



**HAL**  
open science

## Acylcarnitines profile best predicts survival in horses with atypical myopathy

François Boemer, Johann Detilleux, Christophe Cello, Hélène Amory, Christel Marcillaud-Pitel, Eric Richard, Gaby van Galen, Gunther van Loon, Laurence Lefère, Dominique-Marie Votion

### ► To cite this version:

François Boemer, Johann Detilleux, Christophe Cello, Hélène Amory, Christel Marcillaud-Pitel, et al.. Acylcarnitines profile best predicts survival in horses with atypical myopathy. PLoS ONE, 2017, 12 (8), pp.e0182761. 10.1371/journal.pone.0182761 . hal-02167888

**HAL Id: hal-02167888**

**<https://normandie-univ.hal.science/hal-02167888v1>**

Submitted on 7 May 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH ARTICLE

# Acylcarnitines profile best predicts survival in horses with atypical myopathy

François Boemer<sup>1‡</sup>, Johann Detilleux<sup>2‡</sup>, Christophe Cello<sup>1</sup>, Hélène Amory<sup>3</sup>, Christel Marcillaud-Pitel<sup>4</sup>, Eric Richard<sup>5</sup>, Gaby van Galen<sup>3\*</sup>, Gunther van Loon<sup>6</sup>, Laurence Lefère<sup>6</sup>, Dominique-Marie Votion<sup>3\*</sup>

**1** Biochemical Genetics Laboratory, CHU Sart Tilman, University of Liege, Liege, Belgium, **2** Department of animal Productions: Biostatistics, Economy and animal selection, Fundamental and Applied Research for Animals & Health (FARAH), Faculty of Veterinary Medicine, University of Liege, Liege, Belgium, **3** Equine Pole, Fundamental and Applied Research for Animals & Health (FARAH), Faculty of Veterinary Medicine, University of Liege, Liege, Belgium, **4** Réseau d'Epidémiologie et de Pathologie Equine (RESPE), Caen, France, **5** Normandie Université, UNICAEN, Labéo Frank Duncombe, Caen, France, **6** Large Animal Internal Medicine, Gent University, Gent, Belgium

✉ Current address: Department of Large Animal Science, University of Copenhagen, Copenhagen, Denmark

‡ Co-first authors

\* [dominique.votion@ulg.ac.be](mailto:dominique.votion@ulg.ac.be)



**OPEN ACCESS**

**Citation:** Boemer F, Detilleux J, Cello C, Amory H, Marcillaud-Pitel C, Richard E, et al. (2017) Acylcarnitines profile best predicts survival in horses with atypical myopathy. PLoS ONE 12(8): e0182761. <https://doi.org/10.1371/journal.pone.0182761>

**Editor:** Carlos E. Ambrósio, Faculty of Animal Sciences and Food Engineering, University of São Paulo, BRAZIL

**Received:** January 30, 2017

**Accepted:** July 24, 2017

**Published:** August 28, 2017

**Copyright:** © 2017 Boemer et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its Supporting Information files.

**Funding:** The study was supported by the "Institut français du cheval et de l'équitation (Ifce)" of France and by "Les Fonds Spéciaux pour la Recherche (FSR)" of Liege University (Belgium). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

## Abstract

Equine atypical myopathy (AM) is caused by hypoglycin A intoxication and is characterized by a high fatality rate. Predictive estimation of survival in AM horses is necessary to prevent unnecessary suffering of animals that are unlikely to survive and to focus supportive therapy on horses with a possible favourable prognosis of survival. We hypothesized that outcome may be predicted early in the course of disease based on the assumption that the acylcarnitine profile reflects the derangement of muscle energetics. We developed a statistical model to prognosticate the risk of death of diseased animals and found that estimation of outcome may be drawn from three acylcarnitines (C2, C10:2 and C18 -carnitines) with a high sensitivity and specificity. The calculation of the prognosis of survival makes it possible to distinguish the horses that will survive from those that will die despite severe signs of acute rhabdomyolysis in both groups.

## Introduction

Equine atypical myopathy (AM) is caused by hypoglycin A intoxication resulting from the ingestion of seeds or seedlings of certain *Acer* tree species, principally *Acer pseudoplatanus* (sycamore maple) in Europe [1–3]. Ingested hypoglycin A is metabolized into methylenecyclopropyl acetic acid-CoA (MCPA-CoA), a potent inhibitor of acyl-CoA dehydrogenases. In the mitochondria, acyl-CoA dehydrogenases catalyse the initial step in each cycle of fatty acid  $\beta$ -oxidation and their reduced activity leads to accumulation of acyl-CoAs in the mitochondria. These acyl-CoAs may then be scavenged into acylcarnitines which subsequently leave the mitochondrion and reach the peripheral circulation leading to, in AM affected horses, a characteristic blood acylcarnitine profile consistent with a multiple acyl-CoA dehydrogenase

**Competing interests:** The authors have declared that no competing interests exist.

deficiency (MADD) phenotype [4]. Alternatively, MCPA may esterify with carnitine, forming MCPA-carnitine. Both hypoglycin A and MCPA-carnitine have been found in the serum of horses suffering from AM [2, 5–7].

Atypical myopathy has a high fatality rate with about three-quarter of affected horses dying with a mean survival time of less than two days [8]. Although the aetiology of AM has been uncovered, no antidote currently exists, but supportive treatment may be guided by knowledge of the underlying pathophysiological process [9, 10]. Treatment is costly, labour- and resource-intensive, and although treatment significantly increases an animal's probability of survival, outcome is currently difficult to predict on initial assessment of a case [11]. Medical treatment is warranted in selected cases where owners are motivated, but owners may first wish to know their horses' chances of survival and of return to normal sport capacity. From a large survey of European cases, we have learned that survival is not associated to a major risk of sequelae [8].

To determine the prognosis, veterinarians have four major steps in their clinical approach: collection of history, clinical examination, routine laboratory analyses and complementary examinations. Current knowledge concerning prediction of survival is mostly issued from a large-scale epidemiological study performed at a time when the cause was unknown; out of history, signalment, clinical variables and routine clinical pathology, few clinical signs and no biochemical parameters have been identified as predictive of survival [11]. In a more recent study focused solely on UK, a few clinical signs and high serum creatine kinase (CK) activity were associated with reduced odds of survival but only recumbency was retained in multivariate analysis [12]. Owing to recent developments, complementary examinations include the quantification of hypoglycin A [6, 13, 14] and MCPA-carnitine in blood and/or urine samples [5]. However, only a few laboratories offer testing for the hypoglycin A toxin and its principal metabolite, MCPA-carnitine.

We then hypothesized that, although AM survivors and non survivors are clinically almost similar on presentation, outcome may be predicted early in the course of disease based on serum acylcarnitines concentrations as these metabolites indicate the severity of dysfunction of mitochondrial  $\beta$ -oxidation. The aim of this study was to set up a statistical model in order to assess the survival prognosis at early stage of the disease.

