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Relevance of detection of mechanisms of resistance to ALK inhibitors in ALK-rearranged NSCLC in routine practice

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Background

ALK Tyrosine Kinase Inhibitors (ALK TKIs) have demonstrated efficacy in the treatment of ALK-rearranged Non-Small Cell Lung Cancer (NSCLC), but the disease eventually progresses in all patients. In many cases, resistance to ALK TKIs arises through ALK mutations. Although clinical and biological data suggest variations in TKI efficacy according to the mechanism of resistance, ALK mutations are still rarely investigated in routine practice.

Material and methods

We performed a retrospective multicentric study aiming to determine the frequency and clinical relevance of ALK alterations detected by targeted Next-Generation Sequencing (t-NGS) in patients with advanced ALK-rearranged NSCLC following progression on an ALK TKI. Clinical, pathological, and molecular characteristics and patient outcomes were collected.

Results

We identified 23 patients with advanced ALK-rearranged NSCLC who, between January 2012 and May 2017, had undergone at least one repeat biopsy at progression on an ALK TKI. A resistance mechanism was identified in 9 of the 23 patients (39%). The anomalies involved included 9 ALK mutations in 8 patients and one *ALK* amplification. The ALK mutation rate was 15% after failure of a first ALK TKI and 33% after failure of two ALK TKIs. Five out of seven patients who received a different ALK TKI following detection of an ALK mutation achieved an objective response. All the patients who received a TKI presumed to act on the detected ALK mutant achieved disease control.

Conclusions

Targeted NGS is suitable for detecting ALK resistance mutations in ALK-rearranged NSCLC patients in routine practice. It may help select the best treatment at progression on an ALK TKI.

1 **1. Introduction**

2 ALK rearrangements are found in approximately 4% of non-squamous Non-Small Cell Lung
3 Cancer (NSCLC) [1]. The fusion protein resulting from ALK rearrangements has a
4 constitutively activated ALK kinase leading to activation of downstream signaling pathways
5 involved in cell proliferation and cell survival[2,3]. The discovery of ALK rearrangements has
6 led to the development of tyrosine kinase inhibitors (TKIs) targeting the ALK kinase.
7 Crizotinib and ceritinib have demonstrated superiority over chemotherapy against untreated
8 advanced ALK-rearranged NSCLC patients, and alectinib has demonstrated superiority over
9 crizotinib in the same context [4–6]. Several TKIs have proved active following crizotinib
10 failure, including ceritinib, alectinib, brigatinib, and lorlatinib [7–10].

11 Mechanisms of resistance to ALK TKIs in ALK-rearranged NSCLC patients are partly known
12 and are of two main types: i) on-target mechanisms, including mutation of the ALK kinase
13 domain and amplification of the ALK-rearranged gene and ii) off-target mechanisms,
14 including activation of an alternative signaling pathway such as EGFR or KIT and
15 pathological transformation [11].

16 ALK mutations are the main cause of resistance to ALK TKIs. With high-throughput methods
17 of detection [11], they are found in 20-50% of tumor samples obtained at progression on an
18 ALK TKI. Numerous ALK kinase mutations can confer resistance to ALK TKIs in ALK-
19 rearranged NSCLC [12–19], and the frequency and type of ALK mutations detected at
20 progression depend on the type of ALK TKI received [11]. Thus, each ALK TKI has its own
21 spectrum of activity against ALK kinase mutants [20], and a given ALK kinase mutant is not
22 resistant to all ALK TKIs. For example, the L1196M mutation is reported to confer resistance
23 only to crizotinib, the C1156Y mutation is associated with resistance to crizotinib and ceritinib
24 but remains sensitive to alectinib and brigatinib, and the G1202R mutation leads to
25 resistance to most ALK TKIs but not to lorlatinib [21–24].

26 Thus, identifying the mechanism of resistance to an ALK TKI may help in choosing the most
27 appropriate treatment after progression on that ALK TKI, particularly since several ALK

1 inhibitors are now available for use in this context. Here we show that targeted NGS
2 performed in routine practice is suitable and relevant for detecting ALK mutations in tumor
3 tissue samples or circulating tumor DNA from ALK-positive NSCLC patients whose disease
4 has progressed after treatment with an ALK TKI.

5 6 **2. Material and methods**

7 2.1. Study design and patients

8 In this observational, multicentric, retrospective study, we identified patients with advanced
9 ALK-positive NSCLC from whom tumor samples had been obtained at progression on an
10 ALK TKI as part of the routine procedure. Both tissue biopsies and circulating tumor DNA
11 samples were considered. Clinical and molecular data were collected retrospectively at each
12 center. The investigators used RECIST1.1 to evaluate responses to ALK TKIs. The objective
13 response rate was defined as the rate of complete or partial response. The disease control
14 rate was defined as the rate of complete or partial response or stable disease. Overall
15 survival was calculated from the start of the first ALK TKI treatment received.

16 All the data reported in this study were extracted from a database approved by the French
17 National Data Protection Authority (CNIL). This non-interventional study was conducted in
18 accordance with the Declaration of Helsinki and Good Clinical Practice guidelines. It was
19 approved by a national ethics committee and by the French Advisory Committee on
20 Information Processing Related to Research in the Field of Health. All included patients
21 received information from their referring physician.

22 23 2.2. Detection of ALK mutations

24 Targeted NGS (t-NGS) analysis was conducted with the NGS commercial panel “Ion
25 Ampliseq Colon and Lung Research Panel V2” at two molecular centers certified by the
26 French National Cancer Institute. In the case of tissue samples, genomic DNA was extracted
27 from formalin-fixed paraffin embedded (FFPE) samples received in the two laboratories
28 between February 2012 and February 2017. DNA was extracted with the QIAamp FFPE

1 tissue kit (Qiagen) according to the manufacturer's instructions. In the case of blood
2 samples, cell-free DNA was extracted from plasma with the QIAamp Circulating Nucleic Acid
3 Kit (Qiagen) according to the manufacturer's instructions. Extracted DNA was quantified with
4 the Quant-it PicoGreen dsDNA Assay Kit (ThermoFisher Scientific) on a Xenius XC
5 spectrofluorometer (Safas Monaco). AmpliSeq libraries were prepared with the Ion AmpliSeq
6 Library Kit 2.0 Ion Xpress barcode adapters kit, primers (Ion AmpliSeq custom panel or
7 Colon and Lung V2) (ThermoFisher Scientific), and 10 ng or less of each DNA sample
8 according to the manufacturer's instructions (Table S1). Emulsion PCR, enrichment, and chip
9 loading were performed on an Ion Chef instrument with the Ion PI™ Hi-Q™ Chef Kit or the
10 Ion 540™ Kit-Chef. Samples were sequenced with Ion P1 chips on Ion Proton System or Ion
11 540 chips on the Ion S5XL system (ThermoFisher Scientific). Data were analyzed with the
12 Torrent suite software v.5.2.2 (ThermoFisher Scientific). For the ALK, EGFR, and KRAS
13 genes, manual inspection of sequences was done on an integrative genomic viewer (IGV).

