]. To date, the theory that prevails does not consider the decrease in cells number as a hallmark of physiological aging, but rather to its pathological aspect such as Alzheimer disease (AD) affording neuronal and synaptic losses [3-5, 9-11].

Essential relay in the formation of many types of memory traces, the hippocampal formation has been particularly studied, both in 1 u nans and animals [12-15]. Hence, in rodents [16, 17], a decreased ability to consolidate long-term memory has been linked to a weaker capacity to express long-term potentiation (LTP) of synaptic transmission (reviewed in [18]). Besides, a rapid decay of hippocar.p.¹ LTP was correlated to poor spatial memory performances in aging rats, which probably contributes to the increased propensity to forget spatial information [19]. Overal, age-induced decrease in synaptic plasticity efficiency would impaired capacity to encode and/or consolidate new memories and would be in part responsible for loss of pre-existing ones [20-22]. These synaptic functional alterations and the subsequent cognitive deficits have been linked in part to a progressive deregulation of the activation of NMDA receptors (NMDAr). NMDAr is a subtype of glutamate receptors generally views as critical to drive synaptic plasticity in many brain areas [23-26]. In addition to glutamate, NMDAr activation requires the binding of a co-agonist at its specific site. Initially attributed to glycine [27-29], subsequent studies showed that D-serine could also play this role. Indeed, both the specific degradation of the amino acid D-serine by the enzyme Damino acid oxidase (DAAO) [30] and the genetic deletion of its synthesizing enzyme serine

racemase (SR) [31, 32], significantly altered NMDAr activation. These results therefore led to consider D-serine as the main endogenous co-agonist of synaptic NMDAr and further experiments confirmed the pivotal role of this amino acid in the regulation of functional plasticity at many synapses throughout the brain (reviewed in [33]).

2. D-serine in physiological aging

Alterations of functional plasticity associated with cognitive impairments observed across aging naturally raised the question for a role of D-serine. Thereafter, it was observed that a supplementation with the amino acid was able to prevent age-induced deregulation of NMDAr activation and to restore hippocampal synaptic plasticity [34-37]. Beyond the decrease in NMDAr density (or its subunits) initially evoked [38-41], these promising results rather argue for a pivotal role of D-serine in functional mechanisms underlying cognitive aging [42, 43].

The beneficial effect of a D-serine supplementation could reflect a significant decrease in Dserine levels with age, as supported in rodent hippocampal assays [34-36]. Interestingly, this decrease is also found in human plasma levels [44]. And net explanation could have been a decrease in the degree of NMDAr saturation, but autoral'iographic studies performed in aged rodents did not reveal changes in the specific affinit, of the co-agonist binding site [34, 35, 45]. Besides, other results from preclinical investigations performed in rodents indicate that the decrease in D-serine levels could rely of a *weaker* SR expression both at protein and messenger RNA levels [34-36], though whether the activity of the D-serine synthesizing enzyme is altered at the same time remains to be determined. On the other hand, the expression of the catabolizing enzyr. Dr.AO is not affected in aged animals, indicating that an acceleration of the degradation of the amino acid does not contribute to the age-related decrease in D-serine levels [34-36]. The synaptic availability of D-serine is also dependent on the activity of the Asc-1 sub, me of neutral amino acid transporters, which supports the release of D-serine from neu ons [46, 47]. A recent investigation indicates that Asc-1 activity remained unchanged in the aging hippocampus [48]. Considering that mechanisms underlying the degradation and release of D-serine are not altered in physiological aging, alterations in synthesis therefore appears as the critical mechanism underlying the age-related decrease of the NMDAr co-agonist availability [42, 43].

The decrease in SR expression and D-serine production possibly results from the development of the oxidative stress (OS) that normally occurs with aging [49-52]. Indeed, both deficits do not occur in the LOU/C rat [36], a model of successful aging characterized by a high degree of resistance to OS [53-56]. They are also prevented in aged animals receiving a long-term treatment with the reducing compound N-acetyl-cysteine in which the extent of OS is minimized [57]. An increased oxidation of sulfydryl groups of SR [58] and/or changes in its dimer active conformation [59] may be viewed as possible mechanisms underlying the age-

related SR deregulation by OS. Interestingly, aged LOU/C rats show not only intact synaptic plasticity and NMDAr activation but also preserved memory abilities [55, 56, 60]. Compelling all these available functional and behavioural data therefore indicates that a preservation of SR activity and a robust synaptic availability of D-serine in cerebral tissues is essential to maintain potent cognitive capacities in physiological aging (Figure 1). It is interesting to note that the impaired learning of olfactory conditioning in aged drosophila, which also display decreased D-serine levels, was reversed by feeding flies with the amino acid [61]. Nevertheless, a supplementation with D-serine to prevent memory deficits in aged mammals has not been considered yet although treatment with the related agonist D-cycloserine is clearly beneficial in aged rodents [62, 63]. This is presumably because long-term treatment with D-serine induced serious side effects, net hro- and hepato-toxicity notably [64, 65].