## Material and methods

### Horses and sample collection

Blood samples collected from AM cases by the principal investigator between autumn 2006 and spring 2015 were used in this study. The tentative diagnosis of AM was based on previously documented inclusion criteria for the disease [8] which was subsequently confirmed by the identification of a MADD phenotype following determination of the acylcarnitines profile.

Case histories were obtained via a standardized questionnaire available on the Internet ([www.myopathie-atypique.be](http://www.myopathie-atypique.be)) and/or via owner interviews. Data on clinical signs were obtained from medical records. The first signs noted were those considered in this study.

Through routine practice, in the aim to establish a diagnosis and to design a supportive treatment, samples were collected from the jugular vein either at pasture by the referring practitioner ( $n = 9$ ) or on admission at hospital ( $n = 42$ ). Total duration of the blood sampling procedure did not exceed 1 min, and non-painful sampling was confirmed by the absence of any retreat behaviour of these critically-ill animals during the procedure. Blood was centrifuged immediately; the serum was separated and stored at  $-80^{\circ}\text{C}$  until analysis. It is important to underline that the samples taken the more precociously in the disease process on AM cases have been those considered in this study. Information on administered treatments and clinical outcome were extracted from clinical records and added to our database. Determination of the

acylcarnitine profile and the detection of hypoglycin A were conducted with the owner's consent in order to confirm the tentative diagnosis of atypical myopathy.

All hospitalized AM horses received supportive treatment consisting mainly of intravenous fluid therapy with isotonic poly-ionic fluids and glucose solutions with concurrent insulin administration, and supplementation of vitamins, antioxidants, and/or carnitine, and oral carbohydrates.

We have asked advice from Liege University's Animal Ethics Committee who has informed us that legal restrictions do not apply because no animals were handled specifically for this experiment. The procedures performed in this study are non-experimental and routine veterinary practices with livestock species (no laboratory animals).

## Routine laboratory analyses

Serum activities of CK, creatinine, total and ionized calcium ( $\text{Ca}_{\text{Tot}}$  and  $\text{Ca}^{2+}$ , respectively), electrolytes, total protein, haptoglobin, cardiac troponin I, serum amyloid A, triglycerides, glucose and lactate were determined by standard laboratory methods as well as complete blood cell count, haematocrit and arterial partial pressures in  $\text{O}_2$  ( $\text{PaO}_2$ ) on blood sampled on tubes with anticoagulant.

## Complementary biochemical examinations

Serum acylcarnitines concentrations were determined by tandem mass spectrometry [15]. Briefly, serum proteins were precipitated with a methanol solution containing labelled internal standards. Supernatants were evaporated under nitrogen stream and derivatized with butanolic-HCl. Butylated samples were analysed with a TQ5500 mass spectrometer (Sciex, Framingham, MA, USA). Reference ranges for acylcarnitines were derived from serum samples from 44 non-exposed healthy adult horses of various breeds and ages.

Quantification of hypoglycin A in defrosted blood sera was performed using a recently validated methodology using an aTRAQ kit for amino acid analysis of physiological fluids (Sciex, Framingham, MA, USA) [7, 16].

## Statistical analyses

Explanatory variables were grouped in 4 groups: historical data, results of physical examination, biochemical and haematological findings and, concentrations of free carnitine, acylcarnitines (short chain, medium chain and long chains) and hypoglycin A in serum. Statistical analyses were performed for each group separately.

In the first part of the analyses, chi-square and Student *t* tests were used to compare proportions and means between deceased and survivor horses, respectively. Kolmogorov-Smirnov tests were used to test the assumption that continuous explanatory variables were normally distributed. When necessary, they were transformed (Box Cox) so they follow the normal distribution.

All *P* values were corrected for multiple comparisons. However, results were only considered as indicative because Bonferroni correction tends to be too conservative.

In the second part of the analyses, PLS regression was used to handle the problem of the large number of collinear variables measured on a limited number of animals. The method also deals with missing data (EM algorithm). Five PLS components were considered and variables within these components were selected on the basis of their variable importance in the projection (VIP) and magnitude of the regression coefficients. Regression coefficients are for centred and standardized variables (proc PLS)

Variables selected to be included in the final model were those significant in the first part of the analyses ( $P \leq 0.05$ ), with a VIP  $> 1.0$  and a regression coefficient greater than 0.15. We set this strict criterion because of the large number of variables initially available.

The final model is a logistic regression based on which it was possible to compute the ROC curves [17] and threshold with the best sensitivity and specificity (highest Youden index).

## Results

### Individual horses, history and sample collection

During the study period, 51 horses suspected of AM and for which blood samples were available were confirmed to be AM cases based on a MADD phenotype. From these 51 horses, 40 deceased or were euthanized (AM-D group) and 11 survived (AM-S group). Most animals were treated at the faculty of veterinary medicine of Liege or Ugent University (Belgium); seven non survivors and two survivors were treated at their home premises. Blood samples of horses treated at pasture were collected by practitioners on site.

In the AM-D group, 80% of horses were euthanized. After 48 hours, 72.5% of the AM-D group was no longer alive. The duration of disease differed significantly between both groups ( $2.38 \pm 0.25$  days versus  $12.00 \pm 1.48$  days in the AM-D and AM-S group, respectively).

Demographic data are shown in Table 1. Survivors were significantly older than non survivors and frequency distribution of ages indicated that 50% of AM-S were 7 years old or older whereas 64% of AM-D were 4 years old or younger. No significant differences were found among other demographic data. Following Bonferroni correction, the mean age and duration of disease remained significantly different between both groups.

### Clinical signs

Frequency distributions of clinical signs are shown in Table 2. Horses of the AM-S more frequently showed signs of muscle tremors (localized or generalized) and more often remained in

**Table 1. Mean age and frequency distribution of demographic data of horses with atypical myopathy.**

	<i>Non survivors (n = 40)</i>	<i>Survivors (n = 11)</i>
Age (years $\pm$ SEM)	4.7 $\pm$ 0.6	8.9 $\pm$ 1.9*
Type		
<i>Saddle horses</i>	78%	64%
<i>Ponies</i>	20%	27%
<i>Draught horses</i>	3%	9%
Sex		
<i>Fillies and mares</i>	58%	55%
<i>Colts and stallions</i>	18%	9%
<i>Geldings</i>	25%	36%
Body condition		
<i>Thin</i>	0%	10%
<i>Normal weight</i>	94%	80%
<i>Overweight</i>	6%	10%
Use for work	36%	43%

Abbreviation: SEM = standard error of the mean

\* significantly different with a  $P$  value  $\leq 0.05$

<https://doi.org/10.1371/journal.pone.0182761.t001>

**Table 2. Frequency distribution of clinical signs in horses with atypical myopathy.**

Clinical signs	Non survivors (n = 40)	Survivors (n = 11)
	Relative frequency	
Pigmenturia	94%	100%
Stiffness	76%	90%
Depressed mental status	76%	70%
Weakness	74%	90%
Want to eat	70%	71%
Trembling	69%	100%**
Recumbency	<b>63%</b>	44%
Sweating	<b>65%</b>	78%
Pain		
<i>Severe</i>	50%	17%
<i>Medium</i>	35%	33%
<i>Slight</i>	5%	0%
<i>No pain</i>	10%	50%**
Remaining standing	38%	<b>70%*</b>
Signs of colic	31%	40%
Oesophageal obstruction and/or dysphagia	33%	10%*
Anorexia	<b>10%</b>	11%

**Comments:** the clinical signs highlighted in bold are those that were previously found to be of prognostic value for non survival (in bold in the non survivors column) or survival (in bold in the column of survivors) in a large European survey [11].