14

15 2.3. Statistical analysis

16 Quantitative variables are expressed as means (standard deviation, SD) in the case of a
17 normal distribution or as medians (interquartile ranges) otherwise. Categorical variables are
18 expressed as numbers (percentages). The normality of distributions was assessed with
19 histograms and the Shapiro-Wilk test. Survival rates were calculated from the date of
20 introduction of the first ALK TKI until the date of death or last news and derived from Kaplan-
21 Meier survival curves. Medians of survival were calculated with confidence intervals.
22 Statistical testing was conducted at a two-tailed α -level of 0.05. Data were analyzed with the
23 SAS software version 9.4 (SAS Institute, Cary, NC).

24

25 **3. Results**

26 The study included 23 patients with advanced ALK-positive NSCLC having undergone at
27 least one biopsy at progression on an ALK TKI between January 2012 and May 2017 (**Table**
28 **1**). The patients were mostly never smokers (n=19, 83%) and the median age was 57.0

1 years (range 17-70). Sixteen patients (70%) had received platinum-based chemotherapy
2 prior to ALK TKI treatment. Twenty-two patients (96%) had received crizotinib as the first ALK
3 TKI. A second-generation ALK TKI (alectinib, ceritinib, or brigatinib) had been given to one
4 patient (4%) as first-line treatment and to seventeen patients (74%) following crizotinib
5 failure. Three patients having first received crizotinib and then a second-generation ALK TKI
6 had subsequently received a third-generation ALK TKI (lorlatinib).

7

8 The number of biopsies performed on the 23 patients totaled 42, including 26 tissue biopsies
9 and 16 liquid biopsies (cell-free DNA samples). This corresponds to a mean of 1.83 biopsies
10 per patient (range 1-7). Main biopsy sites were lungs in 8 patients, pleural effusion in 5
11 patients and liver in 4 patients (Table S2). The samples studied were from 13 of the 23
12 patients (57%) having progressed on a first TKI, 12 of the 15 patients (80%) having
13 progressed on a second TKI, and 3 of the 7 patients (43%) having progressed on a TKI given
14 as a third TKI or further. The studied samples also included five biopsies performed at
15 progression on therapies other than ALK TKI.

16 The results of targeted NGS applied to these samples are as follows. ALK mutations were
17 detected in eight patients (35%). They included four G1202R mutations and one C1156Y,
18 one V1180L, one I1171N, and one L1196M mutation. In one of the patients where a G1202R
19 mutation was detected initially, a G1202R/G1269A compound mutation was found in a later
20 biopsy. Seven ALK mutations were detected in tissue biopsies and three in circulating free
21 DNA. For 3 patients, liquid and tissue biopsies were performed at the same time. The results
22 were consistent in one patient.

23 An ALK amplification with no concomitant ALK mutation was detected by FISH in one biopsy
24 (4.34 %) (Figure 1). In addition to an ALK mutation, seven patients had a TP53 mutation
25 (A138P, R175H, C238R, C277G, R248Q, R337C, R337P) and one had a MET mutation in
26 exon 18 (c.3686_3686 + 3del). No mutation was identified in any other oncogene tested with
27 this NGS panel..

1 Among the nine ALK mutations detected, one was found in a sample taken at progression on
2 crizotinib given as the first TKI. The eight remaining mutations were identified in samples
3 from patients exposed to a second- or third-generation TKI. The rate of detection of ALK
4 mutations at progression was 15% after a first TKI (2 out of 13 patients tested), 33% after a
5 second TKI (4 out of 12 patients tested) and 66% after a third TKI or more (2 out of 3 patients
6 tested) (**Figure 2**).

7 Among patients on the first ALK TKI, the objective response rate was 74% and the disease
8 control rate was 91%. By the end of data collection, twelve patients (52%) had died. Median
9 overall survival from the start of the first ALK TKI treatment was 37.28 months (95% CI;
10 19.58-NR). We did not observed any difference in OS between patients with or without TP53
11 mutations. Among the eight patients in whom an ALK mutation was identified, seven (87%)
12 received a new ALK TKI after the mutation was found (**Figure 3**). In these seven patients the
13 ORR with the new TKI was 71% (5/7). Among the patients in whom no ALK mutation was
14 found, the ORR with the third ALK TKI was 33% (1/3).

15 Among the seven patients with an ALK mutation who received a subsequent ALK TKI, six
16 received a TKI presumed on the basis of *in vitro* data to act against the identified mutant [11].
17 Among these six patients, five achieved an objective response and one showed stable
18 disease. To the remaining patient alectinib was given despite detection of a G1202R
19 mutation, as at the time alectinib was the only available next-generation TKI and data were
20 still sparse regarding the activity of different ALK TKIs according to the ALK mutation
21 present. This patient displayed progressive disease at first assessment (**Figure 3**).

22

23 **4. Discussion**

24 The present study has focused on targeted NGS, a technique now widely used in daily
25 molecular testing. We show that its use in routine practice allowed identifying an ALK TKI
26 resistance mechanism in 9 out of 23 patients (39%) with advanced ALK-positive NSCLC.
27 Gainor *et al.* report a similar rate of ALK mutations for other NGS panels [11]. Thus, targeted
28 NGS appears suitable for detecting ALK mutations in routine practice.

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It is anticipated that ALK mutations will be detected more frequently in the near future, Gainor *et al.* having shown that they arise more frequently following second- or third-generation TKI treatment (54-71%) than after crizotinib (20% of patients) [11]. Meanwhile two second-generation TKIs, ceritinib and alectinib, have demonstrated efficacy as first-line treatment, [4,25] and ongoing clinical trials are currently evaluating next-generation TKIs (NCT03052608, NCT02737501, NCT02767804) in this context. Although no report so far has provided rates of ALK mutations after progression on second-generation TKIs given as first-line therapy, it is reasonable to speculate that they will exceed the 20% observed after crizotinib failure. Data further suggest that the ALK mutation rate also increases when several ALK TKIs are given sequentially. Solomon *et al.* report ALK mutation rates of 16.6% (14/84) following first-line ALK TKI treatment and 29.2% (31/106) after at least two ALK TKIs [26]. We have likewise observed a 15% ALK mutation rate after failure of a first ALK TKI and a 33% ALK mutation rate after failure of two ALK TKIs. Whether this is due to the number of ALK TKIs administered or to the use of next-generation TKIs remains to be determined.

All these findings and the fact that ALK-positive patients have been shown to benefit from receiving multiple TKIs [8,27,28] suggest that the future will see an increase in the proportion of patients exposed to next-generation ALK TKIs, along with an increase in the rate of ALK mutations.