3. D-serine in Alzheimer's disease

Although our knowledge on pathorbysiological mechanisms underlying the onset and development of Alzheimer's disease (AL) has increased exponentially these last decades, its aetiology is far from being fully clarified. Indeed, AD is complex and multifactorial, with a notable overlap between familial an (s_{β}) radic forms. Its symptomatology has been associated with a convoluted and multiple n echonisms, including an increase in the production of the β -amyloid (A β) peptide that accumulates intracellularly while also aggregates in extracellular deposits, and enhanced internet ironal neurofibrils with hyperphosphorylated TAU protein. These typical features are accompanied with gliosis and synaptotoxicity that finally results in neuronal loss and brain at ophy (see [66] for a review).

The NMDAr represents one of the most predominant extracellular targets for AD-related pathology. Indeed, even though still matter of debates, impairments of NMDAr are viewed as core pathophysiological mechanisms of the disease [67-69]. Whereas synaptic glutamate loss of availability is known to contribute to pathology [70, 71], changes in D-serine levels and putative underlying processes remain so far subject of controversial views.

The first link between D-serine and AD came from biochemical investigations. Indeed, either cerebrospinal fluid (CSF) or plasma changes in amino acid levels were observed in AD patients [72-80]. Unfortunately, divergent results appear from one study to another, regarding the magnitude but also the direction of these changes (Table 1). One possibility to account for

the discrepancies could be found on the Braak stage of the patients sampled since D-serine levels are thought to progressively increase in the CSF across the progression of the pathology [78]. Further, increased expression of the D-serine synthetizing enzyme was found in the hippocampus of Braak stage III-IV AD patients [81]. However, a significant rise of the co-agonist in the pathophysiology of the disease has been questioned by a subsequent analysis but the Braak stage in this study was not clearly defined [72]. Finally, it's worth mentioning that an increase level of extracellular transcript of the phosphoglycerate dehydrogenase (PHGDH) was recently reported in subjects at presymptomatic stage, i.e. before being diagnosed AD [82]. PHGDH catalyzes serine biosynthesis in the brain from glucose[83]. Therefore, the elevation of its extracellular RNA may argue for on increase of D-serine brain level during presymptomatic stage of the disease.

Similarly, this issue does not make consensus in the field of preclinical investigations ([78, 84-86], see Table 1). In fact, an increase of D-set, re levels in hippocampal tissues has been observed in several animal models of AD, inc'aoing those with intracerebroventricular injections of soluble oligomers of the A β pept $\frac{1}{2}$ (A β o) and genetic strains bearing several mutations associated with familiar forms (1 AL [78]. Thus, preliminary results in the 5xFAD mouse model of AD displaying high and *c* ronic elevation of A_b charge show a limited but significant increase in D-serine level. (Ploux et al, abstract IDAR 2019). It has been previously demonstrated *in-vitro* that Af o could promote transcriptional expression of SR and stimulate D-serine release from both activated microglial cells [87, 88] and neurons [70]. More recently, induction of SR e. pression has also been found in vitro in reactive neurotoxic astrocytes [89] and in-vivo in the TgF344 rat model of AD [81]. Quite interestingly, SR knockout mice display similicant reduced brain lesion after acute injections of Aβo [90]. Finally, our preliminary results indicate that the impairment of hippocampal synaptic plasticity and memory deficits associated with the increased D-serine levels in 5xFAD mice are rescued by a concomitant deletion of the SR gene. Altogether, this set of results supports the view that D-serine is required for the induction and development of the Aβo-dependent pathophysiology.

However, this view is not so straightforward (Figure 2). Indeed, observations from two transgenic mice models have extended the debate into the preclinical field on how D-serine contributes to AD. In fact, a decreased synthesis of L-serine from glucose though PHGDH astrocytic enzyme has recently been reported in 3xTg-AD mice [84]. Astrocytic L-serine was proposed to regulate NMDAR activity to sustain D-serine production by neurons [91] that

would thus explain the altered synaptic plasticity observed in AD. The deficits of functional plasticity in the hippocampus and memory impairments in these 3xTg-AD mice [84] are rescued by long-term treatments with either D- or L-serine, confirming the view of severe metabolic dysfunction as a leading hallmark and cause of AD as previously proposed (see [92]).

Besides, a weaker amount of the NMDAr co-agonist was observed in Amyloid Precursor Protein (APP) knock out mice [93]. This suggests that in addition to changes induced by increased synthesis of soluble A β o, the native protein is capable by itself to regulate D-serine homeostasis.

The question of the potential modification of D-serine levels in ... D and its opposite directions found in the literature, points out several important aspects, notably due to predominant role of D-serine as a NMDAr co-agonist, that remain to be solv d. Several explanations could account for the controversial results resumed above. First as recently emphasized [94], the quantification of D-serine levels must comply with strict procedures allowing a precise and specific detection of the amino acid. However, an studies aimed at quantifying D-serine did not fulfil all the validation criteria. Second, a discussed by Le Douce and colleagues [84]), results may differ from one animal mcdel to another since each of them reproduces only partially and differently the mechanicms and severity of AD-associated pathophysiology. As for examples, the intraneuronal A β as r may be very different among the animal models of amyloidogenesis. Also, levels of soluble A β o are not defined in the study of Le Douce [84] which may be too low to impact SR activity. On the other hand, the glycemia and carbohydrate status of the 5xF.,D mice used in the study of Ploux et al has not yet been considered, thus questioning putative changes in L-serine availability. Advantages and limits of modelling the disease in animals has been largely considered in the literature but our review, even by focussing on the specific role of D-serine in AD, indicates that this question still remains an important and open issue [95].

