



HAL
open science

In vitro effects of glyphosate-based herbicides and related adjuvants on primary culture of hemocytes from *Haliotis tuberculata*

Antoine Mottier, Antoine Serpentini, Lorna Dallas, Adèle James, Jean-Marc Lebel,
Katherine Costil

► To cite this version:

Antoine Mottier, Antoine Serpentini, Lorna Dallas, Adèle James, Jean-Marc Lebel, et al.. In vitro effects of glyphosate-based herbicides and related adjuvants on primary culture of hemocytes from *Haliotis tuberculata*. *Fish and Shellfish Immunology*, 2020, 100, pp.1-8. <10.1016/j.fsi.2020.02.058>. <hal-02904691>

HAL Id: hal-02904691

<https://normandie-univ.hal.science/hal-02904691v1>

Submitted on 22 Aug 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons CC BY-NC 4.0 - Attribution - Non-commercial use - International License

1 ***In vitro* effects of glyphosate-based herbicides and related adjuvants on primary culture**
2 **of hemocytes from *Haliotis tuberculata***

3

4 Antoine Mottier^{a,b}, Antoine Serpentine^{a,b}, Lorna Dallas^c, Adèle James^{a,b}, Jean-Marc Lebel^{a,b}, and
5 Katherine Costil^{a,b*}

6

7 ^a Normandie Université, Université de Caen Normandie, F-14032 Caen, France

8 ^b BOREA (Biologie des Organismes et des Ecosystèmes Aquatiques), UCN, MNHN, UPMC,
9 CNRS-7208, IRD-207, IBFA, Université de Caen Normandie, Esplanade de la Paix, CS 14032,
10 14032 Caen Cedex 5, France

11 ^c School of Biological Sciences, University of Plymouth, Drake Circus, Plymouth PL4 8AA,
12 UK

13

14 *corresponding author

15 E-mail address: katherine.costil@unicaen.fr (K. Costil)

16

17 **Declarations of interest:** none

18

19 **Keywords:** hemocytes; *Haliotis tuberculata*; glyphosate; POEAs; viability assay;
20 phagocytosis; neutral red retention assay.

21

22 **Abstract**

23 Glyphosate-based herbicides are among the most produced and widely-used herbicides. Studies
24 have shown that commercial formulations and adjuvants may be more toxic to non-target
25 organisms than the active ingredients alone, but the mechanisms of action of these chemicals
26 remain unclear. The aim of this study was to investigate the *in vitro* effects of glyphosate, a
27 commercial formulation and adjuvant alone using primary culture of hemocytes from the
28 European abalone *Haliotis tuberculata*, a commonly farmed shellfish. Glyphosate was found to
29 have negligible effects on viability, phagocytic activities and lysosome stability even with very
30 high doses (i.e. 100 mg L⁻¹). By contrast, greater effects on viability were observed for the
31 commercial formulation and adjuvant alone, with EC₅₀ values of 41.42 mg L⁻¹ and 1.85 mg L⁻¹,
32 respectively. These results demonstrate that the toxic sublethal effects (i.e. phagocytic activity
33 and destabilization of lysosomal membranes) of formulated glyphosate came from adjuvants
34 and suggest they may be related to cell and organelle membrane destabilization.

35 **1. Introduction**

36 The quality of coastal waters greatly depends on human activities in the upstream areas.
37 Among the various contaminants that may be relevant, pesticides from agricultural and
38 domestic activities can be carried by freshwater *via* run-off and leaching processes and
39 subsequently contaminate marine coastal areas [1-3]. Herbicide formulations containing
40 glyphosate as the active ingredient are among the most commonly used pesticides in the world
41 [4]. In these commercial formulations, adjuvants are used to promote the penetration of the
42 active ingredient into plant cuticles [5]. The most common adjuvants in glyphosate-based
43 herbicides are polyethoxylated tallow amines (POEAs) that can be formulated in an
44 oxide:tallow-amine ratio ranging from 5:1 to 25:1. The half-life of POEAs in water has been
45 estimated from 21 to 42 days suggesting that this adjuvant is relatively persistent in water [6].

46 For glyphosate itself, hydrolysis (>30 days for pH ranging from 5 to 9) and photolysis times in
47 water (69 and 77 days, for pH of 7 and 9, respectively) also reveal its relative persistence (e.g.
48 [7]). Furthermore, studies have shown that glyphosate is detectable in rivers [8-11] with
49 maximum concentrations higher than $100 \mu\text{g L}^{-1}$ [12], in contrast to around $1 \mu\text{g L}^{-1}$ in coastal
50 waters [3,13]. Data from literature have indicated that commercial formulations appeared more
51 toxic for a large panel of non-target organisms compared to glyphosate alone [4,14-16] which
52 may indicate toxicity of adjuvants alone or additive or synergistic toxicity of adjuvants with
53 other(s) component(s) of the formulated compounds. Indeed, the toxicity of POEAs has already
54 been demonstrated in different taxonomic groups such as amphibians [4], freshwater
55 crustaceans [17,18] and molluscs [19,20]. For example, the embryotoxicity of POEAs was
56 quantified after 36 h of exposure considering both arrested development and abnormalities in
57 D-shaped larvae of the Pacific oyster *Crassostrea gigas* [20]; the results suggested that POEAs
58 could be considered very toxic to embryo larval development according to the European
59 toxicity classification [6].

60 The abalone, *Haliotis tuberculata*, is a marine gastropod species, which can be found in the
61 Northeast Atlantic from Senegal to Ireland. Abalone have been used as sensitive species to
62 assess pollution of coastal areas [21,22] or the potential toxicity of chemical compounds [23-
63 26]. In molluscs, the hemocytes are key components of the immune system, responsible for
64 various mechanisms of defense, such as phagocytosis, pathogen hydrolysis or Reactive Oxygen
65 Species (ROS) production [27-30]. In bivalves, such as *Crassostrea gigas*, many types of
66 hemocytes have been described, including eosinophilic, basophilic and an intermix between
67 granulocytes, vesicular and blast-like cells [31]. By contrast in *H. tuberculata*, Travers et al.
68 [30] have described only one type of hemocytes, i.e. hyalinocytes which can be separated into
69 to two sub-types: blast-like and large cells. As hemocytes play an essential role in mollusc
70 immunity, the effects of contaminants on these cells could lead to adverse effects for the whole

71 animal [32,33]. Indeed, experiments conducted in the gastropod *Biomphalaria glabrata* by de
72 Monte et al [34] showed that infection with the platyhelminth *Echinostoma paraensei* caused a
73 decrease of circulating hemocytes, and also that association between infection and exposure to
74 the Roundup[®] (concentration equivalent to 36 mg L⁻¹ of glyphosate) greatly increased the
75 percentage of non-viable cells, making the snails more vulnerable to parasitic infections.

76 *In vitro* studies are useful tools to assess the potential risks induced by anthropogenic
77 contaminants in the aquatic environments. Indeed, these tools provide good alternatives to
78 animal experimentation and take ethical issues into consideration [35]. *In vitro* methodologies
79 also allow assessment of the effects of multiple contaminant concentrations on the cells of a
80 limited number of individuals, thereby reducing variability, and are easily reproducible.
81 Although the use of *in vitro* tools gives a limited view of the physiological processes that occur
82 at the *in vivo* level, cell culture provides precious information on the mechanisms of toxicity
83 [24]. While several studies have been published which focused on the effects of various
84 contaminants on bivalve hemocytes *in vitro* (e.g. [36-38]), few ecotoxicological investigations
85 have been performed on *Haliotis spp.* hemocytes. However, in the 2010s, these limited numbers
86 of studies have increased, including those demonstrating the adverse effects of metals [24,26],
87 antibacterial agents [23] and antidepressants [39] on hemocytes in *H. tuberculata*. However,
88 there is still a lack of data concerning the effects of herbicides on the hemocytes of gastropods
89 including *H. tuberculata*.

90 The aims of this study were to assess the *in vitro* effects of (1) glyphosate; (2) a
91 commercial formulation containing glyphosate as the active ingredient (Roundup Express[®]);
92 and (3) POEA adjuvants on hemocytes; in *H. tuberculata* after 72h exposure by using 3
93 established biomarkers: viability assessment (MTT assay), phagocytosis (fluorescent beads)
94 and lysosomal stability (neutral red retention assay). Phagocytosis by hemocytes is the
95 cornerstone of the molluscan immune system [40]. Lysosomes, cellular organelles, are essential

96 components of the humoral immune response in mollusc species. Lysosomes content is released
97 after phagocytosis in order to digest foreign material.