Detection of ALK mutations in patients having progressed after ALK TKI treatment is becoming crucial to determining the optimal therapeutic strategy. *In vitro*, different ALK mutations have been found to predict different sensitivities to a single ALK TKI, and ALK TKIs do not all share the same spectrum of activity against ALK mutants [11,29]. Similar findings have been reported in patients, although extrapolating from *in vitro* data to patients requires caution. In two patients, for example, the I1171N mutation has been found to drive resistance to alectinib and sensitivity to ceritinib [30,31]. In another patient, the L1152R mutation has on the contrary been reported to confer resistance to ceritinib and to predict a

1 response to alectinib [32]. Consistently, we have found that all six patients who were treated
2 with an ALK TKI presumed to act against the detected mutation achieved disease control,
3 whereas the only patient who was treated with an ALK TKI not presumed to act effectively
4 against the detected mutation displayed progressive disease at first assessment.
5 Interestingly, two patients harboring a G1202R mutation achieved a partial response on
6 brigatinib, reported to exert intermediate activity on G1202R mutants *in vitro*.

7 To date, no clinical trial has assessed the clinical relevance of identifying ALK mutations at
8 progression on an ALK TKI. Yet besides the above-mentioned case reports, data from
9 prospective trials suggest that patients harboring an ALK mutation are more likely to respond
10 to subsequent ALK TKIs. In patients receiving the third-generation TKI lorlatinib after failure
11 of two or three prior ALK TKIs, Shaw *et al.* report objective response rates of 26% in patients
12 with no ALK mutation and 61% in patients with at least one ALK mutation [33]. Interestingly,
13 we have found a similar trend in our set of patients treated in routine practice: we observed a
14 71% ORR with subsequent TKI treatment in patients known to have an ALK mutation and a
15 33% ORR in patients with no ALK mutation, after treatment with a third TKI. Here, the
16 presence of an ALK mutation may reveal sustained dependence on the ALK pathway and
17 thus predict sensitivity to ALK inhibition [26], and the type of ALK mutation should guide the
18 choice of the ALK TKI to be used. Inversely, the absence of any ALK mutation suggests that
19 resistance may be due to an ALK-pathway-independent mechanism that is not sensitive to
20 ALK inhibition. Various ALK-independent resistance mechanisms have been described,
21 including activation of EGFR, IGF1R, and SRC, mutation of *KRAS* or *PIK3CA*, amplification
22 of *cKIT* and gene fusions [12,13,18,34–37]. In *in vitro* studies, cells having acquired
23 resistance to an ALK TKI via an ALK-independent mechanism showed cross-resistance to
24 other ALK TKIs, but sensitivity to ALK TKIs could be restored by combined treatment with
25 another TKI [34,37,38]. Efficacy of ALK TKI in patients having acquired resistance to a given
26 ALK TKI via an ALK-independent mechanism is still poorly known [37]. Patients having
27 progressed on crizotinib with no evidence of an *ALK* mutation have been shown to benefit
28 from second-generation ALK TKIs [39], but whether their resistance was due to an ALK-

1 independent mechanism, to an ALK-dependent mechanism not involving an ALK mutation,
2 or to an ALK mutation that escaped detection because of sensitivity issues remains
3 unknown.

4 Sequential treatment with several ALK TKIs may select resistant clones harboring multiple
5 (i.e. compound) ALK mutations. In a recent study, Yoda *et al.* found a 35% compound
6 mutation rate in patients treated with lorlatinib [40]; in our set, two patients had progressed
7 on lorlatinib and a compound mutation was detected at the time of progression in a patient
8 known to harbor a G1202R mutation. *In vitro*, compound mutations have been found to
9 confer resistance to most ALK TKIs, including lorlatinib [40]. Yet a patient having acquired
10 resistance to crizotinib, ceritinib, and lorlatinib successively and who was found to harbor a
11 C1156Y/L1196M compound mutation achieved an objective response on crizotinib. It is
12 necessary to further determine the sensitivity of compound mutants to ALK TKIs [16].

13 Finally, other molecular alterations may impact patients outcome. Recent findings have
14 shown that TP53 mutations or ALK variants may be associated with shorter PFS and OS in
15 ALK-positive NSCLC patients [41,42]. In our study, no difference in OS was found between
16 patients with or without TP53 mutations, but the number of patients in each subgroup was
17 too low to draw any conclusion from this result. ALK variants were not assessed in our study.

18

19 **Conclusions:**

20 In summary, our findings demonstrate that mechanisms of resistance to ALK TKIs can be
21 detected in routine practice in a third of ALK-positive NSCLC patients. Identifying ALK
22 resistance mutations may help selecting the best subsequent treatment

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1 **References**

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- 3 [1] B. Solomon, M. Varella-Garcia, D.R. Camidge, ALK Gene Rearrangements: A New Therapeutic Target
4 in a Molecularly Defined Subset of Non-small Cell Lung Cancer, *J. Thorac. Oncol.* 4 (2009) 1450–1454.
5 doi:10.1097/JTO.0b013e3181c4dedb.
- 6 [2] R. Chiarle, C. Voena, C. Ambrogio, R. Piva, G. Inghirami, The anaplastic lymphoma kinase in the
7 pathogenesis of cancer, *Nat. Rev. Cancer.* 8 (2008) 11–23. doi:10.1038/nrc2291.
- 8 [3] J.J. Lin, G.J. Riely, A.T. Shaw, Targeting ALK: Precision Medicine Takes on Drug Resistance., *Cancer*
9 *Discov.* 7 (2017) 137–155. doi:10.1158/2159-8290.CD-16-1123.
- 10 [4] J.-C. Soria, D.S.W. Tan, R. Chiari, Y.-L. Wu, L. Paz-Ares, J. Wolf, S.L. Geater, S. Orlov, D. Cortinovis,
11 C.-J. Yu, M. Hochmair, A.B. Cortot, C.-M. Tsai, D. Moro-Sibilot, R.G. Campelo, T. McCulloch, P. Sen,
12 M. Dugan, S. Pantano, F. Branle, C. Massacesi, G. de Castro, First-line ceritinib versus platinum-based
13 chemotherapy in advanced ALK -rearranged non-small-cell lung cancer (ASCEND-4): a randomised,
14 open-label, phase 3 study, *Lancet.* 389 (2017) 917–929. doi:10.1016/S0140-6736(17)30123-X.
- 15 [5] A.T. Shaw, D.-W. Kim, K. Nakagawa, T. Seto, L. Crinó, M.-J. Ahn, T. De Pas, B. Besse, B.J. Solomon,
16 F. Blackhall, Y.-L. Wu, M. Thomas, K.J. O’Byrne, D. Moro-Sibilot, D.R. Camidge, T. Mok, V. Hirsh,
17 G.J. Riely, S. Iyer, V. Tassell, A. Polli, K.D. Wilner, P.A. Jänne, Crizotinib versus Chemotherapy in
18 Advanced *ALK* -Positive Lung Cancer, *N. Engl. J. Med.* 368 (2013) 2385–2394.
19 doi:10.1056/NEJMoa1214886.
- 20 [6] T. Hida, H. Nokihara, M. Kondo, Y.H. Kim, K. Azuma, T. Seto, Y. Takiguchi, M. Nishio, H. Yoshioka,
21 F. Imamura, K. Hotta, S. Watanabe, K. Goto, M. Satouchi, T. Kozuki, T. Shukuya, K. Nakagawa, T.
22 Mitsudomi, N. Yamamoto, T. Asakawa, R. Asabe, T. Tanaka, T. Tamura, Alectinib versus crizotinib in
23 patients with ALK-positive non-small-cell lung cancer (J-ALEX): an open-label, randomised phase 3
24 trial., *Lancet (London, England).* 390 (2017) 29–39. doi:10.1016/S0140-6736(17)30565-2.
- 25 [7] L. Friboulet, N. Li, R. Katayama, C.C. Lee, J.F. Gainor, A.S. Crystal, P.-Y. Michellys, M.M. Awad, N.
26 Yanagitani, S. Kim, A.C. Pferdekamper, J. Li, S. Kasibhatla, F. Sun, X. Sun, S. Hua, P. McNamara, S.
27 Mahmood, E.L. Lockerman, N. Fujita, M. Nishio, J.L. Harris, A.T. Shaw, J.A. Engelman, The ALK
28 Inhibitor Ceritinib Overcomes Crizotinib Resistance in Non–Small Cell Lung Cancer, *Cancer Discov.* 4