3. Conclusion

Together, the results of clinical and preclinical studies show an undeniable variation in Dserine levels in both physiological and pathological aging. Regarding the first aspect, studies agree on a decrease in the availability of the amino acid that would directly impact functional synaptic plasticity and memory capacities. These data are particularly stimulating to initiate the search of future pharmacological treatments to compensate for the loss of D-serine. In

contrast, a therapeutic perspective regarding the pathological aspect of aging still remains utopic since the view of how the amino acid behaves in both humans and animal models of AD does not make consensus. Nevertheless, one may speculate that the opposite decrease or increase in D-serine levels reported along AD-related studies could be speculated as reflecting the slow development of the disease and particularly of its amyloid progression. In conditions of low A β o production, the alteration of glucose intake and consequently the reduced production of the precursor L-serine could be considered as a critical mechanism that decreases D-serine levels in the early stages of the disease. Then, as the A β o production elevates, this mechanism could be overpassed by the direct stimulation of SR activity, thus enhancing D-serine levels. This interesting schema has now to be precisely evaluated and represents an essential milestone since it means that the best effective therapeutic strategy of delivery of the amino acid should closely depend on the st. ge (f the disease.

A further point remains also to be considered for the evaluation of D-serine in AD. As recently demonstrated, epigenetic mechanisms play r key role in the regulation of cerebral D-serine levels [96]. Therefore, the evaluation of h_r i npact of epigenetics in the expression of D-serine degradation (DAAO) and synthesis (CR) enzyme genes should represent a major issue in AD.

On the other hand, this review indicates that there is still clearly a crucial need of animal models really mimicking the slow progression and the changes in A β and Tau protein levels of human AD. Hopefully, a new generation of models such as those currently developed with viral-mediated gene transfer [27, 98] will certainly allow for more consistent results and clearer understanding of the involvement of D-serine in the pathophysiology of AD.

Figure 1: Schematic representation of changes in D-serine availability at CA3/CA1 hippocampal synapses during physiological aging.

Once formed from glucose in astrocytes, the precursor L-serine shuttles to the neuronal compartment to be converted into D-serine by serine racemase (SR). After being released in the synaptic cleft, D-serine binds to NMDAr, together with glutamate, allowing activation of the receptor. During the aging processes, the progressive installation of the oxidative stress decreases the expression or the dimerization of SR, thus reducing D-serine synthesis. Consequently, NMDAr activation is altered that impairs the expression of functional plasticity such as long-term potentiation in neuronal networks and, ultimately of the memory encoding.

Figure 2: Schematic representation of changes in D-serine homeostasis at CA3/CA1 hippocampal synapses during the progressive installation of amyloid-related pathophysiology of AD.

At early stages of the disease with low levels of soluble $A\beta$ oligomers (left), glucose catabolism is impacted that decreases the synthesis of the precursor L-serine, thus reducing D-serine levels. This leads to weaker NMDAr activation, impaired functional plasticity in neuronal networks and ultimately deficits of memory processes.

As levels of soluble A\u00f3o increase (right), astrocytes become reactive and start to express SR and release high amount of D-serine. Together with the inc. Pass in synaptic availability of glutamate, the elevation of D-serine drives NMDAr over-active tion resulting in excitotoxicity, neurodegeneration and profound memory deficits.

Credit author statement

EP, TF and JMB have contributed to the elaboration and writing of the review. EP has built the Table and figures.

Declaration of interests

X The authors declare that they have no known competing financial interests or personal relationships that could have one wed to influence the work reported in this paper.

References:

[1] B.H. Anderton, Ageing of the brain, Mech Ageing Dev 123(7) (2002) 811-7.

[2] L.E.B. Bettio, L. Rajendran, J. Gil-Mohapel, The effects of aging in the hippocampus and cognitive decline, Neurosci Biobehav Rev 79 (2017) 66-86.

[3] S.N. Burke, C.A. Barnes, Senescent synapses and hippocampal circuit dynamics, Trends Neurosci 33(3) (2010) 153-61.

[4] I. Driscoll, D.A. Hamilton, H. Petropoulos, R.A. Yeo, W.M. Brooks, R.N. Baumgartner, R.J. Sutherland, The aging hippocampus: cognitive, biochemical and structural findings, Cereb Cortex 13(12) (2003) 1344-51.

[5] M.M. Esiri, Ageing and the brain, The Journal of pathology 211(2) (2007) 181-7.

[6] D.L. Brown, Bias in image analysis and its solution: unbiased stereology, Journal of toxicologic pathology 30(3) (2017) 183-191.

[7] D.F. Marrone, J.C. LeBoutillier, T.L. Petit, Complementary techniques for unbiased stereology of brain ultrastructure, Journal of electron microscopy 52(4) (2003) 425-8.

[8] R.M.A. Napper, Total Number Is Important: Using the Disector Method in Design-Based Stereology to Understand the Structure of the Rodent Brain, Frontiers in neuroanatomy 12 (2018) 16.
[9] K.A. Baskerville, C. Kent, M.M. Nicolle, M. Gallagher, M. McKinney, Aging causes partial loss of basal forebrain but no loss of pontine reticular cholinergic neurons, Neuroreport 17(17) (2006) 1819-23.

[10] S.A. Chance, Subtle changes in the ageing human brain, Nutrition and health 18(3) (2006) 217-24.

[11] T. Wyss-Coray, Ageing, neurodegeneration and brain rejuvenation, Nature 539(7628) (2016) 180-186.