98

99 2. Materials and methods

100 2.1. Hemocyte primary culture

101 Adult abalone (8-10 cm) were bred and provided by France Haliotis® (Plougerneau,
102 France). The animals were maintained in the Centre de Recherche en Environnement Côtier
103 (CREC; Luc-sur-Mer, Normandy, France) in natural sea water with aeration and *ad libitum*
104 algae supply (*Laminaria sp.* and *Palmaria sp.*) for a minimum of 2-weeks acclimation.

105 Primary cell culture of abalone hemocytes has been previously described [24,26,41,42].
106 Briefly, hemocytes were sampled from the adductor muscle of *H. tuberculata*. Hemolymph was
107 withdrawn from a medio-lateral incision using a syringe fitted with a 25 gauge needle. In order
108 to avoid any cell aggregation, the syringe was moisturized with an Alsever solution (115 mM
109 glucose; 27 mM sodium citrate; 11.5 mM EDTA; 382 mM NaCl). Hemolymph was transferred
110 to a 15 mL centrifuge tube and diluted 1:4 with Alsever solution. Hemocytes were counted in
111 triplicate by using a Thoma cell counting chamber. Cells were plated in 12-well culture plates
112 (NUNC®; Penfield, New York, USA) at a density of 5×10^5 cells per well (MTT assay and
113 phagocytosis analyses) or in 96-well culture plates (neutral red retention assay) at a density of
114 1×10^5 cells per well. Hemolymph was diluted 1:4 (v/v) with sterile artificial sea water
115 (ASSW). After 1 h, ASSW was removed and replaced with 500 μ L (12-well plates) or 200 μ L
116 (96-well plates) of sterile modified Hank's 199 medium (250 mM NaCl, 10 mM KCl, 25 mM
117 MgSO₄, 2.5 mM CaCl₂ and 10 mM HEPES, 2 mM l-glutamine, 100 μ g mL⁻¹ streptomycin, 60
118 μ g mL⁻¹ penicillin G and 2 mM concanavalin; pH: 7.4). Cells were incubated for 24 h before

119 beginning any pesticide exposure. Cells were then exposed for 72 h to the different chemicals
120 and all the cultures were performed in an incubator at 17°C without extra CO₂.

121 2.2. Exposures to chemicals

122 In the present study, glyphosate acid (97% purity, CAS number: 1071-83-6) and the
123 POEA mixture were obtained from Dr. Ehrenstorfer GmbH® (Augsburg, Germany) whereas
124 Roundup Express® (R_{EX}) was purchased from a garden centre. For R_{EX}, all the concentrations
125 given in this study were expressed in glyphosate equivalents. All solutions of chemical
126 compounds used were prepared with sterile Hank's M199 medium. Three different endpoints
127 were studied: viability (MTT assay), lysosomal stability (neutral red retention assay) and
128 phagocytic activities (fluorescent beads). The tested concentrations are provided as
129 supplementary data (S1 Table). For the MTT assay, all the concentrations (from 0.1 to 100,000
130 µg L⁻¹) were tested at least with the cells of four abalones and in triplicate (N ≥ 12). For neutral
131 red retention assay (NRRA) and phagocytic activities the tested concentrations were chosen
132 according to previous MTT results for R_{EX} and POEAs: No Observed Effect Concentration
133 (NOEC) observed from MTT assay, EC₂₀ calculated from MTT, EC₅₀ calculated from MTT and
134 finally EC₈₀ calculated from MTT. Glyphosate tested concentrations for NRRA and phagocytic
135 activities corresponded to one low and two high concentrations. For the NRRA and the
136 phagocytic activities, all the concentrations were tested at least with the cells of three abalones
137 and in triplicate (N ≥ 9).

138 2.3. Studied endpoints

139 2.3.1. MTT assay

140 Cell viability was estimated using the 3-[4,5-dimethylthiazol-2-yl]-2,5-
141 diphenyltetrazolium bromide (MTT) reduction assay. This test was adapted to molluscan cell
142 cultures by Domart-Coulon et al. [43]. It measures the capacity of mitochondrial succinyl

143 dehydrogenase in living cells to convert MTT (yellow) into formazan (dark blue). Briefly, 50
144 μL of a stock solution (5 mg mL^{-1} MTT in PBS) was added to culture plates (10% v/v). After
145 24 h incubation, $50 \mu\text{L}$ of acidified isopropanol (0.04 N HCl) was added to each well in order to
146 dissolve neo-formed formazan. Absorbance was measured at 570 nm with a 630 nm reference,
147 and results were expressed as percentages of viability relative to absorbance of the negative
148 control group.

149 2.3.2. Phagocytic activity

150 The phagocytic rate of hemocytes was assessed by flow cytometry as described
151 previously [24,26,39]. Briefly, the medium was removed and replaced with $500 \mu\text{L}$ of pesticide-
152 free medium containing fluorescent latex beads (100 carboxylate-modified
153 FluoroSpheres[®]/hemocyte, yellow-green fluorescence, $1 \mu\text{m}$ diameter, Molecular Probes[®]).
154 After 1 h incubation, the medium was removed and cells were rinsed and then gently scraped
155 into $500 \mu\text{L}$ MPS (molluscan physiological saline). The hemocyte samples were centrifuged for
156 10 min at 500 g and the resulting pellet was delicately fixed in $300 \mu\text{L}$ of 3% formaldehyde for
157 further analysis. Hemocytes were analyzed by using a Gallios[™] flow cytometer (Beckman
158 Coulter[®]). A minimum of $20,000$ events was considered for each sample. The level of
159 fluorescence was evaluated using FL1 channel. The percentage of phagocytic cells was
160 evaluated as the percentage of hemocytes that had engulfed at least three beads (i.e.
161 immunoefficiency).

162 2.3.3. Neutral red retention assay (NRRA)

163 The stability of lysosomal membranes was assessed by the neutral red retention assay
164 (NRRA) as previously described [39] with modifications. Hank's 199 medium was removed
165 and replaced by $300 \mu\text{L}$ of neutral red working solution (2.10^{-2} M neutral red in MPS; 400 mM
166 NaCl ; 100 mM MgSO_4 ; 20 mM HEPES; 10 mM CaCl_2 ; 10 mM KCl). After 3 h of incubation

167 in the dark, neutral red solution was removed, cells were gently rinsed with MPS and 200 μL
168 of elution solution (1:50:49 v/v/v of acetic acid, absolute ethanol, ultrapure water) was added
169 to each well. Plates were then gently agitated for 30 min in the dark. Finally, the optical density
170 of each well was read using a multiplate reader (FlexStation 3[®], Molecular Devices LLC.;
171 Chicago, USA) at 540 and 650 nm as a reference.

172 2.4. Statistical analyses

173 As the data from MTT assay and neutral red retention do not meet the assumption of
174 normality and homoscedasticity for an ANOVA, these data were analyzed using non-parametric
175 Kruskal-Wallis (K-W) tests ($k > 2$) or Mann-Whitney tests ($k = 2$) for independent samples. In
176 case of H_0 rejection after a K-W test, *post-hoc* Dunn tests were used in order to detect
177 differences among the different concentrations. The data for phagocytic activities and NRRA
178 fulfilled requirements for analysis by one way ANOVAs followed by *a posteriori* Student-
179 Newman-Keuls (SNK) tests or t-tests ($k = 2$). The statistical analyses were performed using
180 STATISTICA 8.0 software (Statsoft[®], Tulsa, OK, USA). EC_{50} values were computed with non-
181 linear regressions (Hill equation) using Excel[®] macro REGTOX [44].

182

183 3. Results

184 3.1. MTT viability assay

185 Glyphosate did not induce any significant decrease in MTT activities even at very high
186 concentrations (100,000 $\mu\text{g L}^{-1}$) (Fig. 1A). No differences from the negative control were found
187 for R_{EX} concentrations lower than 20,000 $\mu\text{g L}^{-1}$ (Fig. 1B). However, exposure to 40,000 $\mu\text{g L}^{-1}$
188 R_{EX} caused a highly significant decrease of MTT values ($p < 0.001$) to 52.30% (± 18.78) of
189 the control viability. The viability declined to 6.84% (± 4.75) of the control viability at 100,000
190 $\mu\text{g L}^{-1}$. The concentration leading to 50% mortality (EC_{50}) was 41,420 $\mu\text{g L}^{-1}$. After POEA

191 exposure a significant ($p < 0.001$) decrease of hemocyte viability was observed from 1280 μg
192 L^{-1} ($67.27 \pm 13.74\%$ of the control viability) to 6400 μg L^{-1} ($3.42 \pm 1.93\%$ of the control
193 viability) (Fig. 1C). Finally, an EC_{50} of 1855 μg L^{-1} was calculated for exposures to POEAs.

