

First Description of Azole-Resistant Aspergillus fumigatus Due to TR 46 /Y121F/T289A Mutation in France

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- Abstract: Azole resistance in Aspergillus fumigatus is an emerging public health concern. 21
- Recently, a novel fungicide-driven mutation in the cyp51A gene and its promoter, 22
- 23 TR₄₆/Y121F/T289A, leading to high-level resistance to voriconazole has been identified in
- 24 the Netherlands, Belgium, Germany, Denmark, Tanzania and India in both clinical and
- environmental samples. Here we report the first description of A. fumigatus carrying this 25
- mutation in France, in a cystic fibrosis patient, underlining the need for extensive monitoring 26
- 27 of Aspergillus resistance.

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Azole-resistant Aspergillus fumigatus are increasingly reported in Europe since the late 2000s. 28 This emerging public health concern occurs through two distinct routes of acquisition: in vivo 29 selection of resistance as a consequence of long-term azole treatment or de novo acquisition 30 of a resistant isolate directly from the environment, linked to the widespread use of azole 31 32 fungicides in agriculture. Besides the TR₃₄/L98H mutation in the cyp51A gene first described in the Netherlands, a novel fungicide-driven mutation, TR₄₆/Y121F/T289A has been recently 33 identified. Up to now, the TR₄₆/Y121F/T289A mutation has been reported in both 34 environmental and/or clinical samples in four countries across Europe (1-7), in Asia (8) and 35 more recently in Africa (9), suggesting a large geographical spread. Here we provide the first 36 37 description of A. fumigatus carrying TR₄₆/Y121F/T289A mutation in France, in a cystic fibrosis patient. 38 39 A 23-year-old male cystic fibrosis patient with follow-up at the Pneumology Department at 40 Rouen University Hospital, France, was seen in consultation in March 2014. This patient had 41 42 high levels of total IgE and Aspergillus-specific IgE with positive Aspergillus-specific IgG antibodies, being suggestive of diagnosis of allergic bronchopulmonary aspergillosis. He had 43 a history of A. fumigatus airway colonization and mold-active azoles exposure since 2002 44 (itraconazole and voriconazole). At the time of the consultation, he was treated with 45 46 voriconazole. Mycological cultures of the sputum collected during the consultation grew A. fumigatus (strain 1). Species identification was obtained by both macroscopic and 47 microscopic characteristics on Sabouraud's agar medium together with sequencing of the 48 beta-tubulin gene (10). According to a local research protocol aiming at the surveillance of 49 azole resistance, this isolate was tested for antifungal susceptibility by the Etest® method 50

(bioMérieux, Marcy-l'Etoile, France). Unexpectedly, this strain exhibited a high-level

resistance to voriconazole (MIC>32µg/mL) in contrast with itraconazole (MIC=8µg/mL) and

posaconazole (MIC=1µg/mL). Antifungal susceptibility was confirmed by the EUCAST broth 53 microdilution reference method (Table 1) (11, 12). Nucleotide sequencing of the cyp51A gene 54 and its promoter, using previously described primers (13, 14) and home-designed primers 55 (CYP51AF-F1: 5'-ATTTCCCTCATCACTGCAA, CYP51AF-R1: 5'-56 5'-AGAAGCGAGATGCTGCTCAT, 57 CATCATGTGCGCAATCTCTT, CYP51AF-F2: CYP51AF-R2: 5'-CCTTTGAAGTCCTCGATGGT), showed the TR₄₆/Y121F/T289A 58 mutation. Antifungal therapy was therefore switched to posaconazole in April 2014 then to 59 caspofungin (50 mg per day) in July 2014 because of pulmonary exacerbation. 60 Given these findings, we retrospectively analyzed all A. fumigatus strains previously isolated 61 62 from this patient since 2007 (n=13), for itraconazole and voriconazole susceptibility, cyp51A sequencing and microsatellite genotyping. As shown in Table 1, our patient had already been 63 colonized by a TR₄₆/Y121F/T289A isolate one year before, in February 2013 (strain 3). All 64 remaining isolates collected before February 2013 were azole-susceptible being either wild-65 type for the cyp51A gene or carrying mutations previously found in both azole-resistant and 66 67 azole-susceptible isolates (15). As a single colony was subjected to in vitro susceptibility testing other azole-resistant isolates could have been missed. Microsatellite genotyping was 68 performed using a panel of nine short tandem repeats as described previously (16). As 69 illustrated in Table 1, both TR₄₆/Y121F/T289A isolates from our patient had the same 70 71 genotype as a strain previously isolated in Germany (7) (Table 1). To gain further insights into the route of acquisition of this azole-resistant isolate in our patient, we conducted an 72 environmental study by soil samplings next to the patient's home as described previously 73 (17), as well as surface samplings (contact agar plates) in his office. Neither A. fumigatus 74 carrying TR₄₆/Y121F/T289A nor TR₃₄/L98H was identified. 75