[12] N. Burgess, E.A. Maguire, J. O'Keefe, The human hippocampus and spatial and episodic memory, Neuron 35(4) (2002) 625-41.

[13] H. Eichenbaum, Memory: Organization and Control, Annu Rev Psychol 68 (2017) 19-45.

[14] B. Opitz, Memory function and the hippocampus, Frontiers of neurology and neuroscience 34 (2014) 51-9.

[15] J.T. Wixted, L.R. Squire, The role of the human hippocanous in familiarity-based and recollection-based recognition memory, Behav Brain Res 215(2) (2) 10, 197-208.

[16] C.A. Barnes, B.L. McNaughton, An age comparison of the rates of acquisition and forgetting of spatial information in relation to long-term enhancement of hopo ampal synapses, Behav Neurosci 99(6) (1985) 1040-8.

[17] J.P. Lister, C.A. Barnes, Neurobiological changes in the hippocampus during normative aging, Arch Neurol 66(7) (2009) 829-33.

[18] J.M. Billard, Ageing, hippocampal synaptic activity and magnesium, Magnes Res 19(3) (2006) 199-215.

[19] C.A. Barnes, B.L. McNaughton, Physiologica¹ compensation for loss of afferent synapses in rat hippocampal granule cells during senescence, J + hysiol 309 (1980) 473-85.

[20] T.C. Foster, Biological markers of a related memory deficits: treatment of senescent physiology, CNS Drugs 20(2) (2006) 153-66.

[21] P.W. Landfield, J.L. McGaugh, G. Lynch, Impaired synaptic potentiation processes in the hippocampus of aged, memory-deficient race, Brain Res 150(1) (1978) 85-101.

[22] E.S. Rosenzweig, C.A. Barnes, trocce of aging on hippocampal function: plasticity, network dynamics, and cognition, Prog Neurobiol 69(3) (2003) 143-79.

[23] T.V. Bliss, G.L. Collingridge, A synaptic model of memory: long-term potentiation in the hippocampus, Nature 361(6407) (1993) 31-9.

[24] G.L. Collingridge, T.V. Bliss, Memories of NMDA receptors and LTP, Trends Neurosci 18(2) (1995) 54-6.

[25] R.C. Malenka, R.A. dicou, NMDA-receptor-dependent synaptic plasticity: multiple forms and mechanisms, Trends Neuror ci 16(12) (1993) 521-7.

[26] R.G. Morris, NMDA receptors and memory encoding, Neuropharmacology 74 (2013) 32-40.

[27] J.W. Johnson, P. Ascher, Glycine potentiates the NMDA response in cultured mouse brain neurons, Nature 325(6104) (1987) 529-31.

[28] N.W. Kleckner, R. Dingledine, Requirement for glycine in activation of NMDA-receptors expressed in Xenopus oocytes, Science 241(4867) (1988) 835-7.

[29] N.W. Kleckner, R. Dingledine, Regulation of hippocampal NMDA receptors by magnesium and glycine during development, Brain Res Mol Brain Res 11(2) (1991) 151-9.

[30] J.P. Mothet, A.T. Parent, H. Wolosker, R.O. Brady, Jr., D.J. Linden, C.D. Ferris, M.A. Rogawski, S.H. Snyder, D-serine is an endogenous ligand for the glycine site of the N-methyl-D-aspartate receptor, Proc Natl Acad Sci U S A 97(9) (2000) 4926-31.

[31] D.T. Balu, Y. Li, M.D. Puhl, M.A. Benneyworth, A.C. Basu, S. Takagi, V.Y. Bolshakov, J.T. Coyle, Multiple risk pathways for schizophrenia converge in serine racemase knockout mice, a mouse model of NMDA receptor hypofunction, Proc Natl Acad Sci U S A 110(26) (2013) E2400-9.

[32] A.C. Basu, G.E. Tsai, C.L. Ma, J.T. Ehmsen, A.K. Mustafa, L. Han, Z.I. Jiang, M.A. Benneyworth, M.P. Froimowitz, N. Lange, S.H. Snyder, R. Bergeron, J.T. Coyle, Targeted disruption of serine racemase affects glutamatergic neurotransmission and behavior, Mol Psychiatry 14(7) (2009) 719-27.
[33] J.M. Billard, D-Amino acids in brain neurotransmission and synaptic plasticity, Amino Acids 43(5) (2012) 1851-60.

[34] J.P. Mothet, E. Rouaud, P.M. Sinet, B. Potier, A. Jouvenceau, P. Dutar, C. Videau, J. Epelbaum, J.M. Billard, A critical role for the glial-derived neuromodulator D-serine in the age-related deficits of cellular mechanisms of learning and memory, Aging Cell 5(3) (2006) 267-74.

[35] B. Potier, F.R. Turpin, P.M. Sinet, E. Rouaud, J.P. Mothet, C. Videau, J. Epelbaum, P. Dutar, J.M. Billard, Contribution of the d-Serine-Dependent Pathway to the Cellular Mechanisms Underlying Cognitive Aging, Front Aging Neurosci 2 (2010) 1.

[36] F.R. Turpin, B. Potier, J.R. Dulong, P.M. Sinet, J. Alliot, S.H. Oliet, P. Dutar, J. Epelbaum, J.P. Mothet, J.M. Billard, Reduced serine racemase expression contributes to age-related deficits in hippocampal cognitive function, Neurobiol Aging 32(8) (2011) 1495-504.