\* significantly different with a *P* value ≤ 0.05

\*\* significantly different with a *P* value ≤ 0.01

<https://doi.org/10.1371/journal.pone.0182761.t002>

standing position. Half of horses of the AM-D group were in severe pain whereas horses of the AM-S group often didn't show signs of pain. Oesophageal obstruction and/or dysphagia were more frequently observed in the AM-D group.

Clinical examination results are reported in Tables 3 and 4. The AM-D group more frequently showed cyanotic mucous membranes whereas capillary refill time was increased in both groups. Even though the mean rectal temperature was normal in both groups, AM-S horses were more frequently hyperthermic. Respiratory difficulties (*i.e.*, tachypnoea and expiratory dyspnoea) were equally seen in both groups. Heart murmurs were heard in both groups but arrhythmias were only detected in the AM-D group. Horses of the AM-S group had normal or diminished gut sounds whereas complete absence of gut sounds occurred in the AM-D group. None of the above variables remained significant following Bonferroni correction; therefore no variables related to clinical examination fulfilled the criteria to be included in a logistic regression to predict the prognosis. So, AM survivors and non survivors were almost clinically similar on presentation.

### Routine laboratory analyses

Laboratory findings are presented in Table 5. Several biochemical and haematological parameters were outside the laboratory's reference range in both groups, but the AM-D group differed from the AM-S by higher CK activities, lower sodium concentration, lower total calcium (Ca<sub>Tot</sub>) and higher total white blood cell counts. One AM-D horse had a mildly increased CK

**Table 3. Results of the clinical examination of horses with atypical myopathy.**

Parameters Relative frequency (%)	Non survivors (n = 40)	Survivors (n = 11)
Mucous membranes		
<i>Normal</i>	14%	<b>22%</b>
<i>Congested</i>	69%	78%
<i>Cyanotic</i>	11%	0%**
<i>Icteric</i>	6%	0%
Temperature		
<i>Normothermia (37–38.5°C)</i>	76%	<b>56%</b>
<i>Hypothermia (&lt; 37°C)</i>	24%	33%
<i>Hyperthermia (&gt; 38.5°C)</i>	0%	11%**
Type of respiration		
<i>Normal rate (&lt; 15 Breaths.min<sup>-1</sup>)</i>	23%	22%
<i>Tachypnoea (≥ 15 Breaths.min<sup>-1</sup>)</i>	<b>77%</b>	78%
<i>Dyspnoea</i>	<b>55%</b>	55%
Abnormal cardiac auscultation		
<i>Murmur</i>	32%	40%
<i>Arrhythmia</i>	10%	0%*
<i>Tachycardia (≥ 45 Beats.min<sup>-1</sup>)</i>	<b>92%</b>	90%
Gut sounds		
<i>Absent</i>	18%	0%**
<i>Diminished</i>	52%	50%
<i>Normal</i>	28%	50%
<i>Increased</i>	3%	0%
Miscellaneous		
<i>Full bladder on palpation</i>	58%	50%
<i>Abnormal rectal examination</i>	33%	29%
<i>Defaecation</i>	71%	<b>50%</b>

**Comments:** the clinical signs highlighted in bold are those that were previously found to be of prognostic value for non-survival (in bold in the non survivors column) or survival (in bold in the column of survivors) in a large European survey [11]

\* significantly different with a *P* value ≤ 0.05

\*\* significantly different with a *P* value ≤ 0.01

<https://doi.org/10.1371/journal.pone.0182761.t003>

(800 IU/L) only; the horse in question was sampled three hours before onset of clinical signs as it was a pasture companion of another case. For this reason, we elected to remove this horse’s CK from the dataset; this had no effect on results of statistical analyses. Following Bonferroni correction, only Ca<sub>Tot</sub> remained significantly different between both groups.

### Ancillary biochemical parameters

Hypoglycin A was detected in serum samples of all horses affected with AM. There was no significant difference in serum levels between survivors and non survivors (4.25±0.49 µmol/L in deceased horses versus 2.67±0.63 µmol/L in survivors; mean ±SD; Fig 1). Hypoglycin A was not found in any blood sample taken from 8 control horses (absence of chromatographic peak).

As presented in Table 6, acylcarnitine profiles were sorted into 3 groups: short, medium and long chain acylcarnitines, corresponding to the metabolic pathways in which they are



**Table 4. Results of the clinical examination of horses with atypical myopathy.**

Parameters (Mean ±SD)	Normal range	Non survivors (n = 40)	Survivors (n = 11)
Rectal temperature (°C)	37–38.5	37.3±0.1	37.2±0.2
Respiratory rate (Breaths.min <sup>-1</sup> )	<15	25.3±1.9*	24.9±2.4*
Heart rate (Beats.min <sup>-1</sup> )	<45	62.8±2.2*	62.8±3.8*
Capillary refill time (s)	≤2	2.8±0.1*	3.0±0.2*

Abbreviation: SD = standard error of the mean

\* Value outside of reference range.

<https://doi.org/10.1371/journal.pone.0182761.t004>

involved. Short chains acylcarnitines are markers of both short chain fatty acids  $\beta$ -oxidation and some amino acids catabolism. Medium chains are markers of medium chain fatty acids  $\beta$ -oxidation while long chains are the reflection of both long chain fatty acids  $\beta$ -oxidation and long chain fatty acids transport into mitochondria (*i.e.* carnitine palmitoyltransferase I and