194 3.2. Phagocytic activity

195 Under our experimental conditions, the percentage of hemocytes that engulfed three
196 beads or more was 33.90 ± 7.91 for the controls. After glyphosate exposure, no significant
197 changes in phagocytic activities were observed (Fig. 2A). Phagocytic activities significantly
198 decreased ($p < 0.001$) after exposures to R_{EX} from 10,000 (NOEC MTT) to 68,000 μg L^{-1} (EC_{80}
199 MTT) (Fig. 2B). Interestingly, the NOEC MTT concentration decreased phagocytic activity
200 ($53.52 \% \pm 16.43$ of the control phagocytosis). Exposure to POEAs led to a significant decrease
201 ($p < 0.001$) in hemocytes phagocytosis at concentrations of 1024 μg L^{-1} ($59.76 \% \pm 15.57$ of
202 the control phagocytosis), 1920 μg L^{-1} ($61.09 \% \pm 25.82$ of the control phagocytosis) and 3200
203 μg L^{-1} (53.55%) (Fig. 2C). An overall comparison between the 3 chemicals tested at the NOEC
204 MTT concentrations revealed significant differences ($p < 0.05$) with a more pronounced effect
205 of R_{EX} compared to glyphosate (the concentration of 100,000 being considered as the NOEC
206 MTT; $p < 0.05$) and POEAs ($p < 0.01$) (Fig. 2). Whereas the effects of R_{EX} and POEAs on
207 phagocytosis efficiency significantly differed when exposed at EC_{20} MTT concentrations ($p <$
208 0.05) it was no longer the case when exposed at EC_{50} MTT ($p = 0.49$) and EC_{80} MTT ($p = 0.48$)
209 concentrations.

210 3.3. Neutral red retention assay (NRRA)

211 Low doses of glyphosate (i.e. 0.1 μg L^{-1}) induced a slight but significant increase ($\times 1.29$)
212 of neutral red retention (NRR) in lysosomes compared to control group ($p < 0.05$). In contrary,
213 the two highest doses of glyphosate (10,000 μg L^{-1} and 100,000 μg L^{-1}) did not lead to any
214 significant changes in lysosomal stability (Fig. 3A) ($p = 0.0256$). A trend toward an increase of

215 neutral red retention in lysosomes was also observed in hemocytes exposed to the lowest doses
216 of R_{EX}[®] (i.e. 0.1 µg L⁻¹) but was not statistically significant (Fig. 3B). Nevertheless, at higher
217 doses of R_{EX}[®], NRR values were significantly ($p < 0.001$) lower compared to the control and
218 ranged from 12.03% (± 4.80%) at 10,000 µg L⁻¹ (NOEC MTT) to 2.74% (± 5.81%) at 68,000
219 µg L⁻¹ (EC₈₀ MTT). Finally, POEA exposure did not induce significant modification of neutral
220 red retention up to 1024 µg L⁻¹ (EC₂₀ MTT) compared to the control (Fig. 3C). However, large
221 significant losses of lysosome neutral red were observed at 1920 (EC₅₀ MTT) and 3200 µg L⁻¹
222 (EC₈₀ MTT), with relative values of 1.61% (± 4.37) and 0.54% (± 3.52), respectively. The
223 overall comparison between the 3 chemicals tested at the NOEC MTT or the 3 EC
224 concentrations for NRR led to similar results than those recorded for phagocytic activity: at the
225 NOEC MTT concentrations, a higher effect of R_{EX} compared to glyphosate ($p < 0.001$) and
226 POEAs ($p < 0.001$), and only at EC₂₀ MTT concentrations, a more marked effect of R_{EX} by
227 comparison to POEAs ($p < 0.001$).

228

229 4. Discussion

230 Hemocytes have a predominant role in the immune response in molluscs and adverse
231 effects to these cells could lead to fatal consequences for the whole animal. Effects on
232 hemocytes could be lethal and reflected by biomarkers such as MTT activities or sublethal by
233 affecting the phagocytic activity and/or the lysosomal system.

234 At the range of concentrations tested, glyphosate did not lead to significant changes in
235 hemocyte MTT activities while exposures to 1024 mg L⁻¹ POEAs and 25,000 µg L⁻¹ R_{EX}
236 induced a significant effect of 20% (EC₂₀). Beyond these values, cellular viability sharply
237 decreased and a dose-response curve was recorded. In the freshwater snail *Biomphalaria*
238 *glabrata*, *in vivo* 24h-exposure to Roundup original[®] at the concentration of 36 mg L⁻¹ induced

239 a significant increase of the number of dead hemocytes assessed by trypan blue vital dye
240 exclusion assay [34]. Viability tests based on mitochondrial activity (such as MTT assay) are
241 also sensitive endpoints that have been previously used to assess the toxicity of different
242 chemical compounds in **marine** molluscs. Domart-Coulon et al. [43] have shown the toxicity of
243 Mexel-432[®] (anti-fouling compound) on the heart cells of *Crassostrea gigas* and gill cells from
244 the clam *Ruditapes decussatus*. In *H. tuberculata* hemocytes, viability tests using mitochondrial
245 activities were used to assess the effect of zinc [26], cadmium [24], triclosan [23] and
246 antidepressants [39]. To our knowledge, no data are available on the cytotoxicity of the
247 compounds studied in this work by using *in vitro* mollusc cell cultures. However, *in vitro*
248 toxicity of commercial formulations and adjuvants has been previously reported in different
249 mammalian cell types and gives interesting points of comparison with our results. Mesnage and
250 his collaborators [45] tested the viability of hepatic (HepG2), embryonic (HEK293) and
251 placental (JEG3) cell lines after 24h exposure to glyphosate or different commercial glyphosate-
252 based formulations and adjuvants. These authors have demonstrated the very low toxicity of
253 glyphosate which was non-toxic on Hep G2 cells and slightly toxic on HEK293 and JEG3 cell
254 lines with EC₅₀ values of 19,300 mg L⁻¹ and 11,192 mg L⁻¹, respectively. In our study, the
255 maximum tested concentration (100 mg L⁻¹) of glyphosate had no effect on hemocyte viability.
256 This result was in accordance with the results from Mesnage and collaborators [45] which
257 correspond to concentration values 100 times higher than those tested in this study. Exposure
258 of hemocytes to R_{EX} and POEAs led to the same patterns than those observed by Mesnage et
259 al. [45]: glyphosate-based commercial formulations expressed a higher toxicity compared to
260 glyphosate alone and adjuvants were the most toxic compounds. Furthermore, the EC₅₀ values
261 for abalone hemocytes were comparable with the EC₅₀ values calculated for JEG3 cell line by
262 Mesnage et al. [45]. Indeed, the exposure to Roundup Grands Travaux[®] led to an EC₅₀ of 32
263 mg L⁻¹ (41.42 mg L⁻¹ for abalone hemocytes exposed to R_{EX}) whereas the POE-15 (one of the

264 ethoxylated amine in adjuvant formulations) induced 50% of cell mortality at the concentration
265 of 1 mg L⁻¹ (1.86 mg L⁻¹ for abalone hemocytes exposed to POEAs). Our results and data from
266 the literature suggest that the toxicity of glyphosate-based compounds is not specific to cell
267 types but seems to be similar between mammalian's cell lines and cells from mollusc primary
268 culture. All these results suggest that the toxicity mechanisms are not specific to a particular
269 metabolic pathway but rather act on common targets for different cell types.