[37] S. Yang, H. Qiao, L. Wen, W. Zhou, Y. Zhang, D-serine enhances appaired long-term potentiation in CA1 subfield of hippocampal slices from aged senescence-accelerated mouse prone/8, Neurosci Lett 379(1) (2005) 7-12.

[38] D.A. Clayton, M.D. Browning, Deficits in the expression of the IR2B subunit in the hippocampus of aged Fisher 344 rats, Neurobiol Aging 22(1) (2001) 165-8.

[39] K.R. Magnusson, Aging of glutamate receptors: Concelations between binding and spatial memory performance in mice, Mech Ageing Dev 104(3) (1, 98, 227-48.

[40] D. Wang, S.A. Jacobs, J.Z. Tsien, Targeting the N! IDA receptor subunit NR2B for treating or preventing age-related memory decline, Expert op nion on therapeutic targets 18(10) (2014) 1121-30.

[41] G.L. Wenk, C.A. Barnes, Regional char ge. in the hippocampal density of AMPA and NMDA receptors across the lifespan of the rat, Brain 8 is 885(1) (2000) 1-5.

[42] J.M. Billard, Serine racemase as a prime taget for age-related memory deficits, Eur J Neurosci 37(12) (2013) 1931-8.

[43] J.M. Billard, Changes in Serine Racer in 2-Dependent Modulation of NMDA Receptor: Impact on Physiological and Pathological Brain Aging, crontiers in molecular biosciences 5 (2018) 106.

[44] M.A. Calcia, C. Madeira, F.V. , 'heira, T.C. Silva, F.M. Tannos, C. Vargas-Lopes, N. Goldenstein, M.A. Brasil, S.T. Ferreira, R. Panizzutti, Plasma levels of D-serine in Brazilian individuals with schizophrenia, Schizophr Res 142(1-3) (2012) 83-7.

[45] Y. Nagata, T. Uehara, V N⁺⁻mura, Y. Nomura, K. Horiike, D-serine content and D-[3H]serine binding in the brain regions of the senescence-accelerated mouse, Mech Ageing Dev 104(2) (1998) 115-24.

[46] D. Rosenberg, S. Artov, A.C. Segal, G. Kolodney, I. Radzishevsky, E. Dikopoltsev, V.N. Foltyn, R. Inoue, H. Mori, J.M. Billard, H. Wolosker, Neuronal D-Serine and Glycine Release Via the Asc-1 Transporter Regulates NMDA Receptor-Dependent Synaptic Activity, J Neurosci 33(8) (2013) 3533-44.

[47] H. Sason, J.M. Billard, G.P. Smith, H. Safory, S. Neame, E. Kaplan, D. Rosenberg, S. Zubedat, V.N. Foltyn, C.T. Christoffersen, C. Bundgaard, C. Thomsen, A. Avital, K.V. Christensen, H. Wolosker, Asc-1 Transporter Regulation of Synaptic Activity via the Tonic Release of d-Serine in the Forebrain, Cereb Cortex 27(2) (2017) 1573-1587.

[48] J.M. Billard, T. Freret, Asc-1 transporter activation: an alternative to rescue age-related alterations in functional plasticity at rat hippocampal CA3/CA1 synapses, J Neurochem 147(4) (2018) 514-525.

[49] W. Droge, H.M. Schipper, Oxidative stress and aberrant signaling in aging and cognitive decline, Aging Cell 6(3) (2007) 361-70.

[50] K. Fukui, K. Onodera, T. Shinkai, S. Suzuki, S. Urano, Impairment of learning and memory in rats caused by oxidative stress and aging, and changes in antioxidative defense systems, Ann N Y Acad Sci 928 (2001) 168-75.

[51] T.R. Golden, D.A. Hinerfeld, S. Melov, Oxidative stress and aging: beyond correlation, Aging Cell 1(2) (2002) 117-23.

[52] A. Kumar, B. Yegla, T.C. Foster, Redox Signaling in Neurotransmission and Cognition During Aging, Antioxid Redox Signal (2017).

[53] J. Alliot, S. Boghossian, D. Jourdan, C. Veyrat-Durebex, G. Pickering, D. Meynial-Denis, N. Gaumet, The LOU/c/jall rat as an animal model of healthy aging?, J Gerontol A Biol Sci Med Sci 57(8) (2002) B312-20.

[54] S. Boghossian, G. Nzang Nguema, D. Jourdan, J. Alliot, Old as mature LOU/c/jall rats enhance protein selection in response to a protein deprivation, Exp Gerontol 37(12) (2002) 1431-40.

[55] M. Kollen, A. Stephan, A. Faivre-Bauman, C. Loudes, P.M. Sinet, J. Alliot, J.M. Billard, J. Epelbaum, P. Dutar, A. Jouvenceau, Preserved memory capacities in aged Lou/C/Jall rats, Neurobiol Aging 31(1) (2010) 129-42.

[56] V. Paban, J.M. Billard, V. Bouet, T. Freret, M. Boulouard, C. Chambon, B. Loriod, B. Alescio-Lautier, Genomic transcriptional profiling in LOU/C/Jall rats identifies genes for successful aging, Brain Struct Funct 218(6) (2013) 1501-12.