**Table 5. Biochemical and haematological findings in horses with atypical myopathy (Mean ±SD).**

Parameters	Normal values	Non survivors	No. of dosages (max 40)	Survivors	No. of dosages (max 11)
<b>Muscle enzymes</b>					
CK (IU/L)	50–200	255,586±37,282*	40	126,877±41,879* <sup>a</sup>	11
<b>Renal functions and ions</b>					
Creatinine (μmol/L)	44–140	118±10	25	94±12	8
Sodium (mmol/L)	130–145	135±1	30	141±1 <sup>b</sup>	7
Potassium (mmol/L)	3.0–4.7	3.93±0.10	28	4.04±0.19	7
Chloride (mmol/L)	95–110	94±1*	27	97±2	7
Ca <sub>Tot</sub> (mmol/L)	2.50–3.40	2.34±0.04*	19	2.72±0.06 <sup>b</sup>	7
Ca <sup>2+</sup> (mmol/L)	> 1.50	1.24±0.06*	14	1.37±0.04*	4
<b>Proteins</b>					
Total protein (g/L)	57–69	73±2*	30	79±3*	9
Haptoglobin (mg/L)	< 500	1,702±116*	20	1,416±167*	7
Troponin I (ng/L)	10–30	4,121±984*	21	3,146±1,538*	7
Serum amyloid A (mg/L)	< 5.0	293±24*	18	303±43*	7
<b>Energetic metabolism</b>					
Triacylglycerols (mmol/L)	< 0.97	2.55±0.28*	28	1.95±0.54*	8
Glycaemia (mmol/L)	3.33–5.55	10.68±0.76*	16	9.01±0.99*	6
Lactate (mmol/L)	< 4	6.5±0.7*	23	5.1±1.3*	7
<b>Haematology</b>					
Hematocrit (%)	32–48	47±1	31	46±2	8
Total white blood cells (10 <sup>9</sup> /L)	6–12	12.5±0.5*	23	9.3±0.6 <sup>a</sup>	5
<b>Arterial blood</b>					
PaO <sub>2</sub> (mmHg)	> 85	86±3	15	91±3	3

Abbreviations: CK, creatine kinase; Ca<sub>Tot</sub> and Ca<sup>2+</sup>, total and ionized calcium, respectively; PaO<sub>2</sub> arterial partial pressures in O<sub>2</sub>; SD = standard deviation

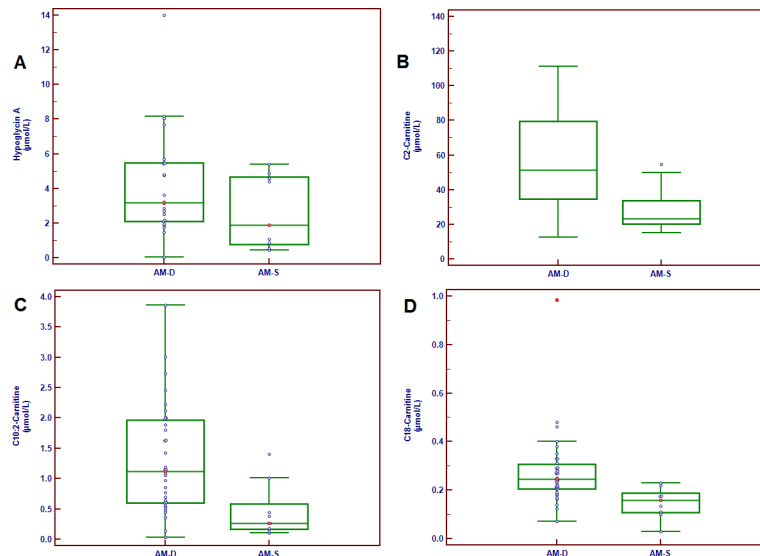
<sup>a</sup> Superscripts indicate that the mean was statistically higher in non survivors *versus* survivors (*t* student or chi-square;  $P \leq 0.05$ ).

<sup>b</sup> Superscripts indicate that the mean was statistically lower in non survivors *versus* survivors (*t* student or chi-square;  $P \leq 0.05$ ).

\* Value outside of reference range.

<https://doi.org/10.1371/journal.pone.0182761.t005>





**Fig 1.** Box plot of (a) hypoglycin A, (b) C2-Carnitine; (c) C10:2-Carnitine, and (d) C18-Carnitine concentrations in sera of atypical myopathy horses among them 11 survived and 40 deceased (hypoglycin A was only measured on 9 survivors and 22 deceased horses). Comments: The central box represents the values from the lower to upper quartile (25 to 75 percentile). The middle line represents the median. A line extends from the minimum to the maximum value, excluding “outside” values (value that is smaller/higher than the lower/upper quartile  $\pm 1.5$  times the interquartile range) or “far out” values (value that is smaller/higher than the lower/upper quartile  $\pm 3$  times the interquartile range) which are displayed as separate points or red marks, respectively.

<https://doi.org/10.1371/journal.pone.0182761.g001>

II, . . .). Mean serum concentrations of all acylcarnitines of the AM-D and AM-S groups were outside percentile 97.5 of reference ranges obtained from healthy control horses. Based on the results, acylcarnitines were classified into 2 subcategories, namely:

- Acylcarnitines which were outside of reference ranges in both AM-S and AM-D but with no significantly different mean serum concentration between the two groups (C3, C6, C8, C10, C10:1, C12, C12:1, C14, C14:1, C16 and C16:1 -carnitine).
- Acylcarnitines which were outside of reference ranges in both AM-S and AM-D and with a mean serum concentration significantly higher in AM-D versus AM-S horses (C2, C3DC, C4, C5, C5-OH, C5DC, C8:1, C10:2, C18 and C18:1 -carnitine).

Acylcarnitines from the first category can be used to confirm a diagnosis of AM but cannot discriminate survivors versus non survivors. The same is true for free carnitine. Acylcarnitines from the second category can also be used to confirm a diagnosis of AM but in addition they are informative regarding prognosis.

### Survival prognosis

Best prognostic factors in each of the four major steps of the clinical approach are listed in [Table 7](#).

Horses younger than 5.25 years have 87.1% chance of dying. This threshold of 5.25 years yields the highest sensitivity (SE) of 64% and specificity (SPE) of 67%. Horses that were still alive at day 3 from onset of clinical sings have 50% chance of surviving.

**Table 6. Serum (mean ±SD) concentrations of free carnitine and acylcarnitines (μmol/L) in horses with atypical myopathy and control horses.**