- 1 (2014). <http://cancerdiscovery.aacrjournals.org/content/4/6/662.long> (accessed April 6, 2017).
- 2 [8] A.T. Shaw, L. Gandhi, S. Gadgeel, G.J. Riely, J. Cetnar, H. West, D.R. Camidge, M.A. Socinski, A.
3 Chiappori, T. Mekhail, B.H. Chao, H. Borghaei, K.A. Gold, A. Zeaiter, W. Bordogna, B. Balas, O. Puig,
4 V. Henschel, S.-H.I. Ou, study investigators, Alectinib in ALK-positive, crizotinib-resistant, non-small-
5 cell lung cancer: a single-group, multicentre, phase 2 trial, *Lancet Oncol.* 17 (2016) 234–242.
6 doi:10.1016/S1470-2045(15)00488-X.
- 7 [9] L. Mezquita, D. Planchard, The role of brigatinib in crizotinib-resistant non-small cell lung cancer.,
8 *Cancer Manag. Res.* 10 (2018) 123–130. doi:10.2147/CMAR.S129963.
- 9 [10] A.T. Shaw, E. Felip, T.M. Bauer, B. Besse, A. Navarro, S. Postel-Vinay, J.F. Gainor, M. Johnson, J.
10 Dietrich, L.P. James, J.S. Clancy, J. Chen, J.-F. Martini, A. Abbattista, B.J. Solomon, Lorlatinib in non-
11 small-cell lung cancer with ALK or ROS1 rearrangement: an international, multicentre, open-label,
12 single-arm first-in-man phase 1 trial, *Lancet Oncol.* 18 (2017) 1590–1599. doi:10.1016/S1470-
13 2045(17)30680-0.
- 14 [11] J.F. Gainor, L. Dardaei, S. Yoda, L. Friboulet, I. Leshchiner, R. Katayama, I. Dagogo-Jack, S. Gadgeel,
15 K. Schultz, M. Singh, E. Chin, M. Parks, D. Lee, R.H. DiCecca, E. Lockerman, T. Huynh, J. Logan, L.L.
16 Ritterhouse, L.P. Le, A. Muniappan, S. Digumarthy, C. Channick, C. Keyes, G. Getz, D. Dias-Santagata,
17 R.S. Heist, J. Lennerz, L. V Sequist, C.H. Benes, A.J. Iafrate, M. Mino-Kenudson, J.A. Engelman, A.T.
18 Shaw, Molecular Mechanisms of Resistance to First- and Second-Generation ALK Inhibitors in ALK-
19 Rearranged Lung Cancer., *Cancer Discov.* 6 (2016) 1118–1133. doi:10.1158/2159-8290.CD-16-0596.
- 20 [12] R. Katayama, A.T. Shaw, T.M. Khan, M. Mino-Kenudson, B.J. Solomon, B. Halmos, N.A. Jessop, J.C.
21 Wain, A.T. Yeo, C. Benes, L. Drew, J.C. Saeh, K. Crosby, L. V. Sequist, A.J. Iafrate, J.A. Engelman,
22 Mechanisms of Acquired Crizotinib Resistance in ALK-Rearranged Lung Cancers, *Sci. Transl. Med.* 4
23 (2012) 120ra17-120ra17. doi:10.1126/scitranslmed.3003316.
- 24 [13] T. Sasaki, J. Koivunen, A. Ogino, M. Yanagita, S. Nikiforow, W. Zheng, C. Lathan, J.P. Marcoux, J. Du,
25 K. Okuda, M. Capelletti, T. Shimamura, D. Ercan, M. Stumpfova, Y. Xiao, S. Weremowicz, M.
26 Butaney, S. Heon, K. Wilner, J.G. Christensen, M.J. Eck, K.-K. Wong, N. Lindeman, N.S. Gray, S.J.
27 Rodig, P.A. Janne, A Novel ALK Secondary Mutation and EGFR Signaling Cause Resistance to ALK
28 Kinase Inhibitors, *Cancer Res.* 71 (2011) 6051–6060. doi:10.1158/0008-5472.CAN-11-1340.