[57] C. Haxaire, F.R. Turpin, B. Potier, M. Kervern, P.M. Sinet, G. Barbanet, J.P. Mothet, P. Dutar, J.M. Billard, Reversal of age-related oxidative stress prevents hippocartic synaptic plasticity deficits by protecting d-serine-dependent NMDA receptor activation, Agir g Ce 11(2) (2012) 336-44.

[58] A.K. Mustafa, M. Kumar, B. Selvakumar, G.P. Ho, J.T. ^chmsen, R.K. Barrow, L.M. Amzel, S.H. Snyder, Nitric oxide S-nitrosylates serine racemase, neurating feedback inhibition of D-serine formation, Proc Natl Acad Sci U S A 104(8) (2007) 2950-5.

[59] W. Wang, S.W. Barger, Cross-linking of serine race nase dimer by reactive oxygen species and reactive nitrogen species, J Neurosci Res 90(6) (2017) ¹.278-29.

[60] C. Menard, R. Quirion, S. Bouchard, G. Ferland, T. Gaudreau, Glutamatergic signaling and low prodynorphin expression are associated within tack memory and reduced anxiety in rat models of healthy aging, Front Aging Neurosci 6 (2014) 21.

[61] D. Yamazaki, J. Horiuchi, K. Ueno, T. Ueno, S. Saeki, M. Matsuno, S. Naganos, T. Miyashita, Y. Hirano, H. Nishikawa, M. Taoka, Y. Yamauchi, T. Isobe, Y. Honda, T. Kodama, T. Masuda, M. Saitoe, Glial dysfunction causes age-related m(mon); impairment in Drosophila, Neuron 84(4) (2014) 753-63.
[62] J. Aura, M. Riekkinen, P. Riekkinen, *i.r.*, Tetrahydroaminoacridine and D-cycloserine stimulate

acquisition of water maze spatial na rigation in aged rats, Eur J Pharmacol 342(1) (1998) 15-20.

[63] M.G. Baxter, T.H. Lanthorn, . M. Frick, S. Golski, R.Q. Wan, D.S. Olton, D-cycloserine, a novel cognitive enhancer, improves spatial memory in aged rats, Neurobiol Aging 15(2) (1994) 207-13.

[64] A.W. Krug, K. Volker, W. Dantzler, S. Silbernagl, Why is D-serine nephrotoxic and alphaaminoisobutyric acid protective, Am J Physiol Renal Physiol 293(1) (2007) F382-90.

[65] R.E. Williams, E.A. Lock, D-serine-induced nephrotoxicity: possible interaction with tyrosine metabolism, Toxicology 201, 1-3) (2004) 231-8.

[66] N. Villain, B. Dubois, Alzheimer's Disease Including Focal Presentations, Semin Neurol 39(2) (2019) 213-226.

[67] H.W. Kessels, S. Nabavi, R. Malinow, Metabotropic NMDA receptor function is required for betaamyloid-induced synaptic depression, Proc Natl Acad Sci U S A 110(10) (2013) 4033-8.

[68] R. Malinow, New developments on the role of NMDA receptors in Alzheimer's disease, Curr Opin Neurobiol 22(3) (2012) 559-63.

[69] S.I. Mota, I.L. Ferreira, A.C. Rego, Dysfunctional synapse in Alzheimer's disease - A focus on NMDA receptors, Neuropharmacology 76 Pt A (2014) 16-26.

[70] J. Brito-Moreira, A.C. Paula-Lima, T.R. Bomfim, F.B. Oliveira, F.J. Sepulveda, F.G. De Mello, L.G. Aguayo, R. Panizzutti, S.T. Ferreira, Abeta oligomers induce glutamate release from hippocampal neurons, Curr Alzheimer Res 8(5) (2011) 552-62.

[71] M. Talantova, S. Sanz-Blasco, X. Zhang, P. Xia, M.W. Akhtar, S. Okamoto, G. Dziewczapolski, T. Nakamura, G. Cao, A.E. Pratt, Y.J. Kang, S. Tu, E. Molokanova, S.R. McKercher, S.A. Hires, H. Sason, D.G. Stouffer, M.W. Buczynski, J.P. Solomon, S. Michael, E.T. Powers, J.W. Kelly, A. Roberts, G. Tong, T. Fang-Newmeyer, J. Parker, E.A. Holland, D. Zhang, N. Nakanishi, H.S. Chen, H. Wolosker, Y. Wang,

L.H. Parsons, R. Ambasudhan, E. Masliah, S.F. Heinemann, J.C. Pina-Crespo, S.A. Lipton, Abeta induces astrocytic glutamate release, extrasynaptic NMDA receptor activation, and synaptic loss, Proc Natl Acad Sci U S A 110(27) (2013) E2518-27.

[72] E.A. Biemans, N.M. Verhoeven-Duif, J. Gerrits, J.A. Claassen, H.B. Kuiperij, M.M. Verbeek, CSF dserine concentrations are similar in Alzheimer's disease, other dementias, and elderly controls, Neurobiol Aging 42 (2016) 213-6.

[73] M.L. Chouinard, D. Gaitan, P.L. Wood, Presence of the N-methyl-D-aspartate-associated glycine receptor agonist, D-serine, in human temporal cortex: comparison of normal, Parkinson, and Alzheimer tissues, J Neurochem 61(4) (1993) 1561-4.