270

271 Phagocytosis has been shown to be impaired by a large panel of contaminants [46].
272 Consequently, the phagocytic activity of marine and freshwater bivalves after *in vitro* exposures
273 is a sensitive endpoint to assess the effects of pollutants at sublethal concentrations [38]. After
274 glyphosate exposure, a slight decrease of phagocytosis occurred at the concentration of 100,000
275 µg L⁻¹ while no decreases in hemocyte viability were observed at this concentration. Similarly,
276 inhibition of phagocytic activities was recorded after R_{EX} exposure from a concentration that
277 did not lead to the inhibition of mitochondrial succinyl deshydrogenase (i.e. 10,000 µg L⁻¹). **A**
278 **similar result** was also observed by Bado-Nilles et al. [36] who have shown the inhibition of
279 phagocytosis in *C. gigas* exposed *in vitro* to pyrene and fluorene without any decrease of cell
280 viability. Likewise, Luna-Acosta et al. [47] observed pronounced inhibition of phagocytosis in
281 *C. gigas* hemocytes after *in vivo* exposure of spat to a mixture of pesticides and pharmaceuticals
282 (5 µg L⁻¹ diuron, 5 µg L⁻¹ isoproturon and 5 µg L⁻¹ ibuprofen) but no decrease in cell viability
283 was recorded. In addition, the exposure of abalone hemocytes to clomipramin, citalopram, and
284 paroxetine decreased phagocytosis whereas amitriptylin induced a dose-related increase of
285 phagocytosis [39]. Authors hypothesized that the increase in phagocytosis could be explained
286 by the interaction of amitriptylin with a variety of receptors (e.g. histaminic, cholinergic,
287 serotonin and adrenergic receptors) even if all of these receptors have not been yet evidenced
288 in abalone hemocytes. Such increases were not observed after glyphosate, R_{EX} or POEA

289 exposures suggesting again that the mechanisms of toxicity are not specific to a particular
290 metabolic pathway. Our results on phagocytosis and those reported by the different authors
291 highlight the potential in studying biomarkers such as phagocytosis which could reveal early
292 effects on cells before any loss of viability. In the Chinese crab *Eriocheir sinensis* exposed to a
293 range of glyphosate concentrations (from 4.4 to 98 mg L⁻¹) up to 96h, Hong et al. [48] studied
294 phagocytic activity by observing cells that had incorporated fluorescent beads under an inverted
295 fluorescence microscope. After 6h exposure to all glyphosate concentrations including 4.4 mg
296 L⁻¹, these authors recorded a significant decrease of phagocytic activity that tended to be at the
297 lowest level at 96h. Therefore, the comparison with the results recorded here in *H. tuberculata*
298 (no significant change for exposure to 10 mg L⁻¹) suggest a higher sensibility of the species *E.*
299 *sinensis* and/or *in vivo* exposure. The EC₅₀ values for cellular tests were higher than the
300 corresponding values for *Daphnia magna* and fish, indicating that the cellular tests (i.e. cell
301 viability) are less sensitive than whole organisms [49].

302

303 Lysosomes are able to sequester and accumulate a large number of different
304 contaminants [50]. Literature reported that the lysosomal system could be a target site for the
305 toxic effects of different type of xenobiotics. Pollutants could not only cause lysosome
306 membrane destabilization but also change the structure, the dynamic and the composition of
307 lysosomal system. Neutral red retention (NRR) is considered as a sensitive biomarker of
308 exposure to chemicals and contaminated areas and it has been used in various mollusc species
309 (e.g. [24,51,52]). In the present study, the three chemicals tested showed significant effects on
310 lysosome membrane but in different ways. Although glyphosate did not affect cell mortality at
311 any tested concentration, this molecule seemed to stimulate lysosomal system at low doses
312 suggesting a like-hormesis effect. Such results have been previously observed by various
313 authors: Canty et al. [53] who have reported a significant increase of NRR after 1 h and 24 h in

314 hemocytes of mussels (*Mytilus edulis*) exposed *in vivo* to the organophosphate pesticide
315 azamethiphos; Bado-Nilles et al. [36] in *C. gigas* hemocytes exposed *in vitro* to dibenzo-[a,h]-
316 anthracene, and Braunbeck and Appelbaum [54] in *Cyprinus carpio* intestinal epithelium
317 exposed *in vivo* to ultra-low doses of the insecticide endosulfan. As abalone hemocytes are non-
318 proliferative cells, the increase in NRR can be considered as an increase of the number and/or
319 the size of lysosomes in the exposed hemocytes. Lysosomes size and number increases have
320 been reported in mollusc for a wide range of contaminants (e.g. [55,56]) and could correspond
321 to an unspecific sign of stress after exposure to a contaminant. The R_{EX} and POEA exposures
322 did not induce a significant increase of NRR even if some trends could be observed at the lowest
323 dose of contaminants. **Conversely**, these two chemicals compounds led to a drastic decrease of
324 the NRR from exposures **to NOEC MTT concentration for R_{EX} and to EC₅₀ MTT concentration**
325 **for POEAs**. It is interesting to note that this decrease appeared at R_{EX} concentrations which did
326 not affect the cell viability (i.e. 10,000 µg L⁻¹). Moreover, at the concentration of POEAs that
327 inhibited 50% of cell viability, the NRR value was near zero. **Such effects cannot be explained**
328 **by cell lysis but very probably by a specific effect of R_{EX}, and to lesser extent POEAs, on the**
329 **lysosomal system**. This **type of effect** has been previously observed in the Haliotidae. In
330 *Haliotis rubra* exposed *in vivo* to air, Song et al. [57] have observed the early response of
331 lysosomal system of hemocytes before any mortalities of exposed abalone. In addition, NRR
332 values drastically decreased when hemocytes from *H. tuberculata* were exposed ***in vitro***, for
333 48, h to different antidepressants [39]. More precisely, the decrease was observed at the
334 concentration equivalent to EC₁₀ MTT for amitryptiline, and EC₂₀ MTT for paroxetine and
335 citalopram. Likewise, a reduction in lysosome membrane stability has been reported in mussel
336 and oysters exposed to heavy metal and this response has been proposed as an indicator of cell
337 damage [50].

338

339 It is important to note that the adverse effects of the three chemical compound and
340 mixtures tested in the present study were observed at non-realistic environmental
341 concentrations ($> 1 \mu\text{g L}^{-1}$ for glyphosate). Furthermore, the bioavailability of all the chemicals
342 was maximized by the experimental design because of the direct exposure of targeted cells. The
343 major disadvantage of *in vitro* methods is the difficulty of extrapolating the results to *in vivo*
344 situations but they also have the advantages of being easy to use and reproducible [39].
345 However, the results provided by the three endpoints, and particularly the values of NOEC and
346 EC determined with the MTT assay, allowed us to classify the toxicity of the chemicals as
347 follows: glyphosate $< R_{EX} <$ POEAs and other studies have already reported this ranking (e.g.
348 [4,16,58]). Further studies should also include *in vivo* exposures in *H. tuberculata* and other
349 immune parameters should be investigated. Indeed, parameters such as the hemocyte
350 concentration in hemolymph and the volume of hemocytes should be considered because
351 significant changes have been recorded after *in vivo* exposures to AMPA or glyphosate for 7
352 days, respectively, from concentrations as low as $1 \mu\text{g L}^{-1}$ in *Mytilus galloprovincialis* [59] and
353 $10 \mu\text{g L}^{-1}$ in *R. philippinarum* [60]. From a case study in *M. galloprovincialis* in which both
354 cellular and biochemical parameters had been affected, Matozzo and his collaborators [61]
355 concluded to a potential risk of glyphosate for aquatic invertebrates. By a transcriptional study
356 conducted in the Pacific oyster, Mottier et al. [62] showed that the level of gene expression
357 significantly increased after sub-chronic exposures to glyphosate from $0.1 \mu\text{g L}^{-1}$ (multi-
358 xenobiotic resistance) or $100 \mu\text{g L}^{-1}$ (GST and metallothioneins). In digestive gland from *M.*
359 *galloprovincialis*, Milan et al. [63] demonstrated a significant effect of the exposure to $10 \mu\text{g}$
360 L^{-1} glyphosate for 21 days on the expression of 111 genes including some involved in
361 endoplasmic reticulum stress response.

362 Mechanisms of POEA toxicity remain not fully explained but several authors have
363 suggested that non-ionic surfactants could interact with the lipid bilayer and membrane proteins

364 altering fluidity and oxygen transport [64-66]. The present results also suggest this mode of
365 action since the effects of Roundup® and POEAs seem to be related to the lysosomal membrane
366 stability and cytoplasmic membrane deformation (phagocytosis).

367

368 **5. Conclusion**

369 This study presents the first results on the effects of glyphosate-based herbicides in *H.*
370 *tuberculta* hemocytes and provided important information to compare the toxicity of the active
371 ingredient with one of its commercial formulations and its associated adjuvants. While no effect
372 on cell viability appeared with any tested concentration of glyphosate or at concentrations
373 below 20 mg L⁻¹ of R_{EX}, effects on cell membranes have been suggested at sublethal
374 concentrations, thus clearly demonstrating the utility of multi-biomarker approaches and *in*
375 *vitro* exposures. The adjuvants seemed to be mainly responsible for the toxicity of commercially
376 formulated glyphosate. However, despite the scarcity or lack of data about the contamination
377 of coastal environments, the toxicity of the tested molecules was most likely at much higher
378 level of magnitude of concentrations than those observed in the environment.

379

380 **Acknowledgement**

381 This study was supported by the European Program Interreg IVA 'Chronexpo' (number: 4059)
382 coordinated by Dr. Bruno Fievet from IRSN (Institut de Radioprotection et de Sureté Nucléaire)
383 of Cherbourg-Octeville and by the Region Basse-Normandie (number: 09P00702). The authors
384 thank Maryline Guillamin (IFR 146 ICORE) for her help in flow cytometry and the technical
385 staff of the C.R.E.C. for their assistance in animal care.