- 1 [14] Y.L. Choi, M. Soda, Y. Yamashita, T. Ueno, J. Takashima, T. Nakajima, Y. Yatabe, K. Takeuchi, T.
2 Hamada, H. Haruta, Y. Ishikawa, H. Kimura, T. Mitsudomi, Y. Tanio, H. Mano, ALK Lung Cancer
3 Study Group, EML4-ALK Mutations in Lung Cancer That Confer Resistance to ALK Inhibitors, N.
4 Engl. J. Med. 363 (2010) 1734–1739. doi:10.1056/NEJMoa1007478.
- 5 [15] T. Sasaki, K. Okuda, W. Zheng, J. Butrynski, M. Capelletti, L. Wang, N.S. Gray, K. Wilner, J.G.
6 Christensen, G. Demetri, G.I. Shapiro, S.J. Rodig, M.J. Eck, P.A. Janne, The Neuroblastoma-Associated
7 F1174L ALK Mutation Causes Resistance to an ALK Kinase Inhibitor in ALK-Translocated Cancers,
8 Cancer Res. 70 (2010) 10038–10043. doi:10.1158/0008-5472.CAN-10-2956.
- 9 [16] A.T. Shaw, L. Friboulet, I. Leshchiner, J.F. Gainor, S. Bergqvist, A. Brooun, B.J. Burke, Y.-L. Deng, W.
10 Liu, L. Dardaei, R.L. Frias, K.R. Schultz, J. Logan, L.P. James, T. Smeal, S. Timofeevski, R. Katayama,
11 A.J. Iafrate, L. Le, M. McTigue, G. Getz, T.W. Johnson, J.A. Engelman, Resensitization to Crizotinib by
12 the Lorlatinib *ALK* Resistance Mutation L1198F, N. Engl. J. Med. 374 (2016) 54–61.
13 doi:10.1056/NEJMoa1508887.
- 14 [17] S. Kim, T.M. Kim, D.-W. Kim, H. Go, B. Keam, S.-H. Lee, J.-L. Ku, D.H. Chung, D.S. Heo,
15 Heterogeneity of Genetic Changes Associated with Acquired Crizotinib Resistance in ALK-Rearranged
16 Lung Cancer, J. Thorac. Oncol. 8 (2013) 415–422. doi:10.1097/JTO.0b013e318283dcc0.
- 17 [18] R.C. Doebele, A.B. Pilling, D.L. Aisner, T.G. Kutateladze, A.T. Le, A.J. Weickhardt, K.L. Kondo, D.J.
18 Linderman, L.E. Heasley, W.A. Franklin, M. Varella-Garcia, D.R. Camidge, Mechanisms of Resistance
19 to Crizotinib in Patients with ALK Gene Rearranged Non–Small Cell Lung Cancer, Clin. Cancer Res. 18
20 (2012). <http://clincancerres.aacrjournals.org/content/18/5/1472.long> (accessed April 6, 2017).
- 21 [19] G. Toyokawa, E. Inamasu, S. Shimamatsu, T. Yoshida, K. Nosaki, F. Hirai, M. Yamaguchi, T. Seto, M.
22 Takenoyama, Y. Ichinose, Identification of a Novel ALK G1123S Mutation in a Patient with ALK-
23 rearranged Non–small-cell Lung Cancer Exhibiting Resistance to Ceritinib, JTO Acquis. 10 (2015) e55–
24 e57. doi:10.1097/JTO.0000000000000509.
- 25 [20] H. Qiao, C.M. Lovly, Cracking the Code of Resistance across Multiple Lines of ALK Inhibitor Therapy
26 in Lung Cancer., Cancer Discov. 6 (2016) 1084–1086. doi:10.1158/2159-8290.CD-16-0910.
- 27 [21] S.-H.I. Ou, M. Azada, D.J. Hsiang, J.M. Herman, T.S. Kain, C. Siwak-Tapp, C. Casey, J. He, S.M. Ali,
28 S.J. Klempner, V.A. Miller, Next-Generation Sequencing Reveals a Novel NSCLC ALK F1174V

- 1 Mutation and Confirms ALK G1202R Mutation Confers High-Level Resistance to Alectinib
2 (CH5424802/RO5424802) in ALK-Rearranged NSCLC Patients Who Progressed on Crizotinib, J.
3 Thorac. Oncol. 9 (2014) 549–553. doi:10.1097/JTO.0000000000000094.
- 4 [22] H.Y. Zou, L. Friboulet, D.P. Kodack, L.D. Engstrom, Q. Li, M. West, R.W. Tang, H. Wang, K.
5 Tsaparikos, J. Wang, S. Timofeevski, R. Katayama, D.M. Dinh, H. Lam, J.L. Lam, S. Yamazaki, W. Hu,
6 B. Patel, D. Bezwada, R.L. Frias, E. Lifshits, S. Mahmood, J.F. Gainor, T. Affolter, P.B. Lappin, H.
7 Gukasyan, N. Lee, S. Deng, R.K. Jain, T.W. Johnson, A.T. Shaw, V.R. Fantin, T. Smeal, PF-06463922,
8 an ALK/ROS1 Inhibitor, Overcomes Resistance to First and Second Generation ALK Inhibitors in
9 Preclinical Models., Cancer Cell. 28 (2015) 70–81. doi:10.1016/j.ccell.2015.05.010.
- 10 [23] N.R. Infarinato, J.H. Park, K. Krytska, H.T. Ryles, R. Sano, K.M. Szigety, Y. Li, H.Y. Zou, N. V. Lee,
11 T. Smeal, M.A. Lemmon, Y.P. Mosse, The ALK/ROS1 Inhibitor PF-06463922 Overcomes Primary
12 Resistance to Crizotinib in ALK-Driven Neuroblastoma, Cancer Discov. 6 (2016) 96–107.
13 doi:10.1158/2159-8290.CD-15-1056.
- 14 [24] A. Passaro, A. Prelaj, A. Pochesci, G. Spitaleri, G. Rossi, E. Del Signore, C. Catania, F. de Marinis,
15 Brigatinib for the treatment of ALK-positive advanced non-small cell lung cancer patients, Drugs of
16 Today. 53 (2017) 435. doi:10.1358/dot.2017.53.8.2676119.
- 17 [25] S. Peters, D.R. Camidge, A.T. Shaw, S. Gadgeel, J.S. Ahn, D.-W. Kim, S.-H.I. Ou, M. Pérol, R.
18 Dziadziuszko, R. Rosell, A. Zeaiter, E. Mitry, S. Golding, B. Balas, J. Noe, P.N. Morcos, T. Mok,
19 Alectinib versus Crizotinib in Untreated ALK -Positive Non-Small-Cell Lung Cancer, N. Engl. J. Med.
20 (2017) NEJMoa1704795. doi:10.1056/NEJMoa1704795.
- 21 [26] B. Solomon, A. Shaw, S. Ou, B. Besse, E. Felip, T. Bauer, R. Soo, A. Bearz, C. Lin, J. Clancy, A.
22 Abbattista, H. Thurm, G. Peltz, E. Masters, J. Martini, L. James, T. Seto, OA 05.06 Phase 2 Study of
23 Lorlatinib in Patients with Advanced ALK⁺/ROS1⁺ Non-Small-Cell Lung Cancer, J. Thorac. Oncol. 12
24 (2017) S1756. doi:10.1016/j.jtho.2017.09.351.
- 25 [27] M. Duruisseaux, B. Besse, J. Cadranet, M. Pérol, B. Mennequier, L. Bigay-Game, R. Descourt, E. Dansin,
26 C. Audigier-Valette, L. Moreau, J. Hureauux, R. Veillon, J. Otto, A. Madroszyk-Flandin, A. Cortot, F.
27 Guichard, P. Boudou-Rouquette, A. Langlais, P. Missy, F. Morin, D. Moro-Sibilot, Overall survival with
28 crizotinib and next-generation ALK inhibitors in ALK-positive non-small-cell lung cancer (IFCT-1302