[74] K. Hashimoto, T. Fukushima, E. Shimizu, S. Okada, N. Komatsu, N. Okamura, K. Koike, H. Koizumi, C. Kumakiri, K. Imai, M. Iyo, Possible role of D-serine in the pathophysiology of Alzheimer's disease, Prog Neuropsychopharmacol Biol Psychiatry 28(2) (2004) 385-8.

[75] S. Kumashiro, A. Hashimoto, T. Nishikawa, Free D-serine in post-mortem brains and spinal cords of individuals with and without neuropsychiatric diseases, Brain Res 581(1-2) (1995) 117-25.

[76] C.H. Lin, C.C. Chiu, C.H. Huang, H.T. Yang, H.Y. Lane, pLG72 levels increase in early phase of Alzheimer's disease but decrease in late phase, Sci Rep 9(1) (2019) 132-1.

[77] C.H. Lin, H.T. Yang, C.C. Chiu, H.Y. Lane, Blood levels of D-amine and oxidase vs. D-amino acids in reflecting cognitive aging, Sci Rep 7(1) (2017) 14849.

[78] C. Madeira, M.V. Lourenco, C. Vargas-Lopes, C.K. Suen nto, C.O. Brandao, T. Reis, R.E. Leite, J. Laks, W. Jacob-Filho, C.A. Pasqualucci, L.T. Grinberg, S.T. Prreira, R. Panizzutti, d-serine levels in Alzheimer's disease: implications for novel biomarker development, Translational psychiatry 5 (2015) e561.

[79] Y. Nagata, M. Borghi, G.H. Fisher, A. D'Anie Ic, Tree D-serine concentration in normal and Alzheimer human brain, Brain Res Bull 38(2) (1995) 181-3.

[80] G. Fisher, N. Lorenzo, H. Abe, E. Fujita, V. H. Fre, C. Emory, M.M. Di Fiore, D.A. A, Free D- and Lamino acids in ventricular cerebrospinal fluic from Alzheimer and normal subjects, Amino Acids 15(3) (1998) 263-9.

[81] D.T. Balu, H. Pantazopoulos, C.C.Y. Hunng, K. Muszynski, T.L. Harvey, Y. Uno, J.M. Rorabaugh, C.R. Galloway, C. Botz-Zapp, S. Berret a, D. Weinshenker, J.T. Coyle, Neurotoxic astrocytes express the d-serine synthesizing enzyme, serine recemase, in Alzheimer's disease, Neurobiol Dis 130 (2019) 104511.

[82] Z. Yan, Z. Zhou, Q. Wu, Z. B. Chen, E.H. Koo, S. Zhong, Presymptomatic Increase of an Extracellular RNA in Blood Plasma Associates with the Development of Alzheimer's Disease, Curr Biol 30(10) (2020) 1771-1782.e3

[83] M. Yamasaki, K. Yama Ia, S. Furuya, J. Mitoma, Y. Hirabayashi, M. Watanabe, 3-Phosphoglycerate dehydrogenase, a key entryme for I-serine biosynthesis, is preferentially expressed in the radial glia/astrocyte lineage and Alfactory ensheathing glia in the mouse brain, J Neurosci 21(19) (2001) 7691-704.

[84] J. Le Douce, M. Maugard, J. Veran, M. Matos, P. Jégo, P.A. Vigneron, E. Faivre, X. Toussay, M. Vandenberghe, Y. Balbastre, J. Piquet, E. Guiot, N.T. Tran, M. Taverna, S. Marinesco, A. Koyanagi, S. Furuya, M. Gaudin-Guérif, S. Goutal, A. Ghettas, A. Pruvost, A.P. Bemelmans, M.C. Gaillard, K. Cambon, L. Stimmer, V. Sazdovitch, C. Duyckaerts, G. Knott, A.S. Hérard, T. Delzescaux, P. Hantraye, E. Brouillet, B. Cauli, S.H.R. Oliet, A. Panatier, G. Bonvento, Impairment of Glycolysis-Derived I-Serine Production in Astrocytes Contributes to Cognitive Deficits in Alzheimer's Disease, Cell metabolism 31(3) (2020) 503-517.e8.

[85] Z. Li, Y. Xing, X. Guo, Y. Cui, Development of an UPLC-MS/MS method for simultaneous quantitation of 11 d-amino acids in different regions of rat brain: Application to a study on the associations of d-amino acid concentration changes and Alzheimer's disease, J Chromatogr B Analyt Technol Biomed Life Sci 1058 (2017) 40-46.

[86] Y. Xing, X. Li, X. Guo, Y. Cui, Simultaneous determination of 18 D-amino acids in rat plasma by an ultrahigh-performance liquid chromatography-tandem mass spectrometry method: application to

explore the potential relationship between Alzheimer's disease and D-amino acid level alterations, Analytical and bioanalytical chemistry 408(1) (2016) 141-50.

[87] S. Wu, A.S. Basile, S.W. Barger, Induction of serine racemase expression and D-serine release from microglia by secreted amyloid precursor protein (sAPP), Curr Alzheimer Res 4(3) (2007) 243-51.

[88] S.Z. Wu, A.M. Bodles, M.M. Porter, W.S. Griffin, A.S. Basile, S.W. Barger, Induction of serine racemase expression and D-serine release from microglia by amyloid beta-peptide, J Neuroinflammation 1(1) (2004) 2.