386

387 **References**

- 388 [1] Auby, I., Bocquene, G., Quiniou, F., Dreno, J.-P., 2007. Etat de la contamination du bassin
389 d’Arcachon par les insecticides et les herbicides sur la période 2005-2006. Impact
390 environnemental. Ifremer Arcachon, RST/LER/AR/07-003 report
391 <https://archimer.ifremer.fr/doc/00000/2398/>
- 392 [2] S. Buisson, V. Bouchart, E. Guerlet, J.P. Malas, K. Costil, Level of contamination and impact
393 of pesticides in cupped oyster, *Crassostrea gigas*, reared in a shellfish production area in
394 Normandy (France). J. Environ. Sci. Health. B. 43 (2008) 655–664.
395 doi:10.1080/03601230802352732
- 396 [3] T. Burgeot, B. Gagnaire, T. Renault, J. Haure, D. Moraga, E. David, I. Boutet, P.G. Sauriau,
397 N. Malet, V. Bouchet, F. Le Roux, S. Lapègue, K. Bouilly, G. Le Moullac, I. Arzul, J. Knoery,
398 F. Quiniou, C. Bacher, P. Soletchnik, Les risques associés au stress environnemental, in:
399 Samain, J.-F., McCombie, H. (Eds.), Summer Mortality of Pacific Oyster *Crassostrea Gigas*.
400 Editions Quae, Versailles, France, 2008, pp. 95–139
- 401 [4] C.M. Howe, M. Berrill, B.D. Pauli, C.C. Helbing, K. Werry, N. Veldhoen, Toxicity of
402 glyphosate-based pesticides to four north American frog species. Environ. Toxicol. Chem. 23
403 (2004) 1928–1938. doi:10.1897/03-71
- 404 [5] R.A. Relyea, The lethal impact of Roundup® on aquatic and terrestrial amphibians. Ecol.
405 Appl. 15 (2005) 1118–1124. doi:10.1890/04-1291
- 406 [6] J.P. Giesy, S. Dobson, K.R. Solomon, Ecotoxicological risk assessment for Roundup®
407 herbicide. Rev. Environ. Contam. Toxicol. 167 (2000) 35–120. doi.org/10.1007/978-1-4612-
408 1156-3_2
- 409 [7] PPDB, 2019. Pesticide Properties Database (PPDB). (last access: October 2019) [WWW
410 Document].

- 411 [8] H. Blanchoud, E. Moreau-Guigon, F. Farrugia, M. Chevreuil, J.M. Mouchel, Contribution
412 by urban and agricultural pesticide uses to water contamination at the scale of the Marne
413 watershed. *Sci. Total Environ.* 375 (2007) 168–79. doi:10.1016/j.scitotenv.2006.12.009
- 414 [9] F. Botta, G. Lavison, G. Couturier, F. Alliot, E. Moreau-Guigon, N. Fauchon, B. Guery, M.
415 Chevreuil, H. Blanchoud, Transfer of glyphosate and its degradate AMPA to surface waters
416 through urban sewerage systems. *Chemosphere* 77 (2009), 133–139.
417 doi:10.1016/j.chemosphere.2009.05.008
- 418 [10] D.W. Kolpin, E.M. Thurman, E.A. Lee, M.T. Meyer, E.T. Furlong, S.T. Glassmeyer,
419 Urban contributions of glyphosate and its degradate AMPA to streams in the United States.
420 *Sci. Total Environ.* 354 (2006) 191–197. doi:10.1016/j.scitotenv.2005.01.028
- 421 [11] S. Pesce, C. Fajon, C. Bardot, F. Bonnemoy, C. Portelli, J. Bohatier, Longitudinal changes
422 in microbial planktonic communities of a French river in relation to pesticide and nutrient
423 inputs. *Aquat. Toxicol.* 86 (2008) 352–60. doi:10.1016/j.aquatox.2007.11.016
- 424 [12] L. Puértolas, J. Damásio, C. Barata, C., A.M.V.M. Soares, N. Prat, Evaluation of side-
425 effects of glyphosate mediated control of giant reed (*Arundo donax*) on the structure and
426 function of a nearby Mediterranean river ecosystem. *Environ. Res.* 110 (2010) 556–564.
427 doi:10.1016/j.envres.2010.05.004
- 428 [13] W. Skeff, C. Neumann, D.E. Schulz-Bull, Glyphosate and AMPA in the estuaries of the
429 Baltic Sea method optimization and field study. *Mar. Pollut. Bull.* 100 (2015) 577–585.
430 doi:10.1016/j.marpolbul.2015.08.015
- 431 [14] L.C. Folmar, H.O. Sanders, A.M. Julin, Toxicity of the herbicide glyphosate and several of
432 its formulations to fish and aquatic invertebrates. *Arch. Environ. Contam. Toxicol.* 8 (1979)
433 269–278. doi:10.1007/BF01056243
- 434 [15] R.M. Mann, J.R. Bidwell, The toxicity of glyphosate and several glyphosate formulations

- 435 to four species of southwestern Australian frogs. *Arch. Environ. Contam. Toxicol.* 36 (1999)
436 193–199. doi.org/10.1007/s002449900
- 437 [16] A. Mottier, V. Kientz-Bouchart, A. Serpentine, J.M. Lebel, A.N. Jha, K. Costil, Effects of
438 glyphosate-based herbicides on embryo-larval development and metamorphosis in the Pacific
439 oyster, *Crassostrea gigas*. *Aquat. Toxicol.* 128–129 (2013) 67–78.
440 doi:10.1016/j.aquatox.2012.12.002
- 441 [17] J.M. Brausch, P.N. Smith, Toxicity of three polyethoxylated tallowamine surfactant
442 formulations to laboratory and field collected fairy shrimp, *Thamnocephalus platyurus*. *Arch.*
443 *Environ. Contam. Toxicol.* 52 (2007), 217–221. doi:10.1007/s00244-006-0151-y
- 444 [18] J.M. Brausch, B. Beall, P.N. Smith, Acute and sub-lethal toxicity of three POEA surfactant
445 formulations to *Daphnia magna*. *Bull. Environ. Contam. Toxicol.* 78 (2007) 510–514.
446 doi:10.1007/s00128-007-9091-0
- 447 [19] R.B. Bringolf, W.G. Cope, S. Mosher, M.C. Barnhart, D. Shea, Acute and chronic toxicity
448 of glyphosate compounds to glochidia and juveniles of *Lampsilis siliquoidea* (Unionidae).
449 *Environ. Toxicol. Chem.* 26 (2007) 2094–2100. doi: 10.1897/06-519R1.1
- 450 [20] A. Mottier, J. Pini, K. Costil, Effects of a POEA surfactant system (Genamin T-200®) on
451 two life stages of the Pacific oyster, *Crassostrea gigas*. *J. Toxicol. Sci.* 39 (2014) 211–215.
452 doi:10.2131/jts.39.211
- 453 [21] J. Gorski, D. Nugegoda, Toxicity of trace metals to juvenile abalone, *Haliotis rubra*
454 following short-term exposure. *Bull. Environ. Contam. Toxicol.* 77 (2006) 732–40.
455 doi:10.1007/s00128-006-1125-5
- 456 [22] X. Zhu, J. Zhou, Z. Cai, The toxicity and oxidative stress of TiO₂ nanoparticles in marine
457 abalone (*Haliotis diversicolor supertexta*). *Mar. Pollut. Bull.* 63 (2011) 334–8.