- 1 CLINALK): a French nationwide cohort retrospective study, *Oncotarget*. 8 (2017) 21903–21917.
2 doi:10.18632/oncotarget.15746.
- 3 [28] T. Mok, D. Spigel, E. Felip, F. deMarinis, M.-J. Ahn, H.J.M. Groen, H.A. Wakelee, T. Hida, L. Crino,
4 M. Nishio, G. V Scagliotti, F. Branle, C. Emeremni, M. Quadrigli, J. Zhang, A.T. Shaw, ASCEND-2: A
5 single-arm, open-label, multicenter phase II study of ceritinib in adult patients (pts) with ALK-
6 rearranged (ALK+) non-small cell lung cancer (NSCLC) previously treated with chemotherapy and
7 crizotinib (CRZ)., *J. Clin. Oncol.* 33 (2015) 8059. doi:10.1200/jco.2015.33.15_suppl.8059.
- 8 [29] S. Zhang, S. Nadworny, S.D. Wardwell, L. Eichinger, B. Das, E.Y. Ye, J.G. Hodgson, V.M. Rivera,
9 Abstract 781: The potent ALK inhibitor AP26113 can overcome mechanisms of resistance to first- and
10 second-generation ALK TKIs in preclinical models, *Cancer Res.* 75 (2015) 781–781. doi:10.1158/1538-
11 7445.AM2015-781.
- 12 [30] A.C. Johnson, P. Dô, N. Richard, C. Dubos, J.J. Michels, J. Bonneau, R. Gervais, Identification of
13 I1171N resistance mutation in ALK-positive non-small-cell lung cancer tumor sample and circulating
14 tumor DNA, *Lung Cancer.* 99 (2016) 38–40. doi:10.1016/j.lungcan.2016.06.010.
- 15 [31] S.-H.I. Ou, J. Greenbowe, Z.U. Khan, M.C. Azada, J.S. Ross, P.J. Stevens, S.M. Ali, V.A. Miller, B.
16 Gitlitz, I1171 missense mutation (particularly I1171N) is a common resistance mutation in ALK-positive
17 NSCLC patients who have progressive disease while on alectinib and is sensitive to ceritinib, *Lung*
18 *Cancer.* 88 (2015) 231–234. doi:10.1016/j.lungcan.2015.02.005.
- 19 [32] N. Tchekmedyan, S.M. Ali, V.A. Miller, E.B. Haura, Acquired ALK L1152R Mutation Confers
20 Resistance to Ceritinib and Predicts Response to Alectinib., *J. Thorac. Oncol.* 11 (2016) e87-8.
21 doi:10.1016/j.jtho.2016.03.018.
- 22 [33] A.T. Shaw, J.-F. Martini, B. Besse, T.M. Bauer, C.-C. Lin, R.A. Soo, G.J. Riely, S.-H.I. Ou, J.S. Clancy,
23 S. Li, H. Thurm, M. Satouchi, D.R. Camidge, S. Kao, R. Chiari, S. Gadgeel, E. Felip, B.J. Solomon,
24 Abstract CT044: Efficacy of lorlatinib in patients (pts) with advanced ALK-positive non-small cell lung
25 cancer (NSCLC) and ALK kinase domain mutations, *Cancer Res.* 78 (2018) CT044-CT044.
26 doi:10.1158/1538-7445.am2018-ct044.
- 27 [34] M. Miyawaki, H. Yasuda, T. Tani, J. Hamamoto, D. Arai, K. Ishioka, K. Ohgino, S. Nukaga, T. Hirano,
28 I. Kawada, K. Naoki, Y. Hayashi, T. Betsuyaku, K. Soejima, Overcoming EGFR Bypass Signal-Induced

- 1 Acquired Resistance to ALK Tyrosine Kinase Inhibitors in ALK-Translocated Lung Cancer, *Mol.*
2 *Cancer Res.* 15 (2017) 106–114. doi:10.1158/1541-7786.MCR-16-0211.
- 3 [35] A.S. Crystal, A.T. Shaw, L. V. Sequist, L. Friboulet, M.J. Niederst, E.L. Lockerman, R.L. Frias, J.F.
4 Gainor, A. Amzallag, P. Greninger, D. Lee, A. Kalsy, M. Gomez-Caraballo, L. Elamine, E. Howe, W.
5 Hur, E. Lifshits, H.E. Robinson, R. Katayama, A.C. Faber, M.M. Awad, S. Ramaswamy, M. Mino-
6 Kenudson, A.J. Iafrate, C.H. Benes, J.A. Engelman, Patient-derived models of acquired resistance can
7 identify effective drug combinations for cancer, *Science* (80-.). 346 (2014) 1480–1486.
8 doi:10.1126/science.1254721.
- 9 [36] C.M. Lovly, N.T. McDonald, H. Chen, S. Ortiz-Cuaran, L.C. Heukamp, Y. Yan, A. Florin, L. Ozretić,
10 D. Lim, L. Wang, Z. Chen, X. Chen, P. Lu, P.K. Paik, R. Shen, H. Jin, R. Buettner, S. Ansén, S. Perner,
11 M. Brockmann, M. Bos, J. Wolf, M. Gardizi, G.M. Wright, B. Solomon, P.A. Russell, T.-M. Rogers, Y.
12 Suehara, M. Red-Brewer, R. Tieu, E. de Stanchina, Q. Wang, Z. Zhao, D.H. Johnson, L. Horn, K.-K.
13 Wong, R.K. Thomas, M. Ladanyi, W. Pao, Rationale for co-targeting IGF-1R and ALK in ALK fusion–
14 positive lung cancer, *Nat. Med.* 20 (2014) 1027–1034. doi:10.1038/nm.3667.
- 15 [37] C.E. McCoach, A.T. Le, K. Gowan, K. Jones, L. Schubert, A. Doak, A. Estrada-Bernal, K.D. Davies,
16 D.T. Merrick, P.A. Bunn, W.T. Purcell, R. Dziadziuszko, M. Varella-Garcia, D.L. Aisner, D.R.
17 Camidge, R.C. Doebele, Resistance Mechanisms to Targeted Therapies in *ROS1*⁺ and *ALK*⁺ Non–small
18 Cell Lung Cancer, *Clin. Cancer Res.* (2018). doi:10.1158/1078-0432.CCR-17-2452.
- 19 [38] T. Tani, H. Yasuda, J. Hamamoto, A. Kuroda, D. Arai, K. Ishioka, K. Ohgino, M. Miyawaki, I. Kawada,
20 K. Naoki, Y. Hayashi, T. Betsuyaku, K. Soejima, Activation of EGFR Bypass Signaling by TGF α
21 Overexpression Induces Acquired Resistance to Alectinib in ALK-Translocated Lung Cancer Cells.,
22 *Mol. Cancer Ther.* 15 (2016) 162–71. doi:10.1158/1535-7163.MCT-15-0084.
- 23 [39] A.T. Shaw, D.-W. Kim, R. Mehra, D.S.W. Tan, E. Felip, L.Q.M. Chow, D.R. Camidge, J.
24 Vansteenkiste, S. Sharma, T. De Pas, G.J. Riely, B.J. Solomon, J. Wolf, M. Thomas, M. Schuler, G. Liu,
25 A. Santoro, Y.Y. Lau, M. Goldwasser, A.L. Borral, J.A. Engelman, Ceritinib in ALK-Rearranged Non–
26 Small-Cell Lung Cancer, *N Engl j Med.* 37013 (2014). doi:10.1056/NEJMoa1311107.
- 27 [40] S. Yoda, J.J. Lin, M.S. Lawrence, B.J. Burke, L. Friboulet, A. Langenbucher, L. Dardaei, K. Prutisto-
28 Chang, I. Dagogo-Jack, S. Timofeevski, H. Hubbeling, J.F. Gainor, L.A. Ferris, A.K. Riley, K.E.