[89] S. Li, Y. Uno, U. Rudolph, J. Cobb, J. Liu, T. Anderson, D. Levy, D.T. Balu, J.T. Coyle, Astrocytes in primary cultures express serine racemase, synthesize d-serine and acquire A1 reactive astrocyte features, Biochem Pharmacol 151 (2018) 245-251.

[90] R. Inoue, K. Hashimoto, T. Harai, H. Mori, NMDA- and beta-amyloid1-42-induced neurotoxicity is attenuated in serine racemase knock-out mice, J Neurosci 28(53) (2008) 14486-91.

[91] S. Neame, H. Safory, I. Radzishevsky, A. Touitou, F. Marchesani, M. Marchetti, S. Kellner, S. Berlin, V.N. Foltyn, S. Engelender, J.M. Billard, H. Wolosker, The NMDA receptor activation by d-serine and glycine is controlled by an astrocytic Phgdh-dependent service shuttle, Proc Natl Acad Sci U S A 116(41) (2019) 20736-20742.

[92] S.M. de la Monte, M. Tong, Brain metabolic dysfunction at the core of Alzheimer's disease, Biochem Pharmacol 88(4) (2014) 548-59.

[93] C. Zou, S. Crux, S. Marinesco, E. Montagna, C. Sgobio, Y. Shi, S. Shi, K. Zhu, M.M. Dorostkar, U.C. Muller, J. Herms, Amyloid precursor protein maintair sconstitutive and adaptive plasticity of dendritic spines in adult brain by regulating D-serine homeost usis, Embo j 35(20) (2016) 2213-2222.

[94] J.P. Mothet, J.M. Billard, L. Pollegioni, J.T. Coyle, J.V. weedler, Investigating brain d-serine: advocacy for good practices, Acta physiologica (Oxfc rd, England) (2019) e13257.

[95] E. Drummond, T. Wisniewski, Alzheimer's discase: experimental models and reality, Acta neuropathologica 133(2) (2017) 155-175.

[96] M. Cuomo, S. Keller, D. Punzo, T. Nuzzo, \bigcirc Affinito, L. Coretti, M. Carella, V. de Rosa, E. Florio, F. Boscia, V.E. Avvedimento, S. Cocozza, F. Errico, A. Usiello, L. Chiariotti, Selective demethylation of two CpG sites causes postnatal activation of the Dao gene and consequent removal of D-serine within the mouse cerebellum, Clinical ϵ oir, ϵ zetics 11(1) (2019) 149.

[97] M. Audrain, R. Fol, P. Dutar, B. Potier, J.M. Billard, J. Flament, S. Alves, M.A. Burlot, G. Dufayet-Chaffaud, A.P. Bemelmans, J. Valetre, P. Hantraye, N. Deglon, N. Cartier, J. Braudeau, Alzheimer's disease-like APP processing in w. d-type mice identifies synaptic defects as initial steps of disease progression, Molecular neurodegeneration 11 (2016) 5.

[98] M. Audrain, B. Souchet S. Ares, R. Fol, A. Viode, A. Haddjeri, S. Tada, N.S. Orefice, C. Josephine, A.P. Bemelmans, T. Delzes aux N. Deglon, P. Hantraye, Y. Akwa, F. Becher, J.M. Billard, B. Potier, P. Dutar, N. Cartier, J. Brandeau, betaAPP Processing Drives Gradual Tau Pathology in an Age-Dependent Amyloid Rat Model of Alzheimer's Disease, Cereb Cortex 28(11) (2018) 3976-3993.

Table 1: Comparison of the different clinical and preclinical studies aimed at measuring Dserine contents in brain tissues, blood or cerebrospinal fluid (CSF). AD: Alzheimer's disease, MCI: Mild cognitive impairment, MMSE: mini-mental state examination, HPC: hippocampus, PCF: Prefrontal cortex

AL	Ref n°	Subjects	AD MMSE	Sample	Cerebral region	D-serine level
	79	AD		Tissues	PFC	=
	75	AD		Tissues	PCF, parietal cortex	=

	73	AD		Tissues	Temporal cortex	=
	78	AD	12.7 (+/-6.2)	Tissues	HPC and parietal cortex	7
		MCI, mild, moderate to severe AD		CSF		7
	80	AD		CSF		7
	72	AD	22.1 (+/-3.3)	CSF		=
	74	AD	18.7 (+/- 5.00)	Serum		7
	76	Amnestic MCI, mild AD, moderate AD and severe AD		Serum		7
	77	MCI, mild, moderate to severe AD	23.2, 18.2, 10.4, 28.1	Peripheral blood	\mathbf{O}	7
PRECLINICAL DATA	Ref n°	Model	Age	Sample	Cerebral region	D-serine level
	85	Wistar rats with Aβ injections		Tisucs	Olfactory bulb, cerebellum	=
				7	HPC, cerebral cortex	7
	78	APPSwe/PS1∆E9 mice	14-16 menths	Tissues	HPC	7
		C57Bl/6 mice with Aβo injections	C months	Tissues	HPC	7
	84	3xTg-AD	(, n onths	Tissues	HPC	7
	86	Wistar rats with Ap injections		Plasma		У

Highlights:

- D-serine levels decr ase in physiological aging
- In Alzheimer's disease, changes in D-serine levels remain unclear
- The direction of changes may depend on the stage of the disease