458 doi:10.1016/j.marpolbul.2011.03.006

459 [23] B. Gaume, N. Bourgougnon, S. Auzoux-Bordenave, B. Roig, B. Le Bot, G. Bedoux, In
460 vitro effects of triclosan and methyl-triclosan on the marine gastropod *Haliotis tuberculata*.
461 Comp. Biochem. Physiol. Part C Comp. Pharmacol. Toxicol. 156 (2012) 87–94.
462 doi:10.1016/j.cbpc.2012.04.006

463 [24] T. Latire, C. Le Pabic, E. Mottin, A. Mottier, K. Costil, N. Koueta, J.M. Lebel, A.
464 Serpentine, Responses of primary cultured haemocytes from the marine gastropod *Haliotis*
465 *tuberculata* under 10-day exposure to cadmium chloride. Aquat. Toxicol. 109 (2012) 213–221.
466 doi:10.1016/j.aquatox.2011.09.017

467 [25] A. Letullier, L. Minguez, K. Costil, M.P. Halm-Lemeille, J.M. Lebel, A. Serpentine, In vitro
468 effect of five pharmaceuticals on the viability of the European abalone hemocytes, *Haliotis*
469 *tuberculata*. J. Xenobiotics 4 (2014) 78–80 doi:10.4081/xeno.2014.4900

470 [26] E. Mottin, C. Caplat, M.L. Mahaut, K. Costil, D. Barillier, J.M. Lebel, A. Serpentine, Effect
471 of in vitro exposure to zinc on immunological parameters of haemocytes from the marine
472 gastropod *Haliotis tuberculata*. Fish & Shellfish Immunol. 29 (2010) 846–53.
473 doi:10.1016/j.fsi.2010.07.022

474 [27] L. Donaghy, H.K. Hong, H.J. Lee, J.C. Jun, Y.J. Park, K.S. Choi, Hemocyte parameters of
475 the Pacific oyster *Crassostrea gigas* a year after the Hebei Spirit oil spill off the west coast of
476 Korea. Helgol. Mar. Res. 64 (2010) 349–355. doi:10.1007/s10152-010-0190-7

477 [28] T.S. Galloway, M.H. Depledge, Immunotoxicity in invertebrates: measurement and
478 ecotoxicological relevance. Ecotoxicology 10 (2001) 5–23. doi:10.1023/A:1008939520263

479 [29] C. Hooper, R. Day, R. Slocombe, J. Handler, K. Benkendorff, Stress and immune
480 responses in abalone : limitations in current knowledge and investigative methods based on
481 other models. Fish & Shellfish Immunol. 22 (2007) 363–379. doi: 10.1016/j.fsi.2006.06.009

- 482 [30] M.A. Travers, P. Mirella Da Silva, N. Le Goïc, D. Marie, A. Donval, S. Huchette, M.
483 Koken, C. Paillard, Morphologic, cytometric and functional characterisation of abalone
484 (*Haliotis tuberculata*) haemocytes. *Fish & Shellfish Immunol.* 24 (2008) 400–411. doi:
485 10.1016/j.fsi.2007.10.001
- 486 [31] S. Chang, S. Tseng, H. Chou, Morphological characterization via light and electron
487 microscopy of the hemocytes of two bivalves: a comparison study between the hard clam
488 (*Meterix lusoria*) and the Pacific oyster (*Crassostrea gigas*). *Zool. Stud.* 44 (2005) 144–152.
- 489 [32] W. Cheng, I.S. Hsiao, C.H. Hsu, J.C. Chen, The immune response of Taiwan abalone
490 *Haliotis diversicolor supertexta* and its susceptibility to *Vibrio parahaemolyticus* at different
491 salinity levels. *Fish & Shellfish Immunol.* 16 (3) (2004) 295–306. doi:10.1016/S1050-
492 4648(03)00111-6
- 493 [33] W. Cheng, I.S. Hsiao, C.H. Hsu, J.C. Chen, Change in water temperature on the immune
494 response of Taiwan abalone (*Haliotis diversicolor*) and its susceptibility to *Vibrio*
495 *parahaemolyticus*. *Fish Shellfish Immunol.* 17 (3) (2004) 235–243.
496 doi:10.1016/j.fsi.2004.03.007
- 497 [34] T.C.C. Monte, T.Q. Chometon, A.L. Bertho, V. Moura, M.C. Vasconcellos, J. Garcia, R.
498 Ferraz-Nogueirac, A.J. Maldonado, M.J. Faro, Changes in hemocytes of *Biomphalaria glabrata*
499 infected with *Echinostoma paraensei* and exposed to glyphosate-based herbicide. *J. Invert.*
500 *Pathol* 160 (2019) 67-75. doi.org/10.1016/j.jip.2018.11.007
- 501 [35] S. Ní Shúilleabháin, C. Mothersill, D. Sheehan, N.M. O'Brien, J. O' Halloran, F.N. van
502 Pelt, M. Kilemade, M. Davoren, Cellular responses in primary epidermal cultures from rainbow
503 trout exposed to zinc chloride. *Ecotoxicol. Environ. Saf.* 65 (2006) 332–41.
504 doi:10.1016/j.ecoenv.2005.08.004
- 505 [36] A. Bado-Nilles, B. Gagnaire, H. Thomas-Guyon, S. Le Floch, T. Renault, Effects of 16

506 pure hydrocarbons and two oils on haemocyte and haemolymphatic parameters in the Pacific
507 oyster, *Crassostrea gigas* (Thunberg). *Toxicol. Vit.* 22 (2008), 1610–7.
508 doi:10.1016/j.tiv.2008.04.011

509 [37] B. Gagnaire, H. Thomas-Guyon, T. Burgeot, T. Renault, Pollutant effects on Pacific oyster,
510 *Crassostrea gigas* (Thunberg), hemocytes: screening of 23 molecules using flow cytometry.
511 *Cell Biol. Toxicol.* 22 (2006) 1–14. doi:10.1007/s10565-006-0011-6

512 [38] S. Sauvé, P. Brousseau, J. Pellerin, Y. Morin, L. Sénécal, P. Goudreau, M. Fournier,
513 Phagocytic activity of marine and freshwater bivalves: in vitro exposure of hemocytes to metals
514 (Ag, Cd, Hg and Zn). *Aquat. Toxicol.* 58 (2002) 189–200. doi.org/10.1016/S0166-
515 445X(01)00232-6

516 [39] L. Minguéz, M.P. Halm-Lemeille, K. Costil, R. Bureau, J.M. Lebel, A. Serpentine,
517 Assessment of cytotoxic and immunomodulatory properties of four antidepressants on primary
518 cultures of abalone hemocytes (*Haliotis tuberculata*). *Aquat. Toxicol.* 153 (2014) 3–11.
519 doi:10.1016/j.aquatox.2013.10.020

520 [40] R.K. Pipe, J.A. Coles, Environmental contaminants influencing immunefunction in marine
521 bivalve molluscs. *Fish & Shellfish Immunol.* 5 (1995) 581–595. doi:10.1016/S1050-
522 4648(95)80043-3

523 [41] J.M. Lebel, W. Giard, P. Favrel, E. Boucaud-Camou, Effects of different vertebrate growth
524 factors on primary cultures of hemocytes from the gastropod mollusc, *Haliotis tuberculata*.
525 *Biol. Cell* 86 (1996) 67–72. doi.org/10.1016/0248-4900(96)89526-8

526 [42] A. Serpentine, C. Ghayor, J.M. Poncet, V. Hebert, P. Galéra, J.P. Pujol, E. Boucaud-Camou,
527 J.M. Lebel, Collagen study and regulation of the de novo synthesis by IGF-I in hemocytes from
528 the gastropod mollusc, *Haliotis tuberculata*. *J. Exp. Zool.* 287 (2000) 275–284.
529 doi.org/10.1002/1097-010X(20000901)287:4<275::AID-JEZ2>3.0

- 530 [43] I. Domart-Coulon, S. Auzoux-Bordenave, D. Doumenc, M. Khalanski, Cytotoxicity
531 assessment of antibiofouling compounds and by-products in marine bivalve cell cultures.
532 Toxicol. Vitr. 14 (2000) 245–251. doi:10.1016/S0887-2333(00)00011-4
- 533 [44] E. Vindimian, 2016. MSEXcel macro Regtox.7.0.7 freely available from Eric Vindimian,
534 IRSTEA, France. http://www.normalesup.org/~vindimian/fr_index.html (last access: October
535 2019)
- 536 [45] R. Mesnage, B. Bernay, G.E. Séralini, Ethoxylated adjuvants of glyphosate-based
537 herbicides are active principles of human cell toxicity. Toxicology 313 (2-3) (2012) 122-128.
538 doi:10.1016/j.tox.2012.09.006
- 539 [46] M. Auffret, S. Rousseau, I. Boutet, A. Tanguy, J. Baron, D. Moraga, M. Duchemin, A
540 multiparametric approach for monitoring immunotoxic responses in mussels from
541 contaminated sites in Western Mediterranean. Ecotoxicol. Environ. Saf. 63 (2006) 393–405.
542 doi:10.1016/j.ecoenv.2005.10.016
- 543 [47] A. Luna-Acosta, T. Renault, H. Thomas-Guyon, N. Faury, D. Saulnier, H. Budzinski, K.
544 Le Menach, P. Pardon, I. Fruitier-Arnaudin, P. Bustamante, Detection of early effects of a single
545 herbicide (diuron) and a mix of herbicides and pharmaceuticals (diuron, isoproturon, ibuprofen)
546 on immunological parameters of Pacific oyster (*Crassostrea gigas*) spat. Chemosphere 87
547 (2012) 1335–1340. doi:10.1016/j.chemosphere.2012.02.022
- 548 [48] Y. Hong, X. Yang, G. Yan, Y. Huang, F. Zuo, Y. Shen, Y. Ding, Y. Cheng, Effects of
549 glyphosate on immune responses and haemocyte DNA damage of Chinese mitten crab,
550 *Eriocheir sinensis*. Fish & Shellfish Immunol. 71 (2017) 19-27.
551 doi.org/10.1016/j.fsi.2017.09.062