	Control horses (n = 44)	Horses with AM Non survivors (n = 40)	Horses with AM Survivors (n = 11)
<b>Free carnitine</b>			
Free carnitine	26.02±1.07	87.04±7.95*	74.41±18.07*
<b>Short chain acylcarnitines (C2 to C5)</b>			
C2-Carnitine	8.77±0.67	55.23±4.24*	36.94±9.28* <sup>a</sup>
C3-Carnitine	0.56±0.03	2.92±0.27*	2.21±0.52*
C3DC-Carnitine	0.03±0.00	0.34±0.04*	0.11±0.02* <sup>a</sup>
C4-Carnitine	0.36±0.02	26.62±3.67*	12.96±4.06* <sup>a</sup>
C5-Carnitine	0.21±0.01	24.71±3.20*	9.32±2.84* <sup>a</sup>
C5-OH-Carnitine	0.06±0.00	0.56±0.07*	0.25±0.05* <sup>a</sup>
C5DC-Carnitine	0.18±0.01	2.20±0.28*	1.10±0.25* <sup>a</sup>
<b>Medium chain acylcarnitines (C6 to C10)</b>			
C6-Carnitine	0.04±0.00	5.40±0.81*	2.93±1.29*
C8-Carnitine	0.02±0.00	1.45±0.18*	1.41±0.67*
C8:1-Carnitine	0.09±0.01	1.26±0.15*	0.44±0.15* <sup>a</sup>
C10-Carnitine	0.02±0.00	0.76±0.09*	1.01±0.51*
C10:1-Carnitine	0.05±0.01	0.67±0.07*	0.44±0.07*
C10:2-Carnitine	0.03±0.00	1.31±0.14*	0.46±0.13* <sup>a</sup>
<b>Long chain acylcarnitines (C12 to C18)</b>			
C12-Carnitine	0.04±0.01	0.28±0.03*	0.38±0.17*
C12:1-Carnitine	0.02±0.00	0.23±0.03*	0.37±0.17*
C14-Carnitine	0.03±0.00	0.21±0.02*	0.25±0.11*
C14:1-Carnitine	0.03±0.00	0.32±0.04*	0.48±0.23*
C16-Carnitine	0.14±0.04	0.68±0.07*	0.52±0.18*
C16:1-Carnitine	0.02±0.00	0.24±0.03*	0.17±0.08*
C18-Carnitine	0.07±0.01	0.28±0.02*	0.15±0.02* <sup>a</sup>
C18:1-Carnitine	0.04±0.01	0.57±0.06*	0.30±0.12* <sup>a</sup>

Abbreviation: SD = standard deviation

\*Mean outside percentile 97.5 of reference range obtained with control horses

<sup>a</sup> Superscripts indicate that the mean was statistically higher in non survivors versus survivors (*t* student or chi-square; *P* ≤ 0.05).

<https://doi.org/10.1371/journal.pone.0182761.t006>

The Ca<sub>Tot</sub> is the most relevant routine laboratory marker to predict prognosis. Affected animals with Ca<sub>Tot</sub> lower than 2.69 mmol/L have a 70% chance of dying. This threshold yields the best SPE (92%) and SE (64%).

According to partial least square (PLS) and logistic regressions, C2, C10:2 and C18–carnitine appeared to be the best parameters to predict survival among the different markers selected (Table 7 and Fig 1). Neither Ca<sub>Tot</sub>, nor age, nor hypoglycin A were retained as significant markers by the algorithm.

The model that best predicted probability of AM horses survival is:

$$\text{Logit}(p) = b_0 + b_1 \text{C2 - Carnitine} + b_2 \text{C10 : 2 - Carnitine} + b_3 \text{C18 - Carnitine}$$

where

b0 = 5.38 (standard error: 2.31)

b1 = -0.06 (standard error: 0.03)

**Table 7. Best prognostic predictors in the four principal components of a clinical approach.**

Parameters	Probability	AUC	Sensitivity	Specificity
<b>In history</b>				
Age	if age < 5.25 years = 87.1% chance of dying	0.79	0.64	0.67
<b>In results of clinical examination</b>				
None				
<b>In routine laboratory analyses</b>				
Ca <sub>Tot</sub>	if < 2.69 mmol/L = 70.0% chance of dying	0.89	0.64	0.92
<b>In complementary biochemical examinations</b>				
C2	if > 17.31 μmol/L = 80.4% chance of dying	0.74	0.82	0.07
C10:2	if > 0.11 μmol/L = 80.9% chance of dying	0.84	0.73	0.15
C18	if > 0.21 μmol/L = 92.6% chance of dying	0.86	0.18	0.37

Abbreviation: AUC = area under the curve

<https://doi.org/10.1371/journal.pone.0182761.t007>

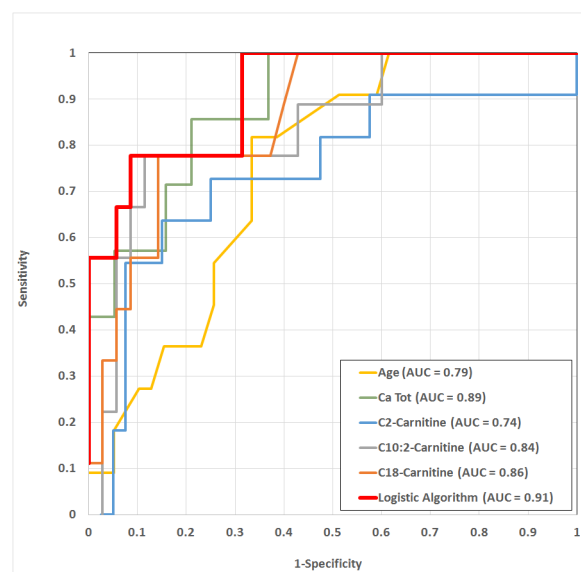
b2 = -0.55 (standard error: 1.01)

b3 = -18.94 (standard error: 9.61)

C2, C10:2 and C18 -Carnitine units: μmol/L

Compared to each individual marker, the corresponding ROC curves demonstrate that the logistic algorithm provides the best sensitivity and specificity, with an area under the curve (AUC) rising to 0.91 (Fig 2).

This multivariate approach has subsequently been validated against independent AM animal's cohorts. Acylcarnitines have been quantified on eight AM horses from Denmark (4 survivors and 4 deceased) [18] and four animals from New-Zealand (2 survivors and 2 deceased) [19]. Except for the case with mildly elevated CK that was collected before symptoms development, all samples were collected the more precociously after symptoms apparition to analyze



**Fig 2. ROC curves comparison (with their corresponding AUC) for age, Ca<sub>Tot</sub>, C2-carnitine, C10:2-Carnitine, C18-Carnitine and the logistic algorithm in the estimation of survival prognosis.** Abbreviation: AUC = area under the curve.

<https://doi.org/10.1371/journal.pone.0182761.g002>

acylcarnitines profile. Concentrations of C2, C10:2 and C18 -Carnitine have been then introduced in the logistic equation to predict their outcome. Estimation of survival probability observed for these animals was in agreement with our expectations: survivors and deceased equines were respectively classified with high and low probability of survival (Table 8).

### Discussion

In this study, factors that enable differentiation between AM cases that are unlikely and those that are likely to survive were identified. Prognostic factors were sought among the four pivotal steps of most clinical approaches, *i.e.* historical data, clinical examination, routine clinical pathology and ancillary testing.

The only historical risk factor associated with a fatal outcome in our study was a young age, which has previously been found to be associated with an increased risk of being affected by AM. [20] This finding suggests that younger individuals have a higher sensitivity to the toxin. As atypical myopathy results from a muscle energetics imbalance, the higher susceptibility of youngsters might be a reflection of higher energetic needs in growing horses.