Aspergillus fumigatus isolates carrying the TR₄₆/Y121F/T289A mutation have been first 77 described in December 2009 in the Netherlands (2). Since then, such isolates have been 78 evidenced in three others European countries namely Belgium (1, 5), Germany (3, 7) and 79 Denmark (4) and recently in India (8) and Tanzania (9) (Table 2, Figure 1). Taken together, 80 81 these findings suggest, as discussed previously for TR₃₄/L98H isolates, a large geographical 82 spread of this resistance mechanism. Several lines of evidence indicate that, as for TR₃₄/L98H, TR₄₆/Y121F/T289A has emerged through a fungicide-driven route (18), such 83 isolates being found in both azole-naïve (1, 2, 6, 7) and azole-exposed patients (2, 3, 5, 7) as 84 well as from the environment (2, 8, 9). Here we report the first description of A. fumigatus 85 86 carrying the TR₄₆/Y121F/T289A mutation isolated from a French patient. Interestingly, our patient organized trips to the Netherlands as a tour operator. For these 87 working purposes, he traveled to Amsterdam in November 2012, three months before the first 88 isolation of the TR₄₆/Y121F/T289A strain from his sputum (February 2013). Moreover he 89 regularly received advertising postal packages from Dutch flower producers which were 90 91 opened in his office. Three hypotheses can explain the route of acquisition of this TR₄₆/Y121F/T289A strain in our patient: (i) inhalation of spores carrying TR₄₆/Y121F/T289A 92 during his trip to the Netherlands (2); (ii) colonization after inhaling spores carrying 93 TR₄₆/Y121F/T289A from his environment in France. Our environmental study conducted 94 95 next to the patient's home, less than 100 km from Belgium (where TR₄₆/Y121F/T289A strains have been recently identified (1)), failed to detect TR₄₆/Y121F/T289A environmental isolates. 96 Nevertheless, environmental isolates carrying this mutation have been recently identified by 97 our team in the same region in France supporting this hypothesis (unpublished data). (iii) The 98 hypothesis is colonization after inhaling A. fumigatus spores carrying 99 100 TR₄₆/Y121F/T289A, escaping while opening the packages received from the Netherlands. 101 Though the French strains are genetically indistinguishable from the German and genetically