1 Kattermann, D. Timonina, R.S. Heist, A.J. Iafrate, C.H. Benes, J.K. Lennerz, M. Mino-Kenudson, J.A.
2 Engelman, T.W. Johnson, A.N. Hata, A.T. Shaw, Sequential ALK Inhibitors Can Select for Lorlatinib-
3 Resistant Compound ALK Mutations in ALK-Positive Lung Cancer., *Cancer Discov.* (2018) CD-17-
4 1256. doi:10.1158/2159-8290.CD-17-1256.

5 [41] A. Kron, C. Alidousty, M. Scheffler, S. Merkelbach-Bruse, D. Seidel, R. Riedel, M.A. Ihle, S. Michels,
6 L. Nogova, J. Fassunke, C. Heydt, F. Kron, F. Ueckerth, M. Serke, S. Krüger, C. Grohe, D. Koschel, J.
7 Benedikter, B. Kaminsky, B. Schaaf, J. Braess, M. Sebastian, K.-O. Kambartel, R. Thomas, T. Zander,
8 A.M. Schultheis, R. Büttner, J. Wolf, Impact of TP53 mutation status on systemic treatment outcome in
9 ALK-rearranged non-small-cell lung cancer., *Ann. Oncol. Off. J. Eur. Soc. Med. Oncol.* 29 (2018)
10 2068–2075. doi:10.1093/annonc/mdy333.

11 [42] J.J. Lin, V.W. Zhu, S. Yoda, B.Y. Yeap, A.B. Schrock, I. Dagogo-Jack, N.A. Jessop, G.Y. Jiang, L.P.
12 Le, K. Gowen, P.J. Stephens, J.S. Ross, S.M. Ali, V.A. Miller, M.L. Johnson, C.M. Lovly, A.N. Hata,
13 J.F. Gainor, A.J. Iafrate, A.T. Shaw, S.-H.I. Ou, Impact of *EML4-ALK* Variant on Resistance
14 Mechanisms and Clinical Outcomes in *ALK* -Positive Lung Cancer, *J. Clin. Oncol.* 36 (2018) 1199–
15 1206. doi:10.1200/JCO.2017.76.2294.

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1 **Legends**

2 Table 1

3 Baseline characteristics

4

5 Figure 1

6 Distribution of mechanisms of resistance to ALK TKIs

7 * *One patient with a G1202R mutation was found, following treatment with lorlatinib, to*
8 *harbor a G1202R/G1269A compound mutation.*

9

10 Figure 2

11 Treatment of ALK-positive NSCLC patients. Swimming plot showing, for each patient, the
12 treatments received and the number, type(s), and time of completion of the biopsies
13 performed. Each bar represents one patient. Treatments received before the first ALK TKI
14 are not represented.

15

16 Figure 3

17 Clinical review of patients treated with a TKI after identification of an ALK mutation.
18 Presumed activity was defined by sensitive or intermediate IC50 based on *in vitro* data by
19 Gainor et al[11].

20

21

1 **Tables**

2 **Table 1**

3

	n = 23
Age, median (years) (range)	57 (17-70)
Sex, n (%)	
- Male	12 (52%)
- Female	11 (48%)
Adenocarcinoma, n (%)	23 (100%)
Stage at diagnosis, n (%)	
- Stage III	2 (9%)
- Stage IV	21 (91%)
Never Smokers, n (%)	19 (83%)
Number of lines of treatment before first ALK TKI, n (%)	
0	7 (30%)
1	13 (57%)
2 or more	3 (13%)
Type of ALK TKI received (n, %)	
1 st generation	22 (96%)
2 nd generation	17 (74%)
3 rd generation	3 (13%)