In a large exploratory data analysis of European cases, negative prognostic factors were recumbency, sweating, anorexia, tachycardia, tachypnoea and dyspnoea [11]. Positive prognostic factors were: remaining standing most of the time, normothermia, normal mucous membranes, presence of defaecation, and vitamin / antioxidant therapy [11]. In the current study, all horses received similar supportive treatment, therefore the effect of treatment could not be assessed but remaining standing up, normal rectal temperature, normal mucous membrane colour and presence of digestive sounds were also found to be associated with a good prognosis. In addition, other clinical parameters were identified as positive prognosis factors such as the absence of signs of pain, a lower frequency of oesophageal obstruction and/or dysphagia and absence of cardiac arrhythmias. As none of the clinical parameters, in the present study, remained significantly different following Bonferroni correction, they cannot be used to predict prognosis with any certainty and should be used with caution in clinical decision-making.

However, it can be stated that the longer the horse survives, the more the outcome becomes positive. Therefore, it may be warranted in future cases attempt to treatment, for at least two

**Table 8. Survival prognosis estimated on independent cohort of horses with atypical myopathy [17, 18].** Prognosis varies from 0 to 1; the closer the value is to zero, the higher the risk of mortality. Conversely, the closer the value approaches 1, the greater the chance of survival. Low mortality risk is marked in bold.

Animal with atypical myopathy	Survival probability
Denmark—AM-S—1	<b>0.512</b>
Denmark—AM-S—2	<b>0.985</b>
Denmark—AM-S—3	<b>0.836</b>
Denmark—AM-S—4	<b>0.250</b>
Denmark—AM-D—1	0.000
Denmark—AM-D—2	0.013
Denmark—AM-D—3	0.024
Denmark—AM-D—4	0.026
New-Zealand—AM-S—1	<b>0.847</b>
New-Zealand—AM-S—2	<b>0.975</b>
New-Zealand—AM-D—1	0.001
New-Zealand—AM-D— 2	0.011

Abbreviations: AM-S: horse suffering from atypical myopathy who survived; AM-D: horse suffering from atypical myopathy who deceased or was euthanized

<https://doi.org/10.1371/journal.pone.0182761.t008>

days from onset of clinical signs if there is no considerable pain or cardiovascular compromise (such as indicated by tachycardia, cardiac arrhythmias and/ or cyanotic mucous membranes) or when they can be reverted by the initial supportive therapy, and if the horse can be fed with a diet rich in carbohydrates in the absence of oesophageal obstruction and/or dysphagia. Recommended supportive therapy for AM is extensively described elsewhere [9, 10]. However, if respiratory difficulties arise or progressively worsen euthanasia may be indicated [21, 22]. In our AM-D group, horses were euthanized because of severe expiratory dyspnoea and/ or compromised gas exchanges as indicated, when measured, by low arterial partial pressures in O<sub>2</sub> and increased arterial partial pressures in CO<sub>2</sub>.

In this study, severe biochemical disorders were found. In particular, massive muscle cell destruction was associated with electrolyte abnormalities and high CK activities in serum. The determination of Ca<sub>Tot</sub> has predictive value for survival. The hypocalcaemia frequently found in AM-affected horses might contribute to dysrhythmias observed in some horses [23]. Intravenous calcium supplementation may be warranted to maintain or restore serum ionized calcium concentration in AM-affected horses [9].

In rat, hypoglycin A is rapidly metabolized *in vivo* [24] and may be undetectable in biological samples. The same is true for human patients suffering from Jamaican vomiting sickness [25], a disease resulting from the ingestion of unripe ackee fruit which contains hypoglycin A [26]. The kinetics of hypoglycin A in horses are currently unknown but high levels have been detected in apparently healthy horses [1, 5]. Also, a wide range of hypoglycin A concentrations was found both in the group of AM-D and AM-S horses whereby mean serum concentration was not significantly different between the two groups. Therefore, detection of hypoglycin A in serum can be considered a biomarker of exposure but has no prognostic value.

Metabolism of hypoglycin A produces the toxic metabolite MCPA which may form ester with carnitine but also with glycine, two important detoxification forms which are excreted in urine [27]. Recently, a method allowing the determination of acylcarnitines and MCPA conjugates in small samples of serum and urine has been validated [6]. The level of MCPA-conjugates might contribute to both diagnosis and prognostication of AM and our predictive model might be further improved by including MCPA-carnitine concentration. MCPA-glycine which is more readily excreted in the urine is less likely to be a valuable serum parameter for estimation of survival. It is unknown which serum concentrations of MCPA need to be reached to induce signs of AM. Since subclinical cases occur [5, 21, 28], we hypothesise that there is a dose-dependent action similar to other species [29] and that the toxic threshold may vary depending on the current energy balance of the horse.

MCPA-CoA inhibits short and medium chain acyl-CoA dehydrogenases [30–34] and reduces several cofactors, such as coenzyme A and carnitine, which are essential to the  $\beta$ -oxidation of long-chain fatty acids thus inhibiting their transport into the mitochondria [35]. Consequently, fatty acids conjugated with carnitines accumulate in the serum. In horses, as obtaining a blood sample is easier than urine collection and because long chain acylcarnitines are not excreted in urine, it was therefore decided to search for the best prognostic biomarkers in blood among short, medium and long chain acylcarnitines thus covering the whole process associated with alteration of the  $\beta$ -oxidation of fatty acids. Also, acylcarnitines profiling is feasible in a large numbers of laboratory facilities, while MCPA-conjugates may only be analysed in a few laboratories that have developed specific methods for diagnosis of hypoglycin A intoxication.

The striking finding of this study is the ability to specify a prognosis based on early blood testing, at the start of the clinical examination by the practitioner or even at a sub-clinical state as shown by one horse that was sampled 3 hours before clinical signs became apparent and which died as would have been predicted based on its serum levels of C2, C10:2 and, C18 -Carnitine (with a prognosis for survival estimated at 0.3%). Considered individually, C2-Carnitine

has good sensitivity for determination of both diagnosis and prognosis, but it is unfortunately a very unspecific marker of ketosis. C10:2-Carnitine is increased in 2,4-dienoyl-CoA reductase deficiency while C18-Carnitine reflects either translocase or carnitine palmitoyltransferase II. Considered all together, these three parameters are unspecific markers of MADD but revealed the highest sensitivity in mortality prediction.

To date, no therapy is effective to cure AM and care is essentially supportive. Many animals die during the course of intoxication and euthanasia often appears as the ultimate veterinary's expedient. However, some intoxicated horses survive despite a dramatic initial clinical picture. In this context, survival prognosis could be of critical interest to prevent euthanasia. Indeed, intensive care may be focused on animals with good survival expectancy while individuals with high death probability could be sacrificed more "pragmatically" when there are signs of suffering that cannot be suppressed. Prognosis of survival may additionally be useful when testing new therapies, in order to evaluate their efficacy. Effect of any new treatment should be relativized if the new drug appears only effective in animals with an initial high predictive value of survival. Contrarily, the efficacy of any new protocol could be highly consolidated if mortality prognosis is initially elevated.