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different from Dutch isolates, the route of acquisition in our patient is unclear, as spores probably follow an airborne migration as hypothesized previously for TR₃₄/L98H (19, 20). The present report now provides evidence that A. fumigatus voriconazole-resistant isolates carrying the TR₄₆/Y121F/T289A mutation can be now isolated from clinical samples in France. As observed with TR₃₄/L98H, a geographical spread of this resistance mechanism is ongoing across Europe and possibly worldwide. These findings together with the high-level voriconazole resistance of the TR₄₆/Y121F/T289A strains both in vitro and in vivo (1, 2, 4-6) underline the need for intensive investigations to determine its prevalence in both clinical and environmental samples. In line with this, as recommended by the ECDC Technical report (18), antifungal susceptibility testing of triazoles should be performed on all clinical A. *fumigatus* isolates before starting antifungal therapy. Acknowledgments: We are grateful to all the technicians of the Parasitology and Mycology Laboratories at Nantes University Hospital and Rouen University hospital for technical assistance. Conflict of interest: PLP received grants from Astellas and Pfizer and speaker's fees from Merck and Gilead. FM received speaker's fees from Gilead and MSD and travel grants from Gilead, MSD, Pfizer and Astellas. JFM received grants from Astellas, Basilea and Merck, has been a consultant to Astellas, Basilea and Merck and received speaker's fees from Merck and

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Table 1- Overview of the characteristics of all Aspergillus fumigatus strains isolated from 193 194 sputum samples of the patient 195 Table 2- Literature review of all studies reporting TR₄₆/Y121F/T289A A. fumigatus isolates 196 Figure 1- Geographical spread of the TR₄₆/Y121F/T289A resistance mechanism (for each 197 strain, the exact location and origin (clinical or environmental) is indicated) Figure 2- STRAf dendrogram highlighting the genetic relatedness between Aspergillus 198

fumigatus isolated from our patient and previously reported TR₄₆/Y121F/T289A isolates

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Table 1- Overview of the characteristics of all Aspergillus fumigatus strains isolated from sputum samples of the patient

	Reference in the dendrogram	Month/Year of isolation	MICs mg/L (EUCAST)					
Strain number					Cyp51A mutations			
			ITC	VRC	-			
1	14-105-2468	Mar 2014	8	>8	TR ₄₆ /Y121F/T289A			
2	14-148-2457	Nov 2013	0.5	0.5	Wild-type			
3	14-148-2460	Feb 2013	>8	>8	TR ₄₆ /Y121F/T289A			
4	141428-459	Jan 2013	0.5	1	Wild-type			
5	14-148-2458	Jan 2013	0.25	0.25	Wild-type			
6	14-148-2456	Dec 2010	Not performed		F46Y, G89G, M172V, N248T, D255E, L358L, E427K, C454			
7	14-148-2455	Sept 2010	Not performed		F46Y, G89G, M172V, N248T, D255E, L358L, E427K, C454			
8	14-148-2454	Jul 2010	Not performed		F46Y, G89G, M172V, N248T, D255E, L358L, E427K, C454			
9	Not performed	Jul 2009	Not pe	rformed	F46Y, G89G, M172V, N248T, D255E, L358L, E427K, C454			
10	14-148-2450	Jul 2009	0.25	1	Wild-type			
11	14-148-2448	Mar 2009	0.25	1	Wild-type			
12	14-148-2447	Dec 2007	0.25	1	Wild-type			
13	14-148-2445	May 2007	0.5	2	Wild-type			
14	Not performed	Feb 2007	0.5	1	F46Y, G89G, M172V, N248T, D255E, L358L, E427K, C454			

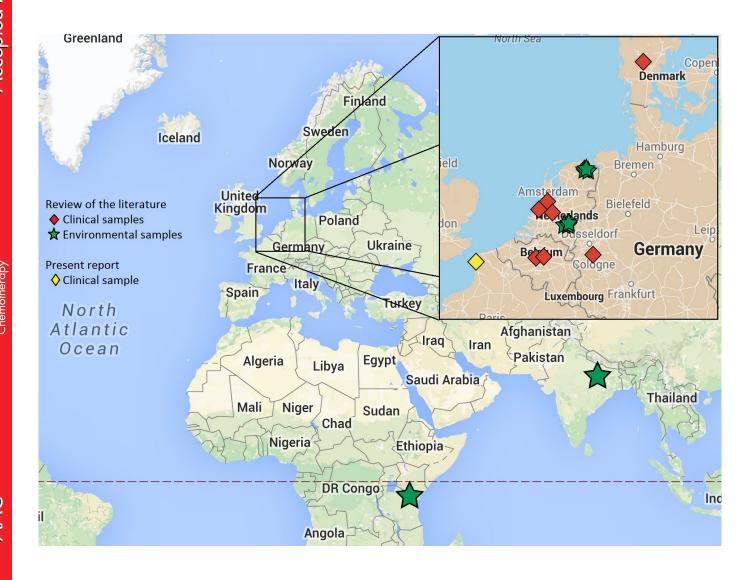
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Table 2- Literature review of all studies reporting TR₄₆/Y121F/T289A A. fumigatus isolates

Reference	Date	Type of sample	Underlying condition	Infection	Antifungal susceptibility CLSI (EUCAST*)			Outcome	Country
					VRC	ITC	POS		
(1)	July 2012	BAL fluid	HSCT	Probable IA	>16	4	1	Death	Belgium
(2)	Dec 2009	Sputum	HSCT	Probable IA	>16	4	0.25	Persistent infection	
	Jan 2010	Ear	Chronic otitis externals/sinusitis	IA	>16	>16	2	Persistent infection	
	Jan 2010	Abdominal abscess	SOT	Proven IA	>16	2	0.5	Death	
	Feb 2010	Sputum	Cystic fibrosis	No IA	>16	4	0.5	Alive	
	Feb 2010	Sputum	Lung carcinoma	No IA	>16	>16	2	Alive	
	March 2010	Sputum	HSCT	Probable IA	>16	1	0.25	Death	
	March 2010	Sputum	Cystic fibrosis, SOT	Proven IA	>16	>16	0.5	Alive	
	May 2010	Biopsy	Chronic otitis, surgery	Proven IA	>16	4	1	Alive	The Netherlands
	May 2010	Sputum	Lung fibrosis	None	>16	>16	1	Alive	
	June 2010	Sputum	Traumatism	None	>16	1	0.25	Death	
	July 2010	Brain biopsy	Beta thalassemia, diabetes mellitus	Proven IA	>16	4	1	Death	
	Sept 2010	Sputum	Cystic fibrosis	ABPA	>16	2	0.5	Alive	
	Oct 2010	Sputum	COPD, SOT	None	>16	>16	2	Alive	
	Nov 2010	Sputum	COPD	No IA	>16	>16	2	Alive	
	Jan 2011	Sputum	HSCT	Probable IA	>16	>16	1	Death	
	Dec 2009 to Jan 2011	Air sampling			ND	ND	ND		
(8)	2012-2013	Soil sampling			>16	1-2	0.25-0.5		India
(3)	Sept 2012	Sputum	Cystic fibrosis	Colonization	>8*	>8*	2*	Alive	Germany
(4)	Jan 2014	Sputum	Bruton's agammaglobulinemia, SOT	Probable IA	>4*	0.25- 0.5*	0.125- 0.25*	Death	Denmark
(9)		Soil sampling			16->16	1-2	0.25-0.5		Tanzania
(5)	Nov 2013	BAL fluid	HSCT	Probable IA	>8	>16	1	Death	Belgium
(7)	Sept 2012	BAL fluid	HSCT	Probable IA	16*	>16*	0.5*	Death	Germany
	July 2012	BAL fluid	HSCT	Proven IA	1*	>16*	0.5*	Death	
Present report	Feb 2013 March 2014	Sputum	Cystic fibrosis	Colonization	>8* >8*	>8* 8*	ND	Alive	France

POS: posaconazole BAL: bronchoalveolar lavage; HSCT: hematopoietic stem cell transplantation; IA: invasive aspergillosis; SOT: solid organ transplantation; ABPA: allergic bronchopulmonary aspergillosis; COPD: chronic obstructive pulmonary disease; ND: Not determined





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