- 552 [49] M. Sandbacka, I. Christianson, B. Isomaa, The acute toxicity of surfactants on fish cells,
553 *Daphnia magna* and fish: a comparative study. *Toxicol. In Vitro*, 14 (2000) 61-68.
554 doi:10.1016/s0887-2333(99)00083-1
- 555 [50] M.N. Moore, J. Icarus Allen, A. McVeigh, Environmental prognostics: an integrated model
556 supporting lysosomal stress responses as predictive biomarkers of animal health status. *Mar.*
557 *Environ. Res.* 61 (2006) 278–304. doi:10.1016/j.marenvres.2005.10.005
- 558 [51] M. Castro, M.M. Santos, N.M. Monteiro, N. Vieira, Measuring lysosomal stability as an
559 effective tool for marine coastal environmental monitoring. *Mar. Environ. Res.* 58 (2004) 741–
560 5. doi:10.1016/j.marenvres.2004.03.088
- 561 [52] A.H. Ringwood, D.E. Conner, A. Dinovob, The Effects of Copper Exposures on Cellular
562 Responses in Oysters. *Mar. Environ. Res.* 46 (1998) 591–595. doi.org/10.1016/S0141-
563 1136(97)00084-6
- 564 [53] M.N. Canty, J. Hagger, R.T.B. Moore, L. Cooper, T.S. Galloway, Sublethal impact of short
565 term exposure to the organophosphate pesticide azamethiphos in the marine mollusc *Mytilus*
566 *edulis*. *Mar. Pollut. Bull.* 54 (2007) 396–402. doi:10.1016/j.marpolbul.2006.11.013
- 567 [54] T. Braunbeck, S. Appelbaum, Ultrastructural alterations in the liver and intestine of carp
568 *Cyprinus carpio* induced orally by ultra-low doses of endosulfan. *Dis. Aquat. Organ.* 36 (1999),
569 183–200. doi:10.3354/dao036183
- 570 [55] L. Giambérini, M.P. Cajaraville, Lysosomal responses in the digestive gland of the
571 freshwater mussel, *Dreissena polymorpha*, experimentally exposed to cadmium. *Environ. Res.*
572 98 (2005) 210–4. doi:10.1016/j.envres.2004.11.003
- 573 [56] I. Marigómez, U. Izagirre, X. Lekube, Lysosomal enlargement in digestive cells of mussels
574 exposed to cadmium, benzo[a]pyrene and their combination. *Comp. Biochem. Physiol. Part C*
575 *Comp. Pharmacol. Toxicol.* 141 (2005) 188–93. doi:10.1016/j.cca.2005.06.005

- 576• [57] L. Song, X. Li, K. Bott, T. Wang, S.M. Clarke, W. Zhao, Effects of air exposure on the
577 lysosomal membrane stability of haemocytes in blacklip abalone, *Haliotis rubra* (Leach).
578 *Aquaculture Res.* 38 (2007) 239-245. doi:10.1111/j.1365-2109.2007.01655.x
- 579 [58] M.T.K. Tsui, L.M. Chu, Aquatic toxicity of glyphosate-based formulations: comparison
580 between different organisms and the effects of environmental factors. *Chemosphere* 52 (2003)
581 1189–1197. doi:10.1016/S0045-6535(03)00306-0
- 582 [59] V. Matozzo, M.G. Marin, L. Masiero, M. Tremonti, S. Biamonte, S. Viale, L. Finos, G.
583 Lovato, P. Pastore, S. Bogialli, Effects of aminomethylphosphonic acid, the main breakdown
584 product of glyphosate, on cellular and biochemical parameters of the mussel *Mytilus*
585 *galloprovincialis*. *Fish and Shellfish Immunol.* 83 (2018) 321–329.
586 doi.org/10.1016/j.fsi.2018.09.036
- 587 [60] V. Matozzo, C. Zampieri, M. Munari, M.G. Marin, Glyphosate affects haemocyte
588 parameters in the clam *Ruditapes philippinarum*. *Mar. Environ. Res.* 146 (2019) 66-70.
589 doi:10.1016/j.marenvres.2019.03.008
- 590 [61] V. Matozzo, J. Fabrello, L. Masiero, F. Ferraccioli, L. Finos, P. Pastore, I.M. Di Gangi, S.
591 Bogialli, Ecotoxicological risk assessment for the herbicide glyphosate to non-target species: a
592 case study with the mussel *Mytilus galloprovincialis*. *Environ. Pollut.* 233 (2018) 623-632. doi:
593 10.1016/j.envpol.2017.10.100
- 594 [62] A. Mottier, A. Séguin, A. Devos, C. Le Pabic, C. Voiseux, J.M. Lebel, A. Serpentine, B.
595 Fievet, K. Costil, Effects of subchronic exposure to glyphosate in juvenile oysters (*Crassostrea*
596 *gigas*): from molecular to individual levels. *Mar. Pollut. Bull.* 95 (2015) 665-677.
597 10.1016/j.marpolbul.2014.10.026

- 598 [63] M. Milan, G. Dalla Rovere, M. Smits, S. Ferraresso, P. Pastore, M.G. Marin, S. Bogialli,
599 T. Patarnello, L. Bargelloni, V. Matozzo, Ecotoxicological effects of the herbicide glyphosate
600 in non-target aquatic species: transcriptional responses in the mussel *Mytilus galloprovincialis*.
601 Environ. Pollut. 237 (2018) 442-451. doi:10.1016/j.envpol.2018.02.049
- 602 [64] T. Cserhádi, Alkyl ethoxylated and alkylphenol ethoxylated nonionic surfactants:
603 interaction with bioactive compounds and biological effects. Environ. Health Perspect. 103
604 (1995) 358-64. doi:10.1289/ehp.95103358
- 605 [65] Å. Lindgren, M. Sjöström, S. Wold, QSAR modelling of the toxicity of some technical
606 non-ionic surfactants towards fairy shrimps. Quant. Struct. Relationships 15 (1996) 208-218.
607 doi:10.1002/qsar.19960150305
- 608 [66] P. Pärt, O. Svanberg, E. Bergström, The influence of surfactants on gill physiology and
609 cadmium uptake in perfused rainbow trout gills. Ecotoxicol. Environ. Saf. 9 (1985) 135-144.
610 doi:10.1016/0147-6513(85)90016-8

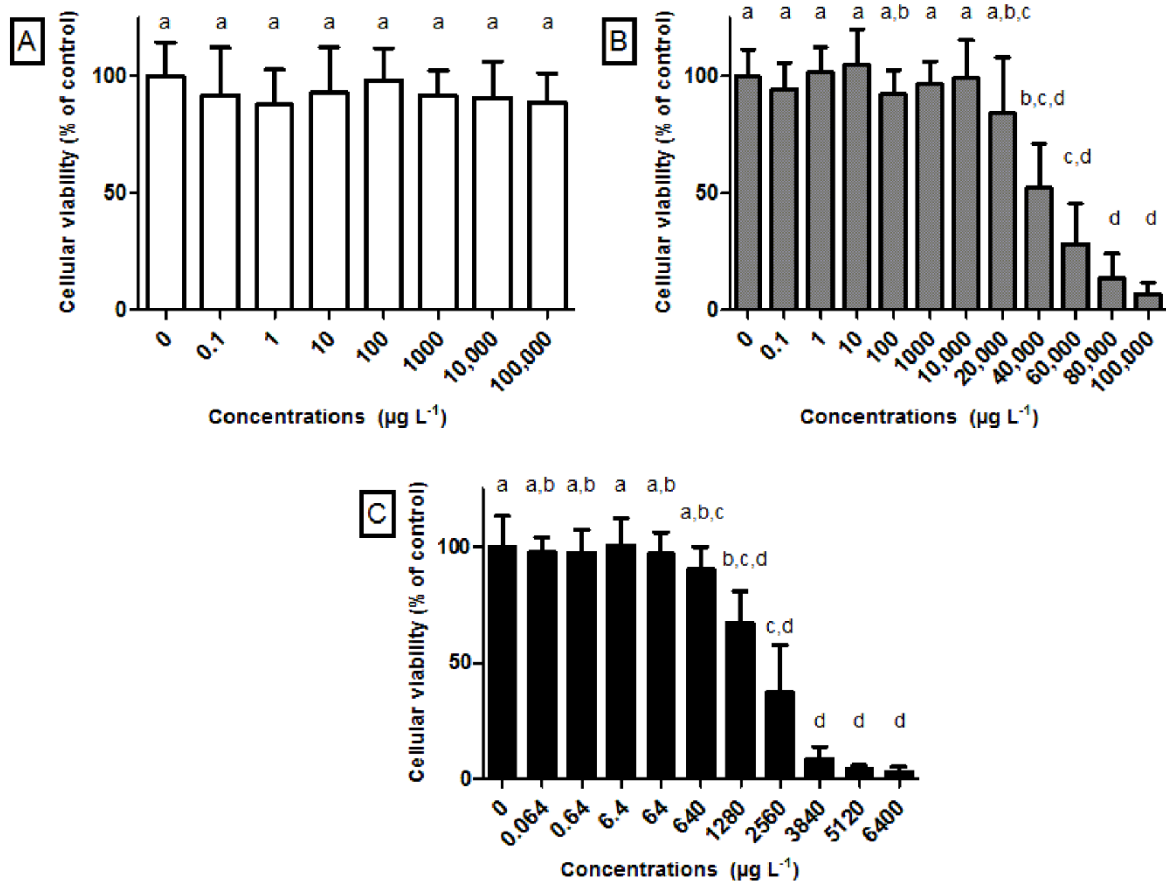


Fig. 1. Cell viability \pm standard deviation (in % of values recorded for control group) after 72h exposure to glyphosate (A) Roundup Express[®] (R_{EX}) (B) and POEAs (C). For each exposure condition, hemocytes came from the hemolymph of at least four abalones and each one's cells were used in triplicate. The concentrations that do not share a letter are significantly different.

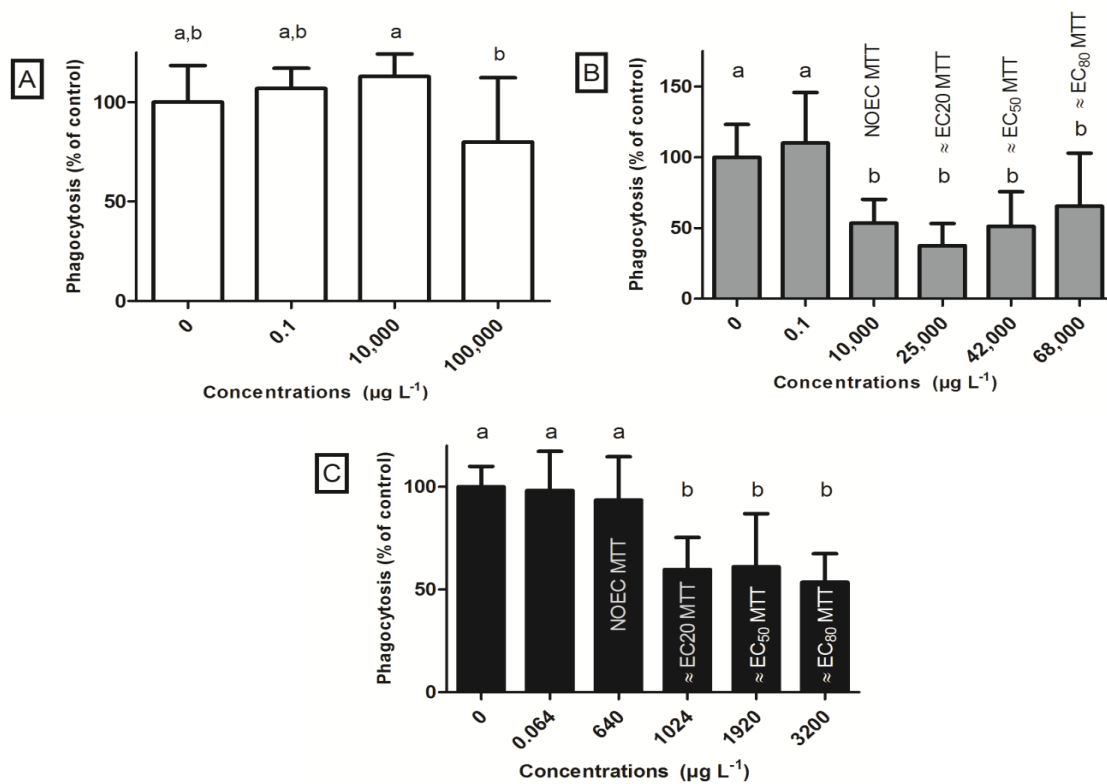


Fig. 2. Phagocytic activity expressed as % of hemocytes that had phagocytosed 3 beads or more \pm standard deviation (in % of values recorded for control group) after 72 h exposure to glyphosate (A) Roundup Express® (R_{EX}) (B) and POEAs (C). For each exposure condition, hemocytes came from the hemolymph of at least three abalones, and a minimum of 20,000 events was considered for each sample. The concentrations that do not share a letter are significantly different.

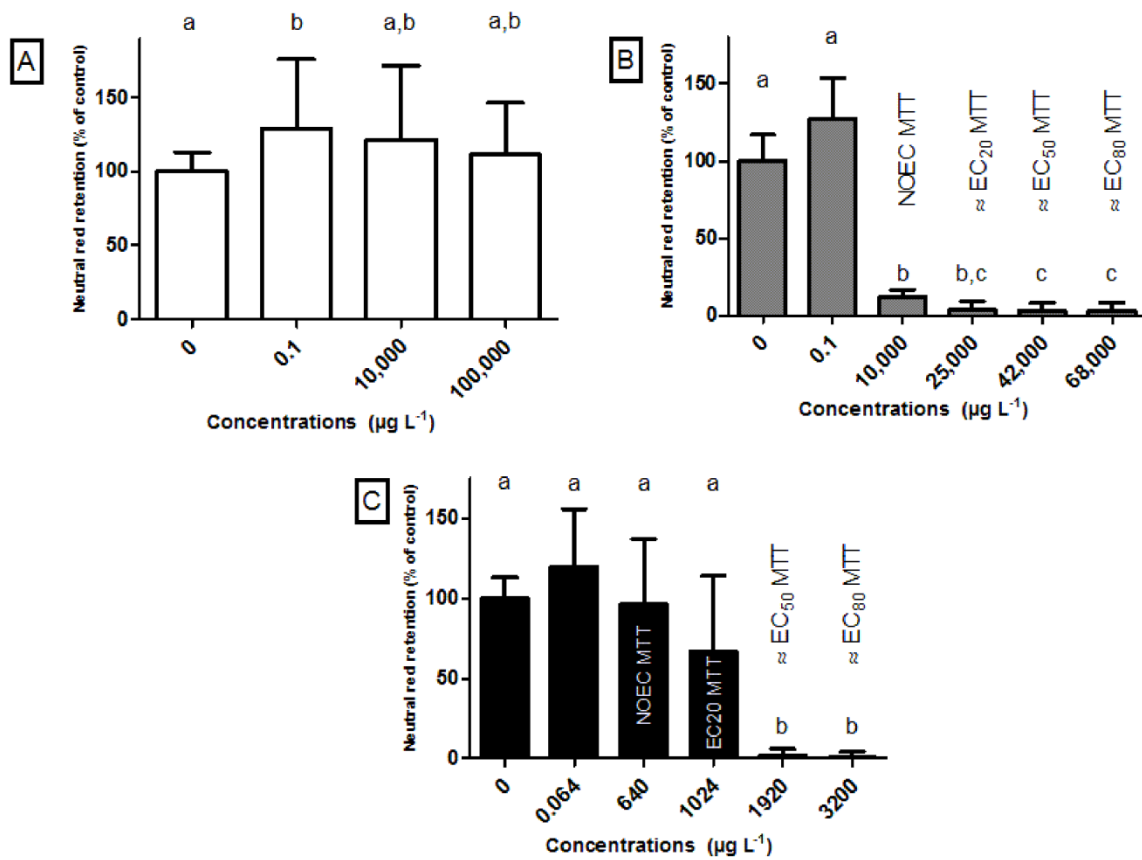


Fig. 3. Neutral red retention \pm standard deviation (in % of values recorded for control group) after 72h exposure to glyphosate (A) Roundup Express® (R_{EX}) (B) and POEAs (C). For each exposure condition, hemocytes came from the hemolymph of at least three abalones and each one's cells were used in triplicate. The concentrations that do not share a letter are significantly different.