Based on the results of this study, we recommend practitioners to pose an initial tentative diagnosis of AM based on history (*i.e.* acute signs of rhabdomyolysis in horses at pasture with possible hypoglycin A ingestion), clinical examination (*i.e.* pigmenturia being the most specific but not pathognomonic signs) and routine laboratory tests (*i.e.* CK activities but also  $Ca_{Tot}$  and other biochemical parameters which are also useful to correct electrolyte imbalances). Confirmation of the diagnosis is best performed by determining acylcarnitines profile and hypoglycin A or MCPA-carnitine concentration in order to confirm exposure to the etiological agent. And finally, the best evaluation of survival prognosis is achieved by integrating the levels of C2, C10:2 and C18:1 -Carnitine into the proposed algorithm. To date, acylcarnitines profile analysis is generally performed in a research context in only few specific laboratories, without any "emergency" consideration. Our results provide however a major incentive for such laboratories to implement, in their own organization, the analysis of corresponding samples on a daily basis to provide prompt and comprehensive diagnostics.

Regardless of eventual outcome of AM, its clinical manifestations are severe signs of acute rhabdomyolysis of mostly postural and respiratory muscles and the myocardium, which all utilise fatty acids as a primary energy source. We developed a scoring system to predict survival in AM affected cases based on early blood sampling. The scoring system proposed in this study may help clinicians with management decisions including euthanasia. In further cases, we sought to apply this procedure and correlate results of this equation with outcomes.

## Supporting information

**S1 Table. Original dataset is available as supporting file of the manuscript.**  
(XLSX)

## Acknowledgments

The members of the AMAG, all clinicians of the equine clinics from Liege and Ugent university hospitals, and all communicating veterinarians and owners of affected horses are gratefully acknowledged for their collaboration. The authors also acknowledge Dr R. Houben for revising this manuscript. We warmly thank Dr E. Baise, who recently passed away, for his collaboration on this topic since more than ten years and who has followed each step of the studies performed.

## Author Contributions

**Conceptualization:** François Boemer, Dominique-Marie Votion.

**Data curation:** Christophe Cello, Hélène Amory, Christel Marcillaud-Pitel, Eric Richard, Gaby van Galen, Gunther van Loon, Laurence Lefère.

**Formal analysis:** François Boemer, Dominique-Marie Votion.

**Methodology:** Johann Detilleux.

**Software:** Johann Detilleux.

**Supervision:** François Boemer, Dominique-Marie Votion.

**Validation:** François Boemer.

**Writing – original draft:** Dominique-Marie Votion.

**Writing – review & editing:** François Boemer.

## References

1. Baise E, Habyarimana JA, Amory H, Boemer F, Douny C, Gustin P, et al. Samaras and seedlings of *Acer pseudoplatanus* are potential sources of hypoglycin A intoxication in atypical myopathy without necessarily inducing clinical signs. *Equine Veterinary Journal*. 2015. <https://doi.org/10.1111/evj.12499> PMID: 26278545
2. Votion DM, van Galen G, Sweetman L, Boemer F, de Tullio P, Dopagne C, et al. Identification of methylenecyclopropyl acetic acid in serum of European horses with atypical myopathy. *Equine Vet J*. 2014; 46(2):146–9. <https://doi.org/10.1111/evj.12117> PMID: 23773055.
3. Westermann CM, van Leeuwen R, van Raamsdonk LW, Mol HG. Hypoglycin A Concentrations in Maple Tree Species in the Netherlands and the Occurrence of Atypical Myopathy in Horses. *J Vet Intern Med*. 2016; 30(3):880–4. <https://doi.org/10.1111/jvim.13927> PMID: 26995161.
4. Westermann CM, Dorland L, Votion DM, de Sain-van der Velden MG, Wijnberg ID, Wanders RJ, et al. Acquired multiple Acyl-CoA dehydrogenase deficiency in 10 horses with atypical myopathy. *Neuromuscul Disord*. 2008; 18(5):355–64. Epub 2008/04/15. S0960-8966(08)00037-0 [pii] <https://doi.org/10.1016/j.nmd.2008.02.007> PMID: 18406615.
5. Bochnia M, Ziegler J, Sander J, Uhlig A, Schaefer S, Vollstedt S, et al. Hypoglycin A content in blood and urine discriminates horses with atypical myopathy from clinically normal horses grazing on the same pasture. *PLoS One*. 2015. <https://doi.org/10.1371/journal.pone.0136785> PMID: 26378918
6. Sander J, Cavalleri JM, Terhardt M, Bochnia M, Zeyner A, Zuraw A, et al. Rapid diagnosis of hypoglycin A intoxication in atypical myopathy of horses. *J Vet Diagn Invest*. 2016; 28(2):98–104. <https://doi.org/10.1177/1040638715624736> PMID: 26965229.
7. Boemer F, Deberg M, Schoos R, Baise E, Amory H, Gault G, et al. Quantification of hypoglycin A in plasma using a TRAQ(R) kit. *Journal of Chromatography B*. 2015; 997:75–80.
8. van Galen G, Marcillaud Pitel C, Saegerman C, Patarin F, Amory H, Baily JD, et al. European outbreaks of atypical myopathy in grazing equids (2006–2009): Spatiotemporal distribution, history and clinical features. *Equine Vet J*. 2012; 44(5):614–20. Epub 2012/03/28. <https://doi.org/10.1111/j.2042-3306.2012.00556.x> PMID: 22448904.
9. van Galen G, Votion DM. Management of cases suffering from atypical myopathy: Interpretations of descriptive, epidemiological and pathophysiological findings. Part 1: First aid, cardiovascular, nutritional and digestive care. *Equine Veterinary Education*. 2013; 25(5):264–70.
10. van Galen G, Votion DM. Management of cases suffering from atypical myopathy: Interpretations of descriptive, epidemiological and pathophysiological findings. Part 2: Muscular, urinary, respiratory and hepatic care, and inflammatory/infectious status. *Equine Veterinary Education*. 2013; 25(6):264–70.
11. van Galen G, Saegerman C, Marcillaud Pitel C, Patarin F, Amory H, Baily JD, et al. European outbreaks of atypical myopathy in grazing horses (2006–2009): Determination of indicators for risk and prognostic factors. *Equine Vet J*. 2012; 44(5):621–5. Epub 2012/03/15. <https://doi.org/10.1111/j.2042-3306.2012.00555.x> PMID: 22413891.
12. Gonzalez-Medina S, Ireland JL, Piercy RJ, Newton JR, Votion DM. Equine Atypical Myopathy in the UK: Epidemiological characteristics of cases reported from 2011 to 2015 and factors associated with survival. *Equine Vet J*. 2017. <https://doi.org/10.1111/evj.12694> PMID: 28445006.