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Figure 1

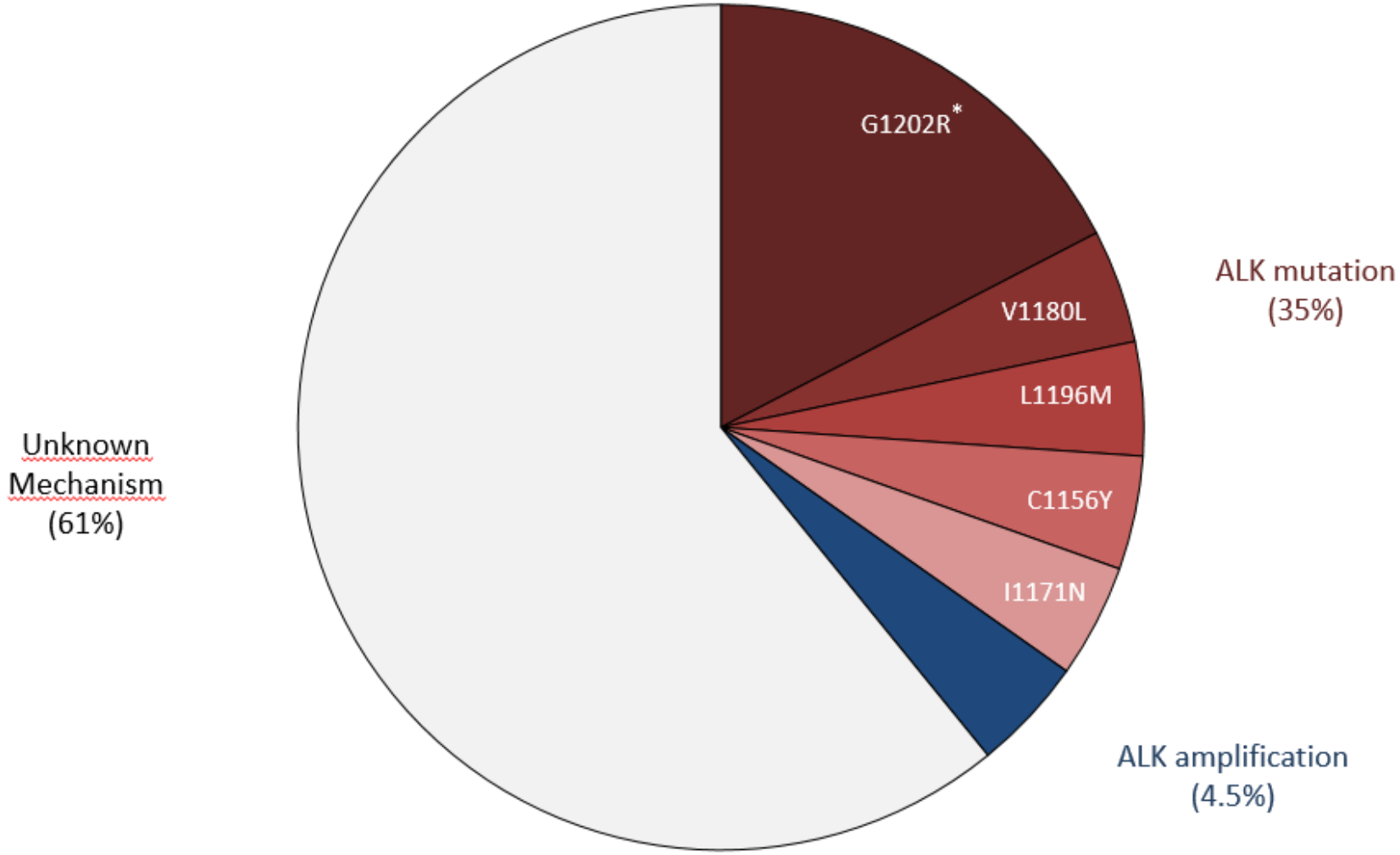


Figure 2

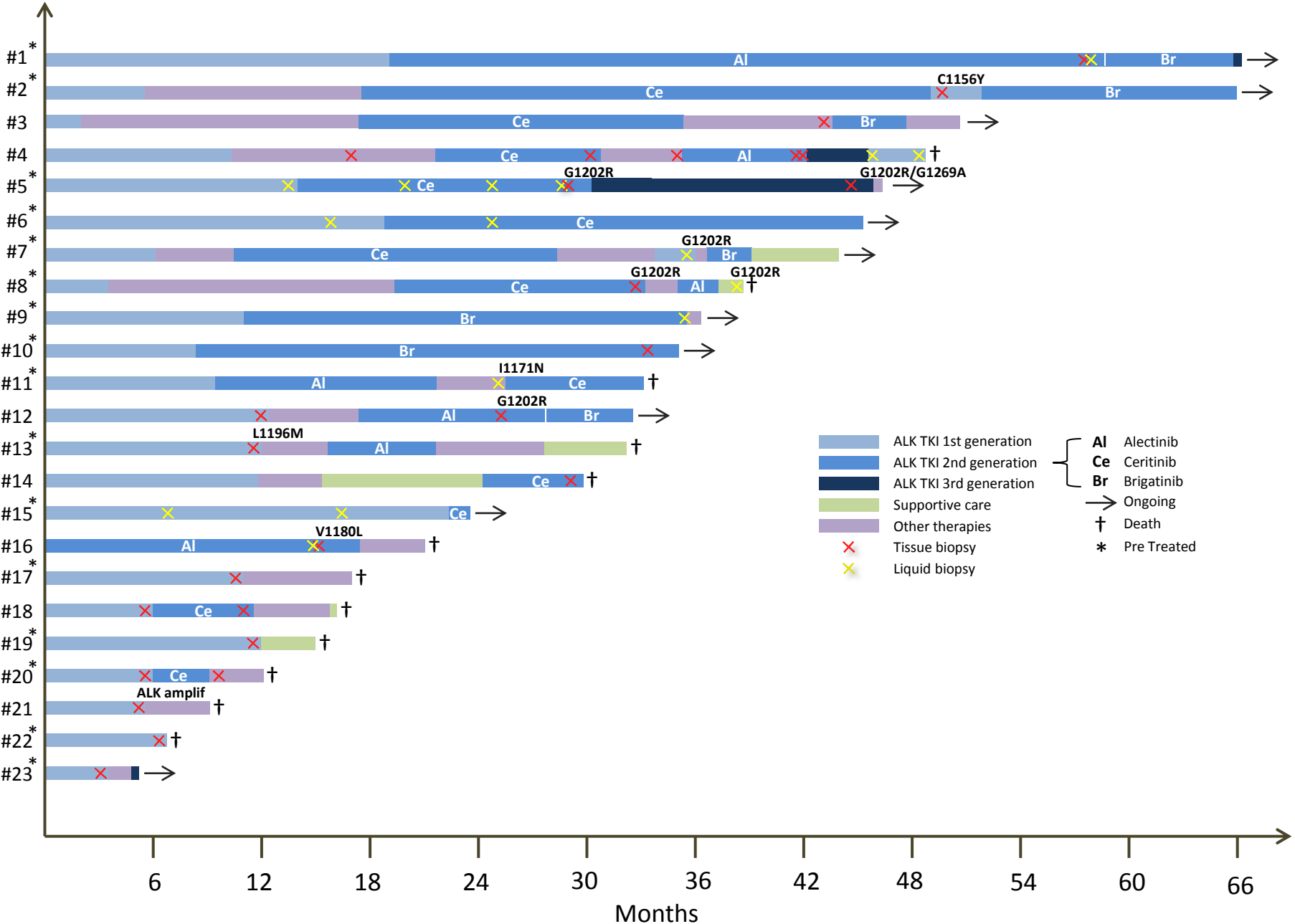
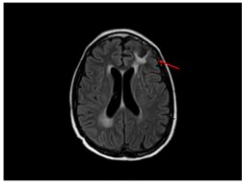
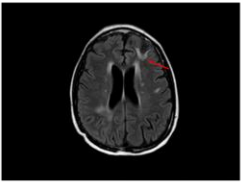


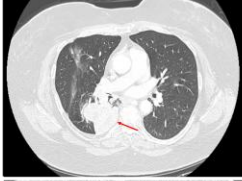



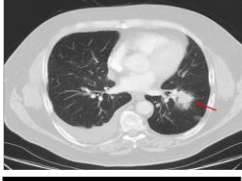

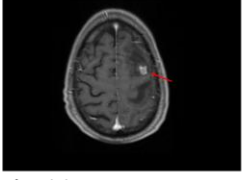
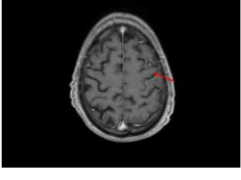




Figure 3

Identification number	Mutation and treatment	Pre treatment	Tumor response	First assessment	Duration of treatment
A. TKI with presumed activity					
#2	EML4/ALK C1156Y <u>Brigatinib</u>		Stable disease →		15.4 months (ongoing)
#13	EML4/ALK L1196M <u>Alectinib</u>		Partial Response →		5.8 months (stopped because of progression)
#6	EML4/ALK G1202R <u>Brigatinib</u>		Partial Response →		3.4 months (stopped because of toxicity)
#7	EML4/ALK G1202R <u>Lorlatinib</u>		Partial Response →		11.4 months (ongoing)
#12	EML4/ALK G1202R <u>Brigatinib</u>		Partial Response →		4.7 months (ongoing)
#11	EML4/ALK I1171N <u>Ceritinib</u>		Partial Response →		7.7 months (stopped because of progression)
B. TKI with no presumed activity					
#8	EML4/ALK G1202R <u>Alectinib</u>		Disease progression →		2.3 months (stopped because of progression)

Tables

Table 1

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