13. Carlier J, Guitton J, Moreau C, Boyer B, Bévalot F, Fanton L, et al. A validated method for quantifying hypoglycin A in whole blood by UHPLC-HRMS/MS. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2015; 978–979:70–7. <https://doi.org/10.1016/j.jchromb.2014.11.029> PMID: 25531872
14. Boemer F, Deberg M, Schoos R, Baise E, Amory H, Gault G, et al. Quantification of hypoglycin A in serum using aTRAQ(R) assay. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2015; 997:75–80. <https://doi.org/10.1016/j.jchromb.2015.06.004> PMID: 26094208.
15. Chace DH, Pons R, Chiriboga CA, McMahon DJ, Tein I, Naylor EW, et al. Neonatal blood carnitine concentrations: normative data by electrospray tandem mass spectrometry. *Pediatr Res*. 2003; 53(5):823–9. Epub 2003/03/04. <https://doi.org/10.1203/01.PDR.0000059220.39578.3D> 01. PDR.0000059220.39578.3D [pii]. PMID: 12612202.
16. Filee R, Schoos R, Boemer F. Evaluation of physiological amino acids profiling by tandem mass spectrometry. *J Inherit Metab Dis*. 2014; 13:119–28.
17. Detilleux J, Arendt J, Lomba F, Leroy P. Methods for estimating areas under receiver-operating characteristic curves: illustration with somatic-cell scores in subclinical intramammary infections. *Prev Vet Med*. 1999; 41(2–3):75–88. PMID: 10448938.
18. Esbjørn Høffer S, Votion DM, Andeberg M, Boemer F, Nautrup Olsen S, Van Galen G. Atypical Myopathy in Denmark Confirmed With the aTRAQ Assay. *Journal of Equine Veterinary Science*. 2016;(47):77–9.
19. McKenzie RK, Hill FI, Habyarimana JA, Boemer F, Votion DM. Detection of hypoglycin A in the seeds of sycamore (*Acer pseudoplatanus*) and box elder (*A. negundo*) in New Zealand; the toxin associated with cases of equine atypical myopathy. *N Z Vet J*. 2016; 64(3):182–7. <https://doi.org/10.1080/00480169.2015.1123123> PMID: 26593855.
20. Votion DM, Linden A, Delguste C, Amory H, Thiry E, Engels P, et al. Atypical myopathy in grazing horses: a first exploratory data analysis. *The Veterinary Journal*. 2009; 180(1):77–87. Epub 2008/04/09. S1090-0233(08)00037-3 [pii] <https://doi.org/10.1016/j.tvjl.2008.01.016> PMID: 18396430.
21. Votion DM, Linden A, Saegerman C, Engels P, Ercicum M, Thiry E, et al. History and clinical features of atypical myopathy in horses in Belgium (2000–2005). *Journal of Veterinary Internal Medicine*. 2007; 21(6):1380–91. Epub 2008/01/17. PMID: 18196750.
22. van Galen G, Cerri S, Porter S, Saegerman C, Lefere L, Roscher K, et al. Traditional and quantitative assessment of Acid-base and shock variables in horses with atypical myopathy. *J Vet Intern Med*. 2013; 27(1):186–93. Epub 2012/12/01. <https://doi.org/10.1111/jvim.12003> PMID: 23193982.
23. Verheyen T, Decloedt A, De Clercq D, van Loon G. Cardiac changes in horses with atypical myopathy. *J Vet Intern Med*. 2012; 26(4):1019–26. Epub 2012/06/01. <https://doi.org/10.1111/j.1939-1676.2012.00945.x> PMID: 22646196.
24. Von Holt C. Methylene cyclopropaneacetic acid, a metabolite of hypoglycin. *Biochim Biophys Acta*. 1966; 125(1):1–10. Epub 1966/08/03. 0005-2760(66)90138-X [pii]. PMID: 5968592.
25. Tanaka K, Kean EA, Johnson B. Jamaican vomiting sickness. Biochemical investigation of two cases. *N Engl J Med*. 1976; 295(9):461–7. Epub 1976/08/26. <https://doi.org/10.1056/NEJM197608262950901> PMID: 940578.
26. Barceloux DG. Akee fruit and Jamaican vomiting sickness (*Blighia sapida* Koenig). *Dis Mon*. 2009; 55(6):318–26. Epub 2009/05/19. S0011-5029(09)00027-3 [pii] <https://doi.org/10.1016/j.disamonth.2009.03.002> PMID: 19446675.
27. Shih VE, Tanaka K. Plasma and urine amino acid changes in rats treated with hypoglycin. *Clin Chim Acta*. 1978; 88(3):539–43. Epub 1978/09/15. PMID: 699343.
28. Delguste C, Cassart D, Baise E, Linden A, Schwarzwald C, Feige K, et al. Myopathies atypiques chez les chevaux au pré: une série de cas en Belgique. *Ann Med Vet*. 2002; 146:231–43.
29. Grunes DE, Scordi-Bello I, Suh M, Florman S, Yao J, Fiel MI, et al. Fulminant hepatic failure attributed to Akee fruit ingestion in a patient with sickle cell trait. *Case Reports in Transplantation*. 2012;(Article ID 739238):4 pages. <https://doi.org/10.1155/2012/739238> PMID: 23259140
30. Osmundsen H, Sherratt HS. A novel mechanism for inhibition of beta-oxidation by methylenecyclopropylacetyl-CoA, a metabolite of hypoglycin. *FEBS Lett*. 1975; 55(1):38–41. Epub 1975/07/15. PMID: 1140426.
31. Tanaka K, Miller EM, Isselbacher KJ. Hypoglycin A: a specific inhibitor of isovaleryl CoA dehydrogenase. *Proc Natl Acad Sci U S A*. 1971; 68(1):20–4. Epub 1971/01/01. PMID: 5276292; PubMed Central PMCID: PMC391092.
32. Tanaka K. On the mode of action of hypoglycin A. 3. Isolation and identification of cis-4-decene-1,10-dioic, cis, cis-4,7-decadiene-1,10-dioic, cis-4-octene-1,8-dioic, glutaric, and adipic acids, N-(methylenecyclopropyl)acetyl glycine, and N-isovalerylglycine from urine of hypoglycin A-treated rats. *J Biol Chem*. 1972; 247(23):7465–78. PMID: 4636318.

33. Ikeda Y, Tanaka K. Selective inactivation of various acyl-CoA dehydrogenases by (methylenecyclopropyl)acetyl-CoA. *Biochim Biophys Acta*. 1990; 1038(2):216–21. Epub 1990/04/19. 0167-4838(90)90208-W [pii]. PMID: [2331485](#).
34. Lai MT, Li D, Oh E, Liu HW. Inactivation of medium-chain acyl-CoA dehydrogenase by a metabolite of hypoglycin: characterization of the major turnover product and evidence suggesting an alternative flavin modification pathway. *J Am Chem Soc*. 1993; 115(5):1619–28.
35. Entman M, Bressler R. The mechanism of action of hypoglycin on long-chain fatty acid oxidation. *Mol Pharmacol*. 1967; 3(4):333–40. Epub 1967/07/01. PMID: [6033633